

# Tumor-dependent Kinetics of Partial Pressure of Oxygen Fluctuations during Air and Oxygen Breathing

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## ABSTRACT

The primary purpose of this study was to examine the kinetics of partial pressure of oxygen (pO<sub>2</sub>) fluctuations in fibrosarcoma (FSA) and 9L tumors under air and O<sub>2</sub> breathing conditions. The overall hypothesis was that key factors relating to oxygen tension fluctuations would vary between the two tumor types and as a function of the oxygen content of the breathing gas. To assist in the interpretation of the temporal data, spatial pO<sub>2</sub> distributions were measured in 10 FSA and 8 9L tumors transplanted into the subcutis of the hind leg of Nembutal-anesthetized (50 mg/kg) Fischer 344 rats. Recessed-tip oxygen microelectrodes were inserted into the tumor, and linear pO<sub>2</sub> measurements were recorded in 50- $\mu$ m steps along a 3-mm path, and blood pressure was simultaneously measured via femoral arterial access. Additionally, pO<sub>2</sub> was measured at a single location for 90 to 120 minutes in FSA ( $n = 11$ ) or 9L tumors ( $n = 12$ ). Rats were switched from air to 100% O<sub>2</sub> breathing after 45 minutes. Temporal pO<sub>2</sub> records were evaluated for their potential radiobiological significance by assessing the number of times they crossed a 10-mm-Hg threshold. In addition, the data were subjected to Fourier analysis for air and O<sub>2</sub> breathing.

FSA and 9L tumors had spatial median pO<sub>2</sub> measurements of 4 and 1 mm Hg, respectively. 9L had more low pO<sub>2</sub> measurements  $\leq 2.5$  mm Hg than did FSA, whereas between 2.5 and 10 mm Hg this pattern was reversed. Pimonidazole staining patterns in FSA and 9L tumors supported these results. Temporal pO<sub>2</sub> instability was observed in all experiments during air and O<sub>2</sub> breathing. Threshold analyses indicated that the 10 mm Hg threshold was crossed 2 to 5 times per hour, independent of tumor type. However, the magnitude of 9L pO<sub>2</sub> fluctuations was approximately eight times greater than FSA fluctuations, as assessed with Fourier transform analysis (Wilcoxon,  $P < 0.005$ ). O<sub>2</sub> breathing significantly increased median pO<sub>2</sub> in FSA from 3 to 8 mm Hg ( $P < 0.005$ ) and caused a significant increase in frequency and magnitude of pO<sub>2</sub> fluctuations. One hundred percent O<sub>2</sub> breathing had no effect on 9L tumor pO<sub>2</sub>, and it decreased the magnitude of pO<sub>2</sub> fluctuations with borderline significance.

These results show that these two tumors differ significantly with respect to spatial and temporal oxygenation conditions under air and O<sub>2</sub> breathing. Fluctuations of pO<sub>2</sub> of the type reported herein are predicted to significantly affect radiotherapy response and could be a source for genetic instability, increased angiogenesis, and metastases.

## INTRODUCTION

Tumor hypoxia is classically depicted as developing as a result of two independent phenomena: chronic hypoxia caused by limitations of oxygen diffusion, and transient hypoxia caused by microvessel flow instabilities.

Although intermittent blood flow and hypoxia have been studied extensively with indirect methods, such as dye mismatch (1, 2), laser Doppler flowmetry (3), and hypoxia markers (4, 5), direct measure-

ment of partial pressure of oxygen (pO<sub>2</sub>) instability has been done only on the R3230Ac tumor with recessed tip microelectrodes (6, 7) and on two human tumor xenograft lines with a fiber optic oxygen sensor (8, 9). It is important to determine the characteristics of intermittent hypoxia in other tumor types with direct methods. Direct measurements of kinetics provide unique information about the frequency of subregions existing under radiobiologically significant pO<sub>2</sub> conditions. In addition, the magnitude of fluctuations can be determined. Both of these variables may be important in governing treatment response and in altering gene expression patterns that could indirectly influence tumor cellular behavior.

The main focus of this article is on the temporal instability in pO<sub>2</sub> during air and O<sub>2</sub> breathing. Spatial pO<sub>2</sub> distributions were also performed for these two tumors to facilitate interpretation of the transient data. Oxygen tension distributions of two tumor lines that grow in the Fischer 344 rat, a fibrosarcoma (FSA; ref. 10) and 9L glioma (9L; ref. 11), were performed, using methods identical to those used previously for the R3230Ac tumor. Second, the kinetics of pO<sub>2</sub> fluctuation were examined under air- and oxygen-breathing conditions.

## MATERIALS AND METHODS

### Animal Model

Forty-one female Fischer 344 rats (Charles River Laboratories, Raleigh, NC) were used in this study. Twenty-one rats received subcutaneous implants of 1- to 2-mm<sup>3</sup> pieces of a rat fibrosarcoma in the left hind limb. The remaining twenty received cell injections of four to six million 9L glioma cells grown in Eagle's basal medium supplemented with 10% fetal calf serum and 5% fibrinogen (cells obtained courtesy of K. Wheeler, Wake Forest University, Winston Salem, NC). After the tumors had reached 1 to 1.5 cm in diameter, rats were anesthetized with an intraperitoneal injection of 50 mg/kg pentobarbital sodium. The femoral artery and vein were cannulated for recording of blood pressure and venous access, respectively. A small portion (~4–10 mm<sup>2</sup>) of the skin and tumor capsule was removed to expose the tumor surface for microelectrode insertion. The surface was moistened by topical application of saline. A small incision was made in the left forelimb and a Ag/AgCl reference electrode was sutured into the subcutis. Body temperature was maintained at 37°C by placing the rat on a regulated water-heated blanket (K-module, Baxter Healthcare, Valencia, CA).

### Oxygen Microelectrodes

Recessed-tip microelectrodes were produced using a previously published technique (12, 13). The electrodes had tip diameters of 8.9 (6.1–12.7)  $\mu$ m (13 electrodes) and recess lengths of ~30  $\mu$ m. Microelectrodes were polarized at -0.7 V using a commercial polarizing box and a picoammeter unit (Chemical microsensor no. 1201, Diamond General, Ann Arbor, MI). Electrodes were calibrated before and after experiments in a 37°C saline-filled tonometer alternately bubbled with 0, 2.5, 5 or 15% O<sub>2</sub> (balance nitrogen). An *in vivo* zero value was obtained by recording microelectrode current in tissue after euthanasia of the rat with an overdose of pentobarbital sodium. The average sensitivity of the electrodes was 0.93 (0.73–1.5) mm Hg picoampere.

