Genotype-Dependent Sensitivity of Uveal Melanoma Cell Lines to Inhibition of B-Raf, MEK, and Akt Kinases: Rationale for Personalized Therapy

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PURPOSE. Inhibitors of B-Raf and MEK kinases hold promise for the management of cutaneous melanomas harboring BRAF mutations. BRAF mutations are rare in uveal melanomas (UMs), but somatic mutations in the G protein α subunits Gq and Gq11 (encoded by GNAQ and GNA11, respectively) occur in a mutually exclusive pattern in ~80% of UM. The impact of B-Raf and MEK inhibitors on Gq-mutant UM remains unknown.

METHODS. The impact of the B-Raf inhibitor PLX4720, the MEK inhibitor AZD6244, and the Akt inhibitor MK2206 on UM cell lines was assessed with the use of cell viability, proliferation, and apoptosis assays and immunoblot analysis.

RESULTS. BRAF-mutant UM cells were sensitive to both PLX4720 and AZD6244, undergoing cell cycle arrest but not apoptosis. UM cells with a G protein–mutant (GNAQ or GNA11) were mildly sensitive to AZD6244 but completely resistant to PLX4720. In fact, PLX4720 paradoxically increased ERK phosphorylation in Gq-mutant UM cells. The combination of AZD6244 and PLX4720 had synergistic antitumor activity in BRAF-mutant cells but not in Gq-mutant cells. The Akt inhibitor MK2206 sensitized BRAF-mutant cells to both PLX4720 and AZD6244 and sensitized Gq-mutant cells to AZD6244 but did not overcome the resistance of the Gq-mutant cells to PLX4720.

CONCLUSIONS. The response of UM cells to inhibition of B-Raf, MEK, and Akt depends on their genotype. Future use of such targeted therapies in clinical trials of UM patients will require careful design and patient selection based on genotype to provide personalized and effective therapy. (Invest Ophthalmol Vis Sci. 2011;52:7248–7255) DOI:10.1167/iovs.11-7398

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Supported by a fellowship from Jackstaedt Stiftung, Essen (AIR) and by National Institutes of Health/National Institute of Diabetes and Digestive and Kidney Diseases Grant 5K01DK081446 (BH).

Submitted for publication February 15, 2011; revised July 10, 2011; accepted July 23, 2011.

Disclosure: N. Mitsiades, None; S.A. Chew, None; B. He, None; A.I. Riechardt, None; T. Karadedou, None; V. Kotoula, None; V. Poulaki, None

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Ras-like domain, abolishing their GTPase activity in a manner similar to that for the NRASQ61R mutation, and resulting in a constitutively active Gα protein that functions as a bona fide oncogene. A second hot spot for mutations has been discovered in exon 4, affecting codon R113 (for both proteins). The presence of these mutations in tumors at all stages of malignant progression suggests that they are early events in UM. However, the sensitivity of Gα-mutant UM cells to the B-Raf and MEK inhibitors currently undergoing clinical development for the management of CM remains unknown.

We investigated the impact of B-Raf and MEK inhibition on UM cell lines using the small molecule inhibitors PLX4720 and AZD6244, respectively, either as monotherapy or in combination with each other and the Akt inhibitor MK2206 (Merck, North Wales, PA). We found that the BRAF-mutant UM cells behave similarly to their cutaneous counterparts, with high sensitivity to inhibition of either B-Raf or MEK that can be further enhanced by concurrent Akt inhibition. However, Gα-mutant UM cells are less sensitive to MEK inhibition (but can be further sensitized by concurrent Akt inhibition) and completely resistant to B-Raf inhibition (even in the presence of the Akt inhibitor). In fact, the B-Raf inhibitor PLX4720 paradoxically increased ERK phosphorylation in Gα-mutant UM cells. Our data demonstrate that the response of UM cells to the inhibition of B-Raf and MEK is genotype dependent. Future use of targeted therapies in clinical trials of UM patients will require careful design and patient selection based on genotype to provide personalized and effective therapy.

