

Perspectives in Diabetes

Diabetic Retinopathy

Seeing Beyond Glucose-Induced Microvascular Disease

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Diabetic retinopathy remains a frightening prospect to patients and frustrates physicians. Destruction of damaged retina by photocoagulation remains the primary treatment nearly 50 years after its introduction. The diabetes pandemic requires new approaches to understand the pathophysiology and improve the detection, prevention, and treatment of retinopathy. This perspective considers how the unique anatomy and physiology of the retina may predispose it to the metabolic stresses of diabetes. The roles of neural retinal alterations and impaired retinal insulin action in the pathogenesis of early retinopathy and the mechanisms of vision loss are emphasized. Potential means to overcome limitations of current animal models and diagnostic testing are also presented with the goal of accelerating therapies to manage retinopathy in the face of ongoing diabetes. *Diabetes* 55:2401–2411, 2006

Despite years of clinical and laboratory investigation, diabetic retinopathy remains the leading cause of vision impairment and blindness among working-age adults, yet the fundamental cause(s) remains uncertain. Retinal photocoagulation to reduce neovascularization and macular edema was developed in the 1950s and is still the standard of care (1). The number of people worldwide at risk of developing vision loss from diabetes is predicted to double over the next 30 years (2), so it is imperative to develop better means to identify, prevent, and treat retinopathy in its earliest stages rather than wait for the onset of vision-threatening lesions. Progress in these areas requires a new perspective on the problem that includes the roles of the neural retina, impaired insulin action, and inflammation. In this way, established neurobiological principles can inform us how

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DCCT, Diabetes Control and Complications Trial; IRS, insulin receptor substrate; PI, phosphatidylinositol; PKC, protein kinase C; VEGF, vascular endothelial growth factor.

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diabetes impairs vision, and knowledge of metabolism, inflammation, and regenerative medicine may lead to new treatments.

This perspective will discuss how the unique anatomy and physiology of the retina may render it vulnerable to the metabolic derangements of diabetes and lead to impaired vision. The intent of this unconventional approach is to encourage consideration of new opportunities for investigations that will advance the field.

NORMAL RETINAL STRUCTURE AND PHYSIOLOGY

Topographic and cellular organization of the retina.

It is instructive to consider the functional organization of the retina (literally a network) to better understand the impact of diabetes (<http://webvision.med.utah.edu>). The retina is a transparent layer of neural tissue between the retinal pigmented epithelium and the vitreous body. Normal vision depends on intact cell-cell communication among the neuronal, glial, microglial, vascular, and pigmented epithelial cells of the retina. The fundamental functions of the retina are to capture photons, convert the photochemical energy into electrical energy, integrate the resulting action potentials, and transmit them to the occipital lobe of the brain, where they are deciphered and interpreted into recognizable images. The retina is partitioned from the systemic circulation by the blood-retinal and blood-aqueous barriers and receives its nutritional supply from the retinal and choroidal circulations and perhaps from the ciliary body by diffusion through the vitreous gel (3). As discussed below, the unique features of retinal anatomy and physiology that allow it to function so efficiently may also be its Achilles' heel under the stressful metabolic conditions of diabetes.

The retina operates in bright light to near darkness, so photoreceptor sensitivity varies over nine orders of magnitude (4). The macula operates in moderate- to bright-light conditions to subserve detailed acuity and color perception, whereas the peripheral retina operates in dim-light conditions, detects motion, and peripheral vision (Fig. 1A). The macula, ~6 mm in diameter in humans, is located between the vascular arcades temporal to the optic disc (5). The macula has the thickest ganglion cell layer and a high number of horizontal neuronal connections to resolve fine detail and discriminate contrast between objects. Beyond the macula, the retina has mostly rod photoreceptors and associated inner neurons, with fewer horizontal interconnections. The macula and peripheral retina operate reciprocally; therefore, the macula is