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## Spatial Profiles of Tumor Partial Pressure of Oxygen

These measurements were done using methods reported previously (14). Briefly, a micromanipulator (model MO102E, Narishige, Narishige, Japan) was positioned so that a dummy electrode could reach the exposed tumor surface, covered in a drop of saline. The actual electrode was then placed in the micromanipulator and advanced into the saline droplet. It was allowed to polarize and then was advanced into the tumor. Total measurement length for each track was 3 mm (50  $\mu\text{m}/\text{step}$ ) with 3 to 4 tracks per tumor (mean of 198 measurements per tumor). After experiments were completed, rats were sacrificed using Euthasol (Delmarva Laboratories, Midlothian, VA), and the *in vivo* zero value for the microelectrode was recorded. Cumulative frequency distributions were obtained for each animal and then were averaged over each tumor type to obtain means and 95% exact confidence intervals for specific pO<sub>2</sub> intervals in the histogram. Partial pressure of oxygen distributions were measured in 18 rats (10 FSA and 8 9L).

## Histology and Immunohistochemistry

**Pimonidazole.** Five rats, three FSA and two 9L, were given injections of 70 mg/kg pimonidazole before spatial profiles of pO<sub>2</sub> were completed. Three hours after injection and after the pO<sub>2</sub> profiles had been completed, the animals were sacrificed. The tumor was removed from the rat and was fixed in 70% formalin before being paraffin embedded within 24 to 48 h. Immunohistochemistry for pimonidazole (hypoxia marker) was carried out as described previously (15).

## Temporal Variations in Blood Flow and Tumor Partial Pressure of Oxygen

Anesthetized rats were placed on a temperature-controlled water-heated blanket, and the left leg was stabilized on a rubber pedestal with tape. The arterial cannula was connected to a blood pressure transducer and amplifier (model 11-G4143-01, Gould Instruments, Valley View, OH), and the amplifier signal was digitized at 25 Hz and recorded using data acquisition software (AT-CODAS, Windaq, DATAQ Instruments, Akron, OH). Two laser Doppler-flow probes (outer diameter, 480  $\mu\text{m}$ ) were inserted into the tumor on the side opposite from the exposed surface. The probes were connected to the flowmeter and data acquisition system (Oxford Array, Oxford Optronix, Oxford, United Kingdom), which acquires data at 20 Hz.

An oxygen microelectrode was inserted into the tumor, using the same method as described above. The microelectrode was moved several millimeters into the tissue until a clearly non-zero pO<sub>2</sub> value was obtained, allowing the possibility for the pO<sub>2</sub> to increase or decrease. The electrode then remained fixed for the rest of the study. Rats breathed air for 45 minutes and were then switched to 100% O<sub>2</sub> for an additional 45 minutes. During the full 90 minutes, pO<sub>2</sub>, blood pressure, and laser Doppler blood flow were continuously recorded. Twenty-three rats were used for this study (11 FSA and 12 9L).

## Fourier Analysis

Fourier analyses of temporal variation in blood flow and pO<sub>2</sub> were performed using commercial software included in the data acquisition package (CODAS, DATAQ Instruments, Akron, OH). Blood pressure and pO<sub>2</sub> recordings could be analyzed directly within the program, because the same software was used to acquire the data. The blood flow data were transferred from the Oxford array system to ASCII text files, which were then converted to CODAS files.

The frequency characteristics of the instrumentation had no impact on the Fourier analysis. The oxygen microelectrodes respond within 40 to 400 milliseconds (13), which is much faster than the fluctuations of interest in this study. Although the high-frequency response of the Chemical Microsensor no. 1201 is somewhat limited in the range (200 picoamperes full scale) that was used in most cases, its cutoff (3 decibels) frequency was 2.3 Hz (138 cycles/min). According to the manufacturer, the Oxford laser Doppler array has a cutoff frequency of 5 Hz (300 cycles/min). Because these cutoff frequencies are relatively high, the resulting attenuation had little or no effect on the apparent power within the frequency range of interest. Therefore, the instrumentation did not contribute to nor did it bias the pO<sub>2</sub> or laser Doppler flowmetry signals.

One record length was analyzed using the Fourier analysis: 33 minutes for

pO<sub>2</sub> and blood pressure; 34 minutes for blood flow. The analysis was performed on records that contained 49,146 points (blood pressure and pO<sub>2</sub>) or 40,960 points (blood flow). The software averaged the records over every 5 or 6 points to obtain files 8,192 points in length. For the recordings during air breathing, each record overlapped the previous record by 50%. For example, the first Fourier analysis of a pO<sub>2</sub> recording covered data from 0.0 to 32.8 minutes, the second from 12.0 to 44.9 minutes. The Fourier analysis of pO<sub>2</sub> during O<sub>2</sub> breathing was performed on the data recorded between 57.0 and 89.8 minutes.

## Analysis of Power Spectra

**Frequency.** An informative way to view and compare frequency information from Fourier power spectra is to examine the frequencies at which the power peaks (7, 16).

**Power and Magnitude of Fluctuations.** The power of the Fourier power spectra (*i.e.*, the magnitude of the fluctuations) was analyzed by examining the total power over a given frequency range (17). Because most of the power in the present study was at very low frequencies, the range chosen for this analysis was 0 to 10 cycles/min. The total power of a spectrum was obtained by a summation of all powers between 0 cycles/min and a given frequency.

## Statistical Analysis

**Partial Pressure of Oxygen Distribution Analysis.** The relative and cumulative frequencies of the spatial pO<sub>2</sub> data were plotted against the pO<sub>2</sub> and analyzed using SAS (SAS Institute, Cary, NC). The point estimate and its exact confidence interval were computed for each type of tumor for both relative and cumulative frequencies (averaged across tracks first, and then across experimental units for each type of tumor). Parametric (*t* test) as well as nonparametric (Wilcoxon and Normal score) tests were carried out for informal comparisons between the two tumor types at each histogram interval. A smoothing version of the histogram technique was explored using SAS procedure kernel density estimation to estimate the density and distribution of the data. The mixed effects model was also used to examine relative or cumulative frequency with respect to tumor type, using a random intercept to account for multiple pO<sub>2</sub> measurements from multiple tracks of one tumor. The results from the last two approaches, however, were very similar to those from the histogram methods and, therefore, are not discussed in detail.