**Materials and Methods**

**Cell Lines and Tissue Culture**

The genotype of OMM1.3 and Mel202 UM cells (both GNAQ-mutant at Q209 and BRAF wt) has been previously reported. The OMM1 UM cells carry a GNA11 Q209 mutation and are wild-type for BRAF and GNAQ. The OCM3 (BRAFV600E/GNAQ wt/GNA11 wt) UM cell line and the A375 and M14 (both BRAFV600E/GNAQ wt/GNA11 wt) CM cell lines were also used. All cell lines examined in this study were carefully genotyped by Sanger sequencing using a cycle sequencing kit (BigDye Terminator v1.1; Applied Biosystems, Foster City, CA) and a genetic analyzer (3130; Applied Biosystems) for GNAQ exons 4 and 5, and GNA11 exons 4 and 5, and BRAF exon 15, and their mutation status was confirmed (Supplementary Fig. S1, http://www.iovs.org/lookup/suppl/doi:10.1167/iovs.11-7398/-/DCSupplemental). Overall, BRAF, GNAQ, and GNA11 mutations are mutually exclusive. All cells were grown in DMEM/F12 (Invitrogen, Carlsbad, CA) with 100 μM penicillin, 100 μg/ml streptomycin, and 10% FBS (Invitrogen).

**MTT, Proliferation, and Apoptosis Assays**

The B-Raf inhibitor PLX4720, the MEK inhibitor AZD6244, and the Akt inhibitor MK2206 were reconstituted in dimethyl sulfoxide (DMSO) and were stored at −20°C until use. Cells were plated in 24-well plates in medium containing 10% FBS. Drugs were added 24 hours later, and the cells were incubated for 96 more hours. Cell numbers were determined in medium containing 10% FBS. Drugs were added 24 hours later, and the cells were incubated for 96 more hours. Cell numbers were determined by BrdU incorporation into DNA (BrdU Cell Proliferation Assay; Calbiochem, San Diego, CA) according to the instructions of the manufacturer and as previously performed.

Apoptosis was detected in UM cells treated with AZD6244 or PLX4720 for 72 hours using the terminal deoxynucleotidyl transferase dUTP nick-end labeling (TUNEL) method with a cell death detection kit (In Situ Cell Death Kit, Fluorescein; Roche Applied Science, Indianapolis, IN), in accordance with the instructions of the manufacturer, and were run on a flow cytometer (BD FACSCanto II; BD Biosciences, San Jose, CA). Flow cytometry software (BD FACSdiva; BD Biosciences) was used to analyze the data.

A positron emission tomography (PET)/CT scan was performed with 18F-fluorodeoxyglucose (FDG) PET/CT at the University of California, San Diego, and the presence of FDG uptake was assessed. The presence of FDG uptake was considered indicative of the presence of cancer cells.

**Statistical Analysis**

To evaluate the differences across various experimental conditions in the viability experiments, one-way analysis of variance was performed, and post hoc tests (Duncan and Dunnett’s T3 tests) served to evaluate differences between pairs of experimental conditions (e.g., vehicle-treated cells vs. cells treated with each concentration of each agent). The additive or synergistic nature of the interaction between agents used in combination was evaluated by isobologram analysis using dose-effect analyzer software (Calcusyn; Biosoft, Ferguson, MO). In all analyses, P < 0.05 was considered statistically significant.

**Results**

**Impact of MEK Inhibition on UM Cell Lines Depends on Their Genotype**

We tested the impact of the MEK inhibitor AZD6244 on UM cell lines using MTT assay and immunoblotting with the BRAF-mutant CM cell lines A375 and M14 serving as controls. We found that the BRAF-mutant OCM3 cells were very sensitive to AZD6244, similar to their CM counterparts (Fig. 1A). The Ga-mutant UM cell lines were less sensitive (higher IC50) than the BRAF-mutant cells but still responded with growth arrest (Fig. 1A).

Immunoblot analysis revealed that AZD6244 decreased levels of phosphorylated ERK, decreased levels of cyclin D1, increased levels of the cyclin-dependent kinase inhibitor p27, and decreased levels of phosphorylated Rb in both BRAF-mutant (OCM3) and Ga-mutant (OMM1.3) UM cell lines (Fig. 1B). All these changes are consistent with and probably mediate the growth-suppressive effect of AZD6244, as has been previously reported for BRAF-mutant CM cell lines.

**Impact of B-Raf Inhibition on UM Cell Lines Depends on Their Genotype**

We tested the impact of the B-Raf inhibitor PLX4720 on UM cell lines using cell survival assays and immunoblot analysis with the BRAF-mutant CM cell lines A375 and M14 serving as controls. We found that the BRAF-mutant OCM3 cells were sensitive to PLX4720, similar to their CM counterparts (Fig. 2A). However, the Ga-mutant UM cell lines were resistant to the anticancer effect of PLX4720, and, in fact, OMM1.3 cells even exhibited a paradoxical mild increase in proliferation (Fig. 2A).

Stimulatory effect of PLX4720 occurred both in the presence of 10% FBS and in serum-free conditions.