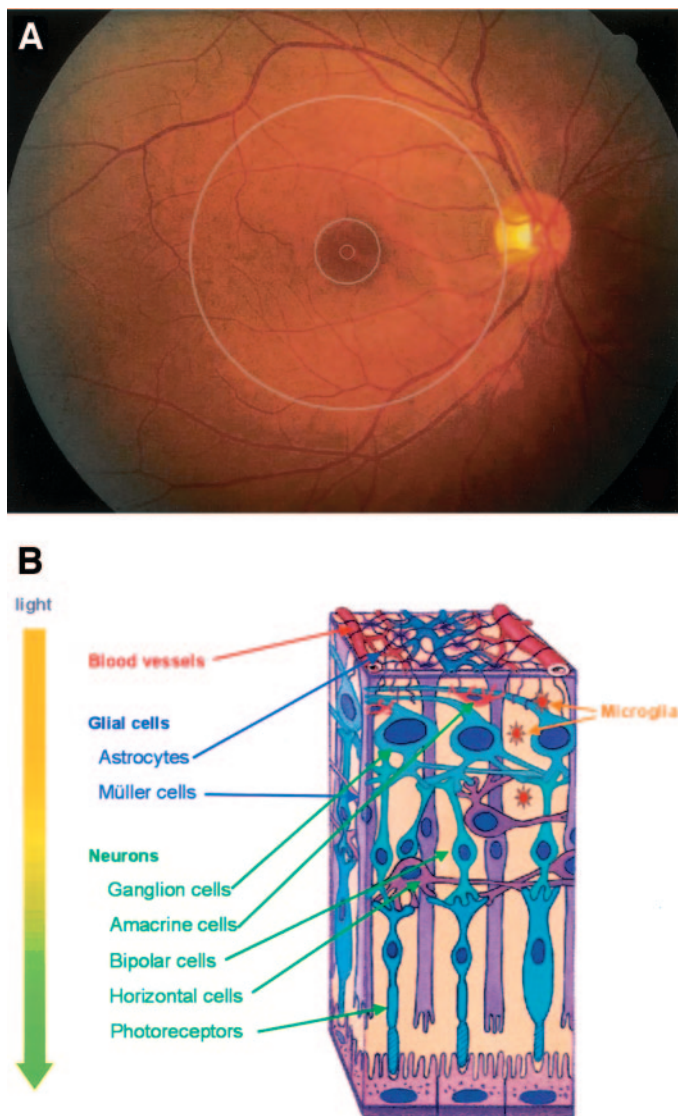


FIG. 1. Photograph of a normal human retina demonstrating the macula, fovea, and foveola in consecutively smaller circles (A). Lamellar retinal structure is shown with neurons, astrocytes, Müller cells, and microglial cells, as well as retinal pigment epithelium (B). Adapted from ref. 135.

more sensitive than the periphery in bright light, and the converse occurs in dim light.

The macula contains cone and rod photoreceptors; the center of the macula contains the fovea (pit), a 1-mm cone-dominated depressed region, and the central-most 200- μ m foveola is specialized for highest spatial resolution. The foveola contains only cone photoreceptors (first-order neurons), and the inner retinal cells are displaced so as not to interfere with light transmission. Outside the fovea, rod photoreceptors dominate and second- and third-order neurons (bipolar, amacrine, and ganglion cells) are present.

The lamellar cellular architecture of the retina (Fig. 1B) has alternating layers of neurons (outer and inner nuclear layers and ganglion cell layer) interposed with two plexiform layers, where neurons communicate at synapses between dendrites and between axons and dendrites. The retina includes five major cell types that perform sensory, regulatory, nutritional, and immunomodulatory functions. The neurons (photoreceptors, bipolar, horizontal, ama-

crine, and ganglion cells) perform sensory functions and define color perception, spatial resolution, and contrast discrimination (6).

Müller cells and astrocytes, two types of glial (glue) cells, provide nutritional and regulatory support for neurons. Müller cells span the retina from the pigment epithelium to the internal limiting membrane, a basement membrane formed by Müller cell end-feet that interfaces with the vitreous gel. Müller cells contact neurons and blood vessels in the plexiform and nerve fiber layers, and astrocytes envelop blood vessels in the nerve fiber and ganglion cell layers and contact ganglion cells and amacrine cells. Müller cells and astrocytes convey substrates, including lactate and amino acids, from the circulation to neurons and regulate blood-retinal barrier properties (7) and synaptic function (8). Müller cells also store glycogen for conversion to lactate, synthesize retinoic acid from retinol, regulate extracellular ion concentrations to modulate plasma membrane polarization/depolarization, participate with neurons in the glutamate/glutamine cycle to control neurotransmission, and protect neurons from glutamate excitotoxicity (9). Glial cells are the interface between the neurons and the vasculature and are thus key regulators of neuronal nutrition and metabolism.