**Fourier Analysis of pO<sub>2</sub> and Blood Flow Fluctuations.** The cumulative magnitude or power in millimeters of (mercury)<sup>2</sup> [(mm Hg)<sup>2</sup>] of pO<sub>2</sub> fluctuations and its 95% confidence interval were first found on the log scale to make its distribution more normal-like and avoid getting a negative lower confidence limit. Data were then transformed back to the original scale. The cumulative power of pO<sub>2</sub> was compared between tumor types for air and O<sub>2</sub> breathing, using the Wilcoxon rank test performed at each frequency point of interest. The difference between O<sub>2</sub> and air was compared within each tumor type using the signed rank test as well as between tumor types using Wilcoxon and normal score tests.

**Changes in pO<sub>2</sub> of Radiobiological Significance during Air and O<sub>2</sub> Breathing.** For hypoxia-reoxygenation kinetics, Wilcoxon and Van der Weerden normal score or  $\chi^2$  (for categorical variables) tests were used to assess for differences between tumor types in total duration and number of episodes, and median pO<sub>2</sub> during hypoxic episodes or the entire interval. A signed rank test was used to assess differences between O<sub>2</sub> and air breathing within each tumor type, and the difference between O<sub>2</sub> and air breathing across the two tumor types was assessed, as before, using Wilcoxon and Van der Weerden normal score tests.

Unless otherwise stated, all of the values presented in the text are medians with interquartile ranges (IQRs).

## RESULTS

### Spatial Partial Pressure of Oxygen Distributions

The median pO<sub>2</sub> values were 4 (2–8) mm Hg and 1 (0–7) mm Hg for FSA and 9L tumors, respectively. By comparison, the median value for the R3230Ac has been reported to be 4 mm Hg (14). FSA and 9L both have ~80% of their pO<sub>2</sub> values below 10 mm Hg, but between 0 and 10 mm Hg, there are differences between the tumor

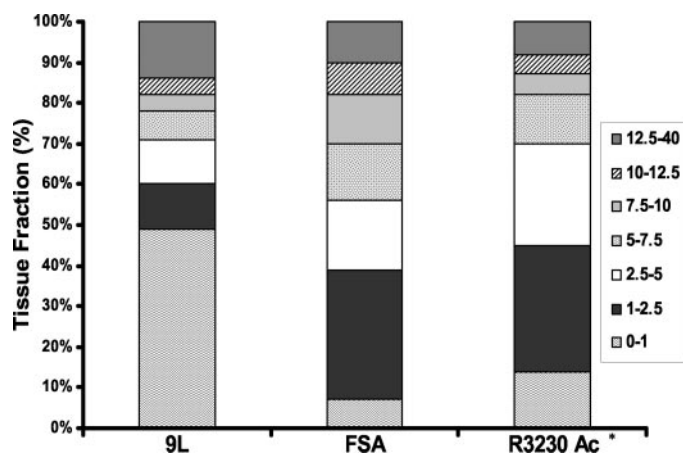


Fig. 1. FSA, 9L, and R3230Ac tumor lines all have 80 to 90% of the tissue fraction below 10 mm Hg; however, 9L has a much larger fraction of its tissue at values less than or equal to 1 mm Hg, and FSA and R3230Ac both have larger fractions (~50%) of moderately hypoxic regions between 1 and 5 mm Hg. See Table 1 for statistical comparisons. \*, from previously published data (1).

Table 1 Summary of partial pressure of oxygen histogram data

pO <sub>2</sub> range (in mm Hg)	Fraction of values*	
	9L	FSA
<1.0	49 (29–58)†	7 (2–13)†
<2.5	60 (48–71)†	39 (31–48)†
<5.0	71 (58–79)†	56 (46–64)†
<7.5	78 (66–84)	70 (58–77)†
<10.0	82 (70–87)	82 (72–87)
<12.5	86 (74–90)	90 (81–93)
<40.0	100 (94–100)	100 (94–100)

\* Percentages shown as means (95% confidence intervals).

† Significantly different from other tumor type.

types (Fig. 1; Table 1). 9L has a greater proportion of severely hypoxic pO<sub>2</sub> values (<2.5 mm Hg) than does FSA, which has the majority of its hypoxic values in an intermediate range (5–7.5 mm Hg). 9L had a significantly higher (71 versus 56%) cumulative percentage of pO<sub>2</sub> values ≤ 5 mm Hg compared with FSA ( $P < 0.05$ ).

### Histology and Immunohistochemistry

**Pimonidazole.** Examples from two tumors are shown in Fig. 2. FSA shows well-oxygenated regions near the edge of the tumor, but also has large regions of moderate hypoxia, shown as light brown staining throughout the tumor. The central region of the tumor also shows small patches of dark brown staining, indicating regions of severe hypoxia. Small regions of necrosis also appear along the bottom edge of the figure. 9L pimonidazole staining shows several dark brown patches indicating severe hypoxia very close to well-oxygenated regions. This tumor also shows many regions of necrosis, surrounded by regions of moderate to severe hypoxia.

### Intermittent Hypoxia

The time interval of the pO<sub>2</sub> measurements averaged 97.2 (92.3–104.0) minutes for FSA, and 93.3 (92.0–95.1) minutes for 9L. The median heart rate for all animals was 345 (321–371) beats per minute (bpm) for the minute previous to beginning O<sub>2</sub> breathing, and 342 (324–370) bpm for the last 33 minutes of O<sub>2</sub> breathing. Mean arterial blood pressure averaged 111 (104–116) and 116 (112–120) mm Hg for air and O<sub>2</sub> breathing, respectively. Thus, systemic cardiovascular conditions were stable during these experiments and within the range reported for unanesthetized rats (18).