Immunoblot analysis revealed that PLX4720 decreased levels of phosphorylated ERK, decreased levels of cyclin D1, increased levels of p27, and decreased levels of phosphory-
lated Rb only in BRAF-mutant (OCM3) cells. On the contrary, in Ga-mutant (OMM1.3) UM cells, PLX4720 induced a paradoxical increase in phosphorylated ERK and an early increase in cyclin D1 and pRb levels and did not stimulate p27 levels (Fig. 2B).

Collectively, our findings suggest that the B-Raf inhibitor PLX4720 does not exert anticancer activity against Ga/H9251-mutant melanoma cell lines (in fact, it can stimulate ERK signaling).

**BrdU Incorporation**

Using the BrdU cell proliferation assay, we studied the impact of AZD6244 and PLX4720 on UM DNA synthesis (S phase). Overall, we obtained parallel results to our MTT data. We found that the BRAF-mutant OCM3 cells were very sensitive to AZD6244, whereas OMM1.3 cells still responded with decreased DNA synthesis (but to a lesser degree; Fig. 3A). We also found that OCM3 cells were sensitive to PLX4720. However, the Ga-mutant OMM1.3 cells were resistant to the anticancer effect of PLX4720 (Fig. 3A).

**TUNEL Assay**

UM cells treated with AZD6244 or PLX4720 for 72 hours were tested for apoptosis using the terminal dUTP nick-end labeling (TUNEL) assay. We did not detect any increase in TUNEL labeling in OCM3 or OMM1.3 cells treated with AZD6244 and PLX4720, suggesting lack of apoptosis (Fig. 3B). The proteasome inhibitor bortezomib (100 nM for 72 hours) served as a positive control for the induction of apoptosis.

Collectively, these data suggest that the mechanism of the observed activity is by growth arrest, not apoptosis. In support, we did not detect any cleavage of caspase-3 or PARP in our immunoblotting analysis of UM cells treated with AZD6244 or PLX4720 for 72 hours (not shown), and the anticancer activity of AZD6244 and PLX4720 was not attenuated by pretreatment with the pan-caspase inhibitor ZVAD-FMK, indicating that apoptosis is not substantially involved in their effects (Supplementary Fig. S2, http://www.iovs.org/lookup/suppl/doi:10.1167/iovs.11-7398/-/DCSupplemental).

**Impact of Combined B-Raf and MEK Inhibition on BRAF-Mutant and Ga-Mutant UM Cells**

We next assessed whether combined B-Raf and MEK inhibition would achieve an enhanced anticancer activity. We found that, in BRAF-mutant OCM3 cells, the combination of sublethal concentrations of PLX4720 and AZD6244 resulted in synergistic anticancer activity (Fig. 4A). However, in Ga-mutant
OMM1.3 cells, the addition of AZD6244 could not overcome their resistance to PLX4720 (Fig. 4B). Therefore, the combination of B-Raf and MEK inhibition appears to be a promising approach only for BRAF-mutant cells.

Impact of B-Raf and MEK Inhibition on the Akt Pathway in BRAF-Mutant and G\(\alpha\)-Mutant UM Cells

We next investigated the impact of B-Raf and MEK inhibition on the Akt pathway in BRAF\(^{V600E}\) and G\(\alpha\)-mutant UM cells. We found that both AZD6244 and PLX4720 induced an early (within 2–6 hours of exposure) increase in the levels of phosphorylated Akt (both Ser473 and Thr308) in both BRAF-mutant (OCM3) and G\(\alpha\)-mutant (OMM1.3) cells. On the contrary, in G\(\alpha\)-mutant (OMM1.3) UM cells, PLX4720 induced a paradoxical increase in phosphorylated ERK and Rb levels and an early increase in cyclin D1 levels but did not stimulate p27 levels.

We hypothesized that the early upregulation of pAkt caused by B-Raf or MEK inhibition might attenuate the anticancer activity of AZD6244, PLX4720, or both, and, therefore, concurrent inhibition of the Akt pathway might enhance the anticancer activity of B-Raf and MEK inhibition. To address this hypothesis, we evaluated the effect of the Akt inhibitor MK2206 in combination with AZD6244 or PLX4720. We found that the Akt inhibitor MK2206 potently enhanced the anticancer activity of the MEK inhibitor (Fig. 5B) and the B-Raf inhibitor (not shown) against the BRAF-mutant OCM3 cells. MK2206 also enhanced the anticancer activity of the MEK inhibitor against the G\(\alpha\)-mutant UM cells but did not overcome the resistance of the G\(\alpha\)-mutant UM cells to the B-Raf inhibitor (Figs. 6A, 6B).