The pigmented epithelial cell layer serves also as a selective conduit of substrates as the outer blood-retinal barrier and allows oxygen diffusion from the choroidal circulation to the outer retina. It removes retinal lactic acid and phagocytoses shed photoreceptor outer segments, comprises the outer blood-retinal barrier, absorbs light, secretes trophic factors such as pigment epithelium-derived factor, and, with the photoreceptors, participates in the cycling of the vitamin A isoforms retinol and retinal (10). Thus, this epithelial cell layer plays a crucial role in vision, although its role in diabetic retinopathy is not clear.

Immunomodulatory functions are provided by a fourth class of cells, microglia. They are a heterogeneous population of resident macrophages that monitor the local environment by interacting with neurons, glia, and endothelium and that react to stresses, including infection, trauma, and retinal detachment, by release of proinflammatory cytokines (11) and clearance of necrotic or apoptotic cells via phagocytosis (12). Microglial cells become activated and help to resolve local injury (13,14), but unrelenting stresses cause persistent inflammatory responses.

The fifth class of cells includes vascular endothelial cells and pericytes. They provide nutritional support and waste product removal for the inner retina and have been the focus of much research in diabetic retinopathy. It is likely that their function depends on as-yet-undefined signals from the neural retina. Blood vessels are the only structures that are visible by clinical examination because they convey erythrocytes containing the visible pigment hemoglobin. Despite their prominent appearance by clinical examination, the vasculature constitutes less than 5% of the retinal mass, so the retina is a vascularized neural tissue (15).

Retinal physiology may underlie its vulnerability to diabetes. The unique retinal structure imparts special physiologic constraints compared with other nervous system tissues because of the requirement for transparency, and these features may contribute to its susceptibility to diabetes. For example, retinal axons are not ensheathed by myelin, since myelin is opaque and blocks light transmission. Unmyelinated nerves require more energy to maintain membrane potentials than myelinated axons