### Air versus Oxygen Breathing Results

**Tumor pO<sub>2</sub> Fluctuations.** Tumor pO<sub>2</sub> fluctuations were measured in 11 FSA, and 12 9L tumors. An example of one FSA tumor recording is shown in Fig. 3A. In this example, pO<sub>2</sub> remained fairly constant during the air-breathing period, staying between 4–5 mm Hg. After O<sub>2</sub> breathing was started, pO<sub>2</sub> gradually increased to nearly 15 mm Hg. The magnitude of fluctuations changed with the switch from air to O<sub>2</sub>. During air breathing, fluctuations varied less than ~1 mm Hg. After the rat began breathing O<sub>2</sub>, the magnitude of fluctuations increased to 4 to 5 mm Hg (Fig. 3A). For FSA overall, the median pO<sub>2</sub> value during the 30 minutes before O<sub>2</sub> breathing was 3 (2–12) mm Hg, and it rose to 8 (4–25) mm Hg during the 30-minute period beginning 12 minutes after the onset of O<sub>2</sub> breathing. The increase in pO<sub>2</sub> was statistically significant ( $P < 0.005$ , signed rank test).

Fig. 3B shows one trace of a 9L tumor pO<sub>2</sub> recording. During the first 48 minutes of air breathing, pO<sub>2</sub> fluctuated and then gradually dropped from 16 to 2 mm Hg. The magnitude of fluctuations of pO<sub>2</sub> visibly decreased when the rat was switched to O<sub>2</sub> breathing (Fig. 3B). The median of pO<sub>2</sub> values during the last 30 minutes of air breathing was 9 (6–18) mm Hg, and was 9 (1–15) mm Hg before O<sub>2</sub> breathing. Thus, pO<sub>2</sub> did not significantly change in this tumor type on switching from air to O<sub>2</sub> breathing ( $P = 0.34$ ).

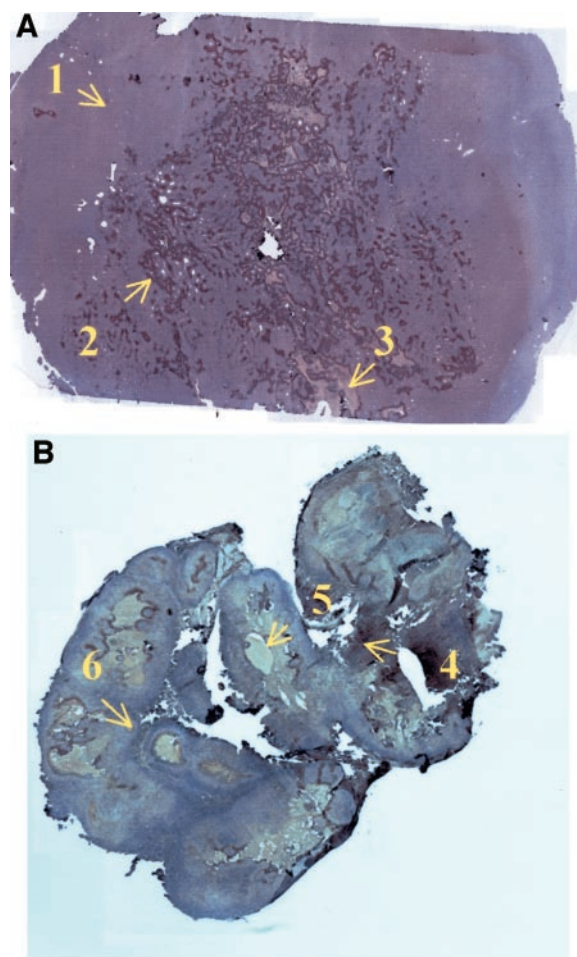


Fig. 2. Examples of pimonidazole staining in FSA (A) and 9L (B). FSA shows large regions of moderate hypoxia (arrow 1) and regions of well-oxygenated tissue. There are also small patches of severe hypoxia (arrow 2), and some necrosis (arrow 3). 9L pimonidazole staining shows large patches of severe hypoxia (arrow 4) and necrosis (arrow 5). There are also regions of severe hypoxia very close to well-oxygenated regions (arrow 6).

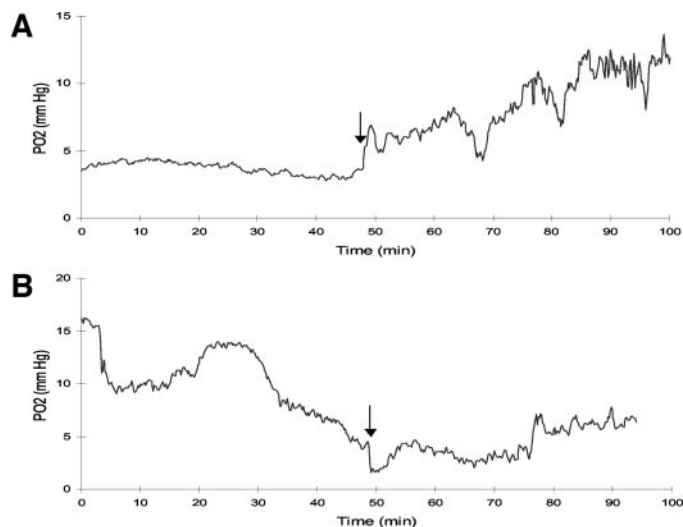


Fig. 3. Examples of temporal pO<sub>2</sub> traces for FSA (A) and 9L (B); arrow, the time at which the rat was switched from air to oxygen breathing. For FSA, pO<sub>2</sub> increased with the onset of O<sub>2</sub> breathing. The 9L tumor trace shows an immediate slight decrease in pO<sub>2</sub> when O<sub>2</sub> breathing began, but the level then returned to the baseline pO<sub>2</sub> a few minutes later.