DISCUSSION

Recent advances in our understanding of the molecular pathophysiology of cancer have allowed for rational development of targeted therapies designed to interrupt molecular pathways critical for cell growth and survival. Kinases represent such “druggable” targets and an area of very active clinical research in oncology. Various malignancies that exhibit “oncogenic addiction” to select kinase pathways respond clinically to treatment with respective kinase inhibitors (e.g., imatinib in chronic myelogenous leukemia and gastrointestinal stromal tumors, erlotinib in EGFR-mutant non–small cell lung carcinoma, and others). Because of the biological heterogeneity and interindividual variation in human cancers, molecular profiling of each patient’s tumor is necessary to guide selection of the appropriate targeted therapy for the patient most likely to benefit from it. The goal of this personalized approach is to avoid exposing patients to drugs from which they are unlikely to benefit, thus sparing them unnecessary toxicity and cost. For example, responses of non–small cell lung carcino-
mas to EGFR small molecule inhibitors (such as erlotinib) are generally limited to tumors harboring somatic mutations in the EGFR tyrosine kinase domain. Moreover, colorectal carcinomas harboring certain activating somatic mutations in KRAS are resistant to EGFR-targeting monoclonal antibodies, and use of such agents should be avoided in these particular carcinomas. In the present study, we investigated the activity of small molecule inhibitors of MEK, B-Raf and Akt against UM cells in vitro. Our results demonstrate that the sensitivity of UM cells to these inhibitors is genotype dependent and make a strong case for a personalized approach in the management of UM with targeted therapies.

CMs frequently harbor mutations in BRAF (usually BRAF\(^{V600E}\)) or NRAS\(^{4-6}\) that cause constitutive activation of

**Figure 3.** Growth arrest without apoptosis in UM cell lines treated with AZD6244 or PLX4720 monotherapy. (A) Using the BrdU cell proliferation assay, we studied the impact of AZD6244 and PLX4720 on UM DNA synthesis (S phase). The BRAF-mutant OCM3 cells and the Ga-mutant OMM1.3 cells were treated with AZD6244 or PLX4720 (or DMSO) at the indicated concentrations in 24-well plates for 48 hours. Subsequently, BrdU was added to the wells for an additional 24 hours, and the amount of incorporation into DNA was quantified according to the instructions of the manufacturer and was expressed as percentage of control (DMSO) wells (average ± SD). We found that the BRAF-mutant OCM3 cells were sensitive to AZD6244 and PLX4720, whereas Ga-mutant OMM1.3 cells still responded with decreased DNA synthesis to AZD6244 (but to a lesser degree than OCM3 cells) though they were resistant to the anticancer effect of PLX4720 (\(P < 0.05\), \(P < 0.001\) vs. respective DMSO controls). (B) Using the TUNEL assay, we studied the impact of AZD6244 and PLX4720 on UM cell apoptosis. UM cells were treated with AZD6244 or PLX4720 at 3 \(\mu\)M (or DMSO control) for 72 hours and then labeled for apoptosis using the enzyme terminal deoxynucleotidyl transferase (TdT) and the fluorescein-dUTP. We did not detect any shift in fluorescence (TUNEL labeling) in OCM3 or OMM1.3 cells treated with AZD6244 or PLX4720 monotherapy, suggesting a lack of apoptosis. The proteasome inhibitor bortezomib (100 nM for 72 hours) served as a positive control for the induction of apoptosis. Cells incubated without TdT or fluorescein-dUTP served as a negative control.

**Figure 4.** Impact of combined B-Raf and MEK inhibition on BRAF-mutant and Ga-mutant UM cells. The BRAF-mutant OCM3 cells (A) and the Ga-mutant OMM1.3 (B) and Mel202 cells were treated with the combination of PLX4720 and AZD6244 at the indicated concentrations for 96 hours in medium containing 10% FBS. Cell number was quantified with the MTT assay and expressed as a percentage of control wells (average ± SD). In BRAF-mutant OCM3 cells, the combination of sublethal concentrations of PLX4720 and AZD6244 resulted in synergistic anticancer activity (A), whereas in Ga-mutant OMM1.3 cells (B) and Mel202 cells (not shown), the addition of AZD6244 could not overcome their resistance to PLX4720.
the MEK/ERK pathway and cell proliferation. CMs harboring BRAFV600E mutations exhibit increased sensitivity to inhibitors of MEK and B-Raf. We found similar sensitivity to the MEK and B-Raf inhibitors in BRAFV600E UM cells, suggesting that such targeted therapies may represent a promising option for this subset of UM patients as well. In agreement with their CM counterparts, BRAFV600E UM cells treated with AZD6244 or PLX4720 exhibited decreased levels of phosphorylated ERK, decreased levels of cyclin D1, increased levels of the cyclin-dependent kinase inhibitor p27, and decreased levels of phosphorylated Rb. All these changes are consistent with and probably mediate the growth-suppressive effect (inhibition of cell proliferation) of MEK or B-Raf inhibition.