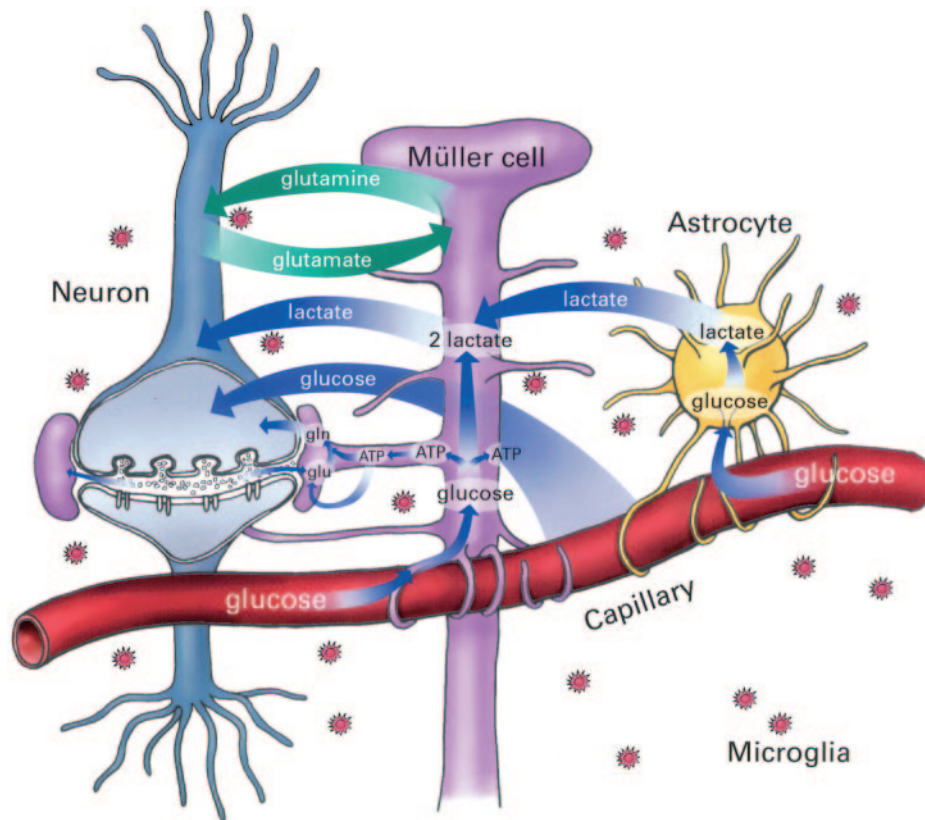


FIG. 2. Functional anatomy of the retina. Metabolic interactions in the retina between blood vessels (red), astrocytes (yellow), Müller cells (purple), and glutamatergic neurons (blue). Glucose (green) can pass directly from blood vessels to neurons. However, glucose is not oxidized in astrocytes and Müller cells but, instead, converted to lactate, which is transported out of the glia and into neurons for oxidation. Glutamate and glutamine are interconverted between glia and neurons. Adapted from ref. 136.

(16). Second, the density of blood vessels that would absorb light is relatively low, so the oxygen tension of the inner retina is relatively hypoxic with a pO_2 of only ~ 25 mm (17). The pO_2 gradient of the retina declines from the outer retina to the inner retina (17,18). Third, the inner retina possesses relatively few mitochondria that contain light-absorbing heme-based cytochrome proteins of the electron transport chain. Müller cells are relatively enriched in mitochondria and are found in regions of higher pO_2 in plexiform layers and ganglion cells (19) but not as prominently in the nuclear layers (20). Thus, the inner retina relies heavily on glycolysis, a less efficient means of generating ATP than oxidative phosphorylation, which predominates in the outer retina, where the pO_2 is ~ 80 mmHg (18,21). In spite of this sparse vascularity and low pO_2 , the retina has one of the highest metabolic demands of any tissue (22). ATP is required for phototransduction to maintain ion gradients across cell membranes, for neurotransmission at synapses, to (on a daily basis) replenish photoreceptor outer segment membranes, and to transport proteins and neurotransmitters anterograde and retrograde via axons to the optic nerve and lateral geniculate body of the thalamus (23). The combination of high metabolic demand and minimal vascular supply may limit the inner retina's ability to adapt to the metabolic stress of diabetes. By contrast, the outer retina receives its oxygen and nutrients by diffusion from the choroid through the pigmented epithelium and is relatively spared from the early insults of diabetes.

The retina is further specialized because, like the brain, its metabolic functions are compartmentalized between

glia and neurons. In glia of the inner retina, glucose metabolism occurs mostly via glycolysis, whereas cells of the outer retina fully oxidize glucose to CO_2 and water via oxidative phosphorylation (24). In the inner retina, metabolic substrates, such as glucose, flow from vascular endothelium to astrocytes to neurons. In the outer retina, substrates reach Müller cells and photoreceptors from the choroid via the pigmented epithelium. Thus, glial cells are vital to neuronal function because they convey circulating glucose into the retina for ATP production and provide intermediary compounds such as lactate. The functional anatomy of the retina is illustrated in Fig. 2.

Taken together, these findings strongly suggest that the unique anatomic and physiologic specialization required for vision demands intact cell-cell communication. This specialization may, in turn, predispose the retina to diabetes-induced damage if the metabolic derangements typical of diabetes interfere with the generation of neurotransmitters, macromolecule synthesis, or induce proapoptotic or proinflammatory responses.

DIABETIC RETINOPATHY: BEYOND GLUCOSE-INDUCED MICROVASCULAR DISEASE

Numerous investigators have suggested that the pathogenesis of diabetic retinopathy includes glucose-mediated microvascular damage. Previously implicated pathways related to excess glucose include oxidative stress, activation of protein kinase C (PKC), and activation of advanced glycation end products and their receptor (25–29). Mechanisms of vascular injury include increased vascular per-

meability due to tight junction disassembly (30) and endothelial cell-mediated leukostasis (31). These studies have led to potential therapeutic approaches, including PKC inhibitors