### Changes in pO<sub>2</sub> of Radiobiological Significance during Air and O<sub>2</sub> Breathing.

FSA and 9L pO<sub>2</sub> traces during air and O<sub>2</sub> breathing were examined with respect to a 10-mm Hg threshold value for hypoxia. A threshold of 10 mm Hg was chosen because, below this value, radiosensitivity is most sensitive to changes in pO<sub>2</sub>. Any two hypoxic episodes separated by less than 30 seconds were collapsed into a single episode, under the condition that the difference in the median pO<sub>2</sub> between the two episodes was not greater than 5 mm Hg. After combining those episodes, if the duration of a hypoxic episode was no more than 10 seconds, it was treated as a nonhypoxic episode. These rules were set, keeping in mind that a typical radiation fraction usually requires several minutes to apply. Thus, very short periods of hypoxia would not likely affect radioresponse appreciably. Experiments for each tumor were classified as not showing hypoxia-reoxygenation if they remained either above or below the 10-mm-Hg threshold for the 30-minute observation during air breathing and the 30-minute observation during O<sub>2</sub> breathing (Table 2). For both tumor types, the number of times per hour that pO<sub>2</sub> crossed the 10-mm-Hg

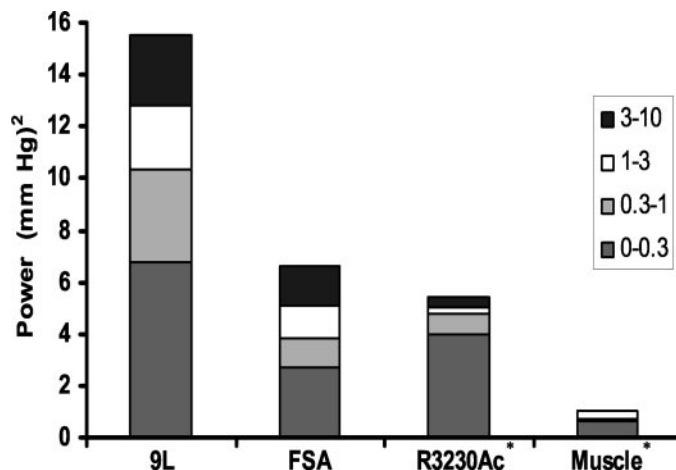


Fig. 4. Summary of pO<sub>2</sub> power versus fluctuation frequency for tumor lines and normal muscle. 9L has greater cumulative power than FSA, R3230Ac, and muscle tissue over all frequencies shown. FSA and 9L have a large portion of their cumulative power occurring at higher frequencies (0.3–10 cpm), whereas most of the cumulative power for R3230Ac occurs between 0 and 0.3 cpm. FSA cumulative power is roughly equivalent to that of the R3230Ac, both of which were higher in cumulative power than muscle tissue over all frequencies shown. \*, from previously published data (9).

threshold ranged from 0 to 10 was an average of 2. The frequency of threshold crossing was independent of the breathing gas.

For FSA the change from air to O<sub>2</sub> breathing caused tumor pO<sub>2</sub> to increase. During O<sub>2</sub> breathing, fewer tumors always remained below 10 mm Hg, down from seven tumors during air to four during O<sub>2</sub> breathing. Of those FSA tumors that fluctuated below the 10 mm Hg threshold, the pO<sub>2</sub> of the hypoxic intervals significantly increased during O<sub>2</sub> breathing ( $P < 0.05$ , signed rank test).

For the 9L tumor, there was no significant difference ( $P = 0.34$ ) in pO<sub>2</sub> comparing air (median, 9 mm Hg) to O<sub>2</sub> breathing (median, 9 mm Hg). The median pO<sub>2</sub> during hypoxic episodes, however, decreased with borderline significance from 6 mm Hg during air breathing to 1 mm Hg during O<sub>2</sub> breathing ( $P = 0.08$ , signed rank test), which was opposite to the change in median FSA pO<sub>2</sub>.

**Fourier Analysis of Tumor pO<sub>2</sub> Fluctuations.** Fourier analysis showed that pO<sub>2</sub> fluctuations typically occurred at very low frequencies (<1 cpm) for both FSA and 9L tumors. There were differences in the power spectra however (Fig. 4). The cumulative power of 9L is greater than FSA, indicating that fluctuations in 9L pO<sub>2</sub> were signif-

Table 2. Summary of partial pressure of oxygen fluctuations across a 10 mm Hg threshold

Variable	FSA		9L	
	Air pO <sub>2</sub> < 10	O <sub>2</sub> pO <sub>2</sub> < 10	Air pO <sub>2</sub> < 10	O <sub>2</sub> pO <sub>2</sub> < 10
Never hypoxic*	3 of 11	4 of 11	3 of 12	3 of 12
Always hypoxic	7 of 11	4 of 11	3 of 12	3 of 12
No. available for fluctuation analysis	8 of 11	7 of 11	9† of 12	9‡,§ of 12
Median no. of events/h (range) for all tumors	2 (0–2)	2 (0–6)	2 (0–8)	2 (0–10)
Median % time (range) hypoxic	100 (0–100)	77 (0–100)	62 (0–100)	79 (0–100)
Median no. of minutes (range) hypoxic	30 (0–30)	23 (0–30)	19 (0–30)	24 (0–30)
Median pO <sub>2</sub> (range) during interval	3 (1–15)¶,**,††	8 (0–31)¶,††	9 (1–22)**,††	9 (0–32)††
Median pO <sub>2</sub> (range) during hypoxic episodes‡‡	2 (0–8)§§,   ,††	5 (0–9)   ,††	6 (1–10)§§,††	1 (0–9)††

\* Hypoxic is defined as pO<sub>2</sub> < 10 mm Hg for at least 20 seconds (pO<sub>2</sub> was averaged over 10-second intervals).

† One tumor with a hypoxic duration of 29 minutes and 10 seconds out of 30 minutes.

‡ One tumor with a hypoxic duration of 29 minutes and 50 seconds out of 30 minutes.

§ One tumor with a median hypoxic pO<sub>2</sub> of 0 for a hypoxic duration of 28 minutes and 20 seconds out of 30 minutes.

|| Median pO<sub>2</sub> during the interval is the median pO<sub>2</sub> of the entire set of tumors for the full 30-minute period of analysis.

¶  $P < 0.005$  signed rank for air versus O<sub>2</sub>.

\*\*  $P < 0.05$  Wilcoxon for FSA versus 9L.

††  $P < 0.006$  Wilcoxon for FSA versus 9L in the difference between air and O<sub>2</sub>.

‡‡ Median pO<sub>2</sub> during hypoxic episodes is the median pO<sub>2</sub> below the 10-mm-Hg threshold for the subset of tumors available for fluctuation analysis.