However, BRAFV600E mutations are rare in UMs. Instead, an equivalent oncogenic event is frequently present in the form of mutually exclusive somatic mutations in the heterotrimeric G protein α-subunit GNAQ (~40%-50% of primary UMs) or its related GNA11 (~30% of primary UMs). These mutations apparently occur early in UM carcinogenesis, contrary to other genetic events associated with increased metastatic potential such as the recently described BRCA1-associated protein 1 (BAP1) mutations. The GNAQQ209 and GNA11Q209 mutations affect the Ras-like domain of these G proteins, specifically corresponding to NRASQ61R, abolishing GTPase activity, and resulting in constitutive activation. In this study, we examined the sensitivity of GNAQ-mutant and GNA11-mutant UM cell lines to MEK and B-Raf inhibition. We found that Gα-mutant UM and CM cells are sensitive to MEK inhibition (exhibiting decreased levels of pERK, cyclin D1, and pRb, increased levels of p27, and decreased proliferation) but to a lower degree (higher IC50) than BRAF-mutant UM and CM cells. In direct contrast to BRAF-mutant

![Figure 5](http://doi.org/10.1167/iovs.11-7253) Impact of B-Raf and MEK inhibition on the Akt pathway in BRAF-mutant and Gα-mutant UM cells. (A) Immunoblot analysis revealed that both AZD6244 and PLX4720 induced an early (within 2–6 hours of exposure) increase in the levels of phosphorylated Akt (at residues Ser473 and Thr308) in both BRAF-mutant OCM3 and Gα-mutant OMM1.3 cells. Eventually, and with prolonged drug exposure, the phosphorylation of Akt returned to baseline in Gα-mutant OMM1.3 cells and decreased even below baseline levels in BRAF-mutant OCM3 cells. (B) The Akt inhibitor MK2206 potently enhanced the anticancer activity of the MEK inhibitor AZD6244 against the BRAF-mutant OCM3 cells. The cells were treated with drugs at the indicated concentrations for 96 hours in medium containing 10% FBS. Cell number was quantified with the MTT assay and expressed as a percentage of control wells (average ± SD).
In view of the resistance of Go-mutant UM cells to PLX4720, we investigated whether combination of the B-Raf inhibitor with MEK or Akt inhibition would restore sensitivity. Unfortunately, in Go-mutant UM cells, the addition of the MEK inhibitor AZD6244 or the Akt inhibitor MK2206 did not restore sensitivity to PLX4720. On the contrary, in B-Raf-mutant cells, combined B-Raf and MEK inhibition resulted in synergistic anticancer activity. Furthermore, the Akt inhibitor MK2206 enhanced the anticancer activity of both the B-Raf inhibitor and the MEK inhibitor against the B-Raf-mutant UM cells and the anticancer activity of the MEK inhibitor against the Go-mutant UM cells. This finding is in agreement with the early upregulation of phosphorylated Akt that we observed on MEK or B-Raf inhibition in our models and with reports of the synergistic activity of AZD6244 in combination with the inhibition of Akt in B-RafV600E cutaneous melanoma and lung carcinoma cell lines. Moreover, in UM cells, cotreatment with the PI3K inhibitor LY294002, in combination with the MEK inhibitor U0126 or with the B-Raf inhibitor Bay45-4900, has been reported to result in more potent inhibition of cell proliferation. These data highlight the combined inhibition of MEK and Akt pathways as a rationally designed targeted therapeutic approach for Go-mutant UMs, that deserves investigation in a clinical trial.

In conclusion, we report that the Go mutations, present in the majority of UMs, are functionally similar to the NRAS mutations seen in CMs in that they are associated with lower sensitivity to MEK inhibition and complete resistance to the anticancer activity of the B-Raf inhibitor PLX4720. Combined inhibition of MEK and Akt results in synergistic anticancer activity and represents a promising targeted therapeutic approach for Go-mutant UMs. Our findings emphasize the importance of a personalized, genotype-based approach to the use of targeted therapies in clinical trials of patients with UM.

Acknowledgments

The authors thank Martine Jager (Department of Ophthalmology, Leiden University Medical Center, Leiden, The Netherlands) for generously providing UM cell lines.

References

enhances cell migration and proliferation of BRAF melanoma cells.