§§  $P < 0.05$  Wilcoxon for FSA versus 9L.

|||  $P < 0.05$  signed rank for air versus O<sub>2</sub>.

icantly larger than in FSA ( $P < 0.005$ , Wilcoxon). The power spectra are similar between FSA and previously published data for R3230Ac (7). The power spectra for all tumors are greater than that which has been reported for normal rat muscle, using identical measurement methods (7).

**Tumor Blood Flow Fluctuations.** Tumor blood flow was measured at two sites in seven rats with FSA and nine rats with 9L tumors. Small variations of ~5% in blood flow occurred continuously in all of the tumors. In general, however, the range of fluctuations for blood flow was less than 30%, with 9L showing greater changes in relative blood flow than did FSA. Temporal characteristics of the two recordings made at different sites in the same tumor varied, with some tumors showing two completely independent traces and some tumors showing two temporally coordinated traces. There was no discernible change in fluctuations of relative blood flow for either FSA or 9L when the animal was switched from air to O<sub>2</sub>.

**Fourier Analysis of Tumor Blood Flow Fluctuations.** Fourier analysis showed dominant fluctuations occurring at low frequencies (<1 cpm) for FSA and 9L tumors, although contributing frequencies occurred over the range of 0 to 10 cpm (data not shown). There were differences in the power spectra, showing that the cumulative power of the 9L was greater than that of FSA during air breathing, particularly at low frequencies, although they did not reach statistical significance. O<sub>2</sub> breathing did not affect the frequency distribution appreciably for either tumor.

**Fourier Analysis of Changes in Frequency and Power in pO<sub>2</sub> and Blood Flow after Switch from Air to O<sub>2</sub> Breathing.** There was a significant increase in frequency of O<sub>2</sub> fluctuations in FSA during O<sub>2</sub> breathing between 0 and 10 cpm (data not shown). There was also a borderline significant increase in magnitude of fluctuations over the frequency range (0–10 cpm), with mean increases on the order of 400%. The relative frequency of fluctuations for 9L tumors was not changed when going from air to O<sub>2</sub> (data not shown). In contrast to FSA, however, there was a borderline significant decrease in the fluctuation magnitude (power) over the 0 to 10 cpm frequency range. The change in power spectra when switching from air to O<sub>2</sub> breathing between FSA and 9L in the fluctuation power was statistically significant ( $P = 0.01$ , Wilcoxon). FSA and 9L blood flow showed no significant changes in frequency or power with the switch from air to O<sub>2</sub> breathing.

## DISCUSSION

### Spatial Partial Pressure of Oxygen Distributions

These tumor lines were similar as assessed by pO<sub>2</sub> histograms, yet they were quite different with respect to details of the distribution. For comparison, previously published data for R3230Ac are included (7). Fig. 1 shows that all three tumors had a similar percentage of tissue below 10 mm Hg, that is, between 80 and 90%. However, 9L has a much larger fraction of its tissue at values less than or equal to 1 mm Hg, and FSA and R3230Ac both have a larger fraction of moderately hypoxic tissue (between 1 and 5 mm Hg) than does 9L. These measurements are in qualitative agreement with the pimonidazole staining seen in Fig. 2.

The 9L glioma showed large track lengths of pO<sub>2</sub> near zero, alternating with track lengths that were well above 10 mm Hg. This tumor is typically reported to have a very small radiobiological hypoxic fraction (11), but some investigators have found it to be radiobiologically hypoxic (19). In these experiments, we suspect that the long track lengths of near-zero pO<sub>2</sub> corresponded to zones of necrosis. The discontinuity, between the electrode histograms and what is known about the radiobiological hypoxic fraction, points to

one of the primary difficulties of using electrode measurements to predict radiation response. If measurements are made in areas of necrosis or nonclonogenic cells, the resultant electrode hypoxic fraction will overestimate the radiobiological hypoxic fraction (20, 21). The radiation response of FSA has not yet been determined, although the data from this study suggest that O<sub>2</sub> breathing would probably improve the radiation response.

### Partial Pressure of Oxygen Fluctuations

**Temporal Changes in pO<sub>2</sub> and Radiobiological Significance.** Description of temporal instability in tumor oxygenation can be done several ways based on measurements from this report. Important features of the description should include (a) the percentage of time that pO<sub>2</sub> lies within a radiobiologically significant range and the frequency that this occurs within a given experiment, and (b) the percentage of measurements in which pO<sub>2</sub> fluctuates into the range that would be considered radiobiologically important. These features of the temporal instability could influence radiotherapy response (3). The magnitude of the changes in pO<sub>2</sub> is also important, because this may influence the susceptibility of the tissue to hypoxia-reoxygenation injury.

We did not perform any experiments in which we observed stable pO<sub>2</sub> values. Stability here is defined as remaining at a value within the resolution limit of the electrodes, which is 1 mm Hg. This does not mean that all tumor regions are in a state of unstable oxygenation, however. Because we required a non-zero baseline pO<sub>2</sub> (to allow for increases or decreases in pO<sub>2</sub> over time), we selected for better oxygenated areas that might be more susceptible to fluctuations in pO<sub>2</sub> than areas of lower overall baseline pO<sub>2</sub>. For example, the average pO<sub>2</sub> from the histogram measurements in the 9L tumor was 1 mm Hg, whereas the average pO<sub>2</sub> for the fluctuant oxygenation measurements was  $14 \pm 9$  mm Hg. In contrast, the same pO<sub>2</sub> values averaged 4 and  $8 \pm 2$  mm Hg for FSA, respectively. There was no significant difference in baseline pO<sub>2</sub> measurements between the two tumor lines for the fluctuation studies ( $P = 0.11$ ). This minimizes the chance that there was selection bias that could have influenced the analysis of hypoxia fluctuation results.

Approximately 30% of the experiments remained above a 10-mm-Hg threshold for the duration of the kinetic study and, therefore, were not in a radiobiologically significant range. Similarly, some experiments remained below this threshold for the duration of the study (Table 2). For those experiments in which the threshold was crossed, the average number of crossings per hour was 2, regardless of the tumor type or the breathing gas. These results were similar to those published previously for the R3230Ac tumor (6, 7).

In recently published articles (22), we used the experimentally derived data from the present work to model the effects of pO<sub>2</sub> instabilities on radiotherapy response. For conventionally fractionated radiotherapy, we predicted that tumor control probability would be dominated by the average pO<sub>2</sub> observed during the 30-minute interval of observation. For more coarse fractionation schemes, such as stereotactic radiosurgery (six-field technique) or intraoperative radiotherapy with a high-dose-rate remote after-loading method, tumor control probability was substantially reduced, compared with conventional external beam fractionation, because there is increased likelihood that at least some of the treatment time is dominated by low pO<sub>2</sub> conditions. Oxygen enhancement ratio did not average near the maximal value of 3 in any experiments, indicating that the pO<sub>2</sub> fluctuations simulate a moderate degree of hypoxia and, therefore, a moderate degree of radioresistance. Wouters and Brown (23) have argued that cells of intermediate radiosensitivity are likely to dominate radiotherapy response, based on the argument that full reoxygenation does not

occur between fractions of radiation. Our data support the conclusion that intermediate levels of hypoxia are dominant, but this comes from a completely different logic, based on the direct measurements of pO<sub>2</sub> transients.

**Kinetics of pO<sub>2</sub> Fluctuations.** There is a substantial body of evidence supporting the idea that temporal kinetics of pO<sub>2</sub> and blood flow fluctuation in other rodent models and in human tumors are dominated by relatively slow fluctuations, on the order of 2 to 5 cycles per hour. For example, the radiation response of cells located near and far from perfused vessels, as marked with the perfusion marker drug Hoechst 33342, has been examined when the dye was given before or simultaneously with radiotherapy (3). When animals received the dye and were irradiated simultaneously, brightly stained cells were clearly more radiosensitive, supporting the theory that cells located nearer to vessels would be better oxygenated. However, when several minutes expired (typically 20–30 min) between dye administration and irradiation, the radiosensitivity of brightly and dimly stained cells was the same. These data suggested that, in the intervening minutes between dye administration and irradiation, some cells that were aerobic became hypoxic and *vice versa*. The 20-to-30-minute time interval to see this effect is consistent with our observation of cycle times on the order of 2 to 5 events per hour.

Similar conclusions have been made using pairs of fluorescent dyes that could be given either simultaneously or separated in time (2). Durand (1) suggested that subtle differences in staining intensity should also be taken into account, because such differences could indicate fluctuations in flow that could also contribute to intermittent hypoxia. Accordingly, he has argued that fluctuant hypoxia is likely to be a common physiological feature of tumors.

Work from this laboratory directly supports Durand's theory and also reveal a mechanism for the pO<sub>2</sub> instabilities. We measured perivascular pO<sub>2</sub> in skin-fold window-chamber tumors, concomitantly with measurements of microvessel red cell flux in the same vessels (24). Red cell fluxes varied temporally by as much as several orders of magnitude, but typical variations averaged ~2-fold. Perivascular pO<sub>2</sub> was found to be directly proportional to red cell flux within each microvessel. Thus, this work provided the first direct evidence that temporal variations in microvessel red cell flux can alter tissue oxygenation. The cycle frequencies were also on the order of 2 to 3 events per hour (7). Green's function models of typical networks, predicted that as much as 30% of a tumor region could experience transient hypoxia as a result of 2-fold changes in microvessel red cell flux (24). This prediction was higher than the observed incidence of vascular stasis in this model and in other models in which dye mismatch has been evaluated by Durand and LePard (1) and Trotter *et al.* (2).

One has to be cautious about extrapolating data from a window-chamber model to a larger more three-dimensional tumor. However, laser Doppler flow studies of red cell flux instability performed in a number of preclinical models as well as in human tumors show typical cycle times of 2-to-5-per-hour variation magnitudes of a factor of 1.5- to 3-fold. Thus, these data are in the range that we predicted would be sufficient to cause transient hypoxia using the Green's function models (1, 3, 25–27).

Brurberg *et al.* published two articles recently (8, 9) that directly corroborate the notion that tumor oxygenation is temporally unstable with a frequency of a few cycles per hour. In their work, they used the Oxford Optronix fiber optic oxygen sensor to measure transients in pO<sub>2</sub> in multiple sites in two early-passage human tumor xenograft lines. Fourier analysis as well as the direct evaluation of the number of crossings across a 10-mm-Hg threshold showed remarkable consistency with our prior data on the R3230Ac mammary tumor and with the data in this article (6, 7).

The process of hypoxia reoxygenation injury generates free radi-

cals, which may induce overexpression of defense mechanisms to protect cells from oxidative damage as well as being a potential cause of genetic instability (28–30). Cairns *et al.* (31) recently reported that exogenous manipulation of tumor oxygenation imposed by exposing animals to various levels of ambient oxygen can lead to increased frequency of single strand breaks and propensity toward metastasis *in vivo*.

Temporal instability in oxygenation may also have significant influence on angiogenesis. For example, we have recently shown that hypoxia-reoxygenation injury, such as that seen in this paper, leads to increases in the transcription of pro-angiogenic genes regulated by the hypoxia responsive promoter, hypoxia inducible factor-1 (HIF-1; Ref. 32). Two sources of increased HIF-1 activity were described. The first was related to stabilization of HIF-1 $\alpha$  created by the presence of elevated levels of reactive oxygen species. The second mechanism involved disaggregation of hypoxia-mediated stress granules during reoxygenation. It was shown that these granules contained protected pro-angiogenic HIF-1-mediated mRNA transcripts, such as vascular endothelial growth factor and plasminogen activator inhibitor 1.

**Causes of Temporal pO<sub>2</sub> Instability in Tumors.** There are at least four potential causes of fluctuant hypoxia: arteriolar vasomotion, vascular remodeling, variations in distribution of red cells at bifurcation points, and changes in oxygen consumption rate. Intaglietta *et al.* (33) used skin-fold window-chamber tumors to examine tumor-feeding arteriolar diameter while simultaneously examining downstream microvessel red cell flux. They noted temporal coordination in these events and suggested that arteriolar vasomotion could cause transient variations in perfusion that could be important for drug and nutrient transport (33). We have performed serial measurements of tumor arteriolar diameter, while simultaneously measuring downstream microvessel red cell flux. In some instances, the two processes appear to be temporally coordinated and occur with a periodicity of 2 to 3 cycles per hour (34, 35). This result suggests that arteriolar vasomotion may contribute to the temporal instability in red cell flux and vascular oxygenation in some cases. However, coordination between these events did not always occur, and this suggests that there are likely other mechanisms playing a role.

Vascular remodeling that occurs during tumor angiogenesis may be another key factor. Patan *et al.* (36), used skin-fold window chambers to perform serial observations of vascular architecture over periods of 1 to 2 hours. They commonly observed evidence for vascular intussusception, a process of vascular remodeling that would cause changes in microvessel flow resistance. The process of intussusception involves the in-growth of endothelial pillars into a vascular lumen, the consequence of which is to split a vessel segment into two or more smaller diameter vessels.

Variations in how red cells distribute at bifurcations can also alter flow resistance because microvessel hematocrit directly influences flow resistance (37). The influence of variations in red cell distribution on flow resistance would be exaggerated in tumors because the microenvironmental conditions cause reduced red cell membrane fluidity and an increased propensity toward Rouleaux formation (38).

**Fourier Analysis of Changes in Frequency and Power in pO<sub>2</sub> and Blood Flow after Switch from Air to O<sub>2</sub> Breathing.** Fourier analysis showed significant changes in power and frequency of pO<sub>2</sub> fluctuations when switching from air to O<sub>2</sub> breathing for both types of tumors. Analysis of FSA traces showed an increase in frequency and power of fluctuations after O<sub>2</sub> breathing began. Conversely, hyperoxic breathing had a dampening effect on the power of the 9L pO<sub>2</sub>, significantly decreasing the magnitude but not the frequency of the pO<sub>2</sub> fluctuations.

The frequency and magnitude of blood flow fluctuations did not change significantly for FSA or 9L tumors with the onset of O<sub>2</sub>

breathing. However, blood flow was measured using laser Doppler probes, which measure over larger areas of tissue than do microelectrodes (sensor diameter, ~200  $\mu\text{m}$  compared with 6  $\mu\text{m}$  for polarographic electrodes; Ref. 14). The lack of a measured concurrent change in blood flow could be due to the difference in spatial resolution between the two methods of measurement. Changes in microregional blood flow could be occurring over spatial volumes that are not measurable with laser Doppler. Similarly, microregional changes in flow could be occurring in different directions, such that when averaged over a relatively large volume, they yield no net change in flow as detected by laser Doppler.

This is the first report showing that oxygen breathing can affect O<sub>2</sub> transients and that there are tumor-specific effects. The cause of these changes in frequency and magnitude of the pO<sub>2</sub> fluctuations in tumors during air and O<sub>2</sub> breathing is unknown, but one possible reason for the fluctuations in pO<sub>2</sub> and blood flow may be changes in red blood cell flux induced by changes in vasomotor activity of tumor arterioles. Future work will examine potential mechanisms.

**Limitations of Study and Future Directions.** The data reported in this study are limited to roughly a 1-hour period of observation. It is possible that fluctuations occur over longer time intervals. Vascular remodeling, as occurs concomitantly with angiogenesis, could contribute to this process. A recent study using matched hypoxia markers showed that up to 15% of tumor cells may experience transient changes in oxygenation over a 12-hour period (5).

An important unknown with respect to fluctuant hypoxia is the size of regions that are affected. Does fluctuant hypoxia occur at the individual microvessel level, or are the fluctuations coordinated over small networks or an entire tumor? Existing data suggest that fluctuant hypoxia is not at the individual microvessel level. Temporal fluctuations in microvessel red cell flux in small groups (regions with 200–500- $\mu\text{m}$  diameters) of microvessels is typically coordinated (24). Vascular stasis, on the other hand, tends to occur sporadically (5–10% incidence) and involves single-vessel segments (34). The two papers by Brurberg *et al.* (8, 9) also substantiate that at least small microregions are being affected, as assessed by multiple Oxford Optronix sensors within individual tumors. These results suggest that the variations in red cell flux are not controlled at the whole tumor level. However, the probes are quite large, relative to the oxygen diffusion distance and sample a surface area around the probe tip that is  $3.8 \times 10^4 \mu\text{m}^2$  (14). A square region this size would be ~190  $\mu\text{m}$  in length. Such microregions would contain several microvessels (39). These results, then, suggest that the variations in red cell flux and consequent pO<sub>2</sub> occur over regions that comprise at least small groups of vessels (8, 9). Additional work is needed to clarify this issue further.

**Issues Related to Data Interpretation and Methods of Analysis: Effect of Measurement Device Drift on Fluctuations in Red Cell Flux and pO<sub>2</sub>.** We have ruled out the possibility that the fluctuations in pO<sub>2</sub> and red cell flux are due to drift in the measurement devices. First, the most compelling evidence comes from the work of Kimura *et al.* (24). In those experiments, red cell flux of individual microvessels was measured directly in-window-chamber tumors by observing the movement of fluorescently labeled red cells. Second, the ranking of power spectra for laser Doppler flow and pO<sub>2</sub> showed that the measurement for the 9L tumor was greater than that for the FSA. The fact that these two methods independently ranked the 9L tumor as having greater instability is indirect evidence that the fluctuations are not due to measurement drift. Finally, when the oxygen electrodes are calibrated *in vitro*, the currents obtained at set levels of oxygen are very stable (<1.0 mm Hg), indicating little or no drift.

Baseline instabilities in pO<sub>2</sub> and laser Doppler flow cause some difficulty in interpreting the response of the tissue to any oxygen-

manipulation protocol. Because it is not possible to obtain a stable baseline value, the challenge is to distinguish changes in pO<sub>2</sub> or blood flow that are caused by a physiological manipulation from fluctuation in these values due to the underlying physiology. In prior work, we developed a Bayesian statistics method to distinguish changes in pO<sub>2</sub> created by a manipulation from random fluctuation (40). Although Bayesian statistics were not used in this article, the results are fairly typical of what we observed in the prior Bayesian analysis for the R3230Ac tumor (40).

The change in Fourier transform results, after the switch from air to oxygen breathing, is less subject to interpretation errors because the data are obtained from blocks of time before and after the manipulation. Fourier analysis of a discrete sample assumes that the sample has periodic events with constant frequencies over the period of observation. Nothing was done during the experiments that would invalidate this assumption.

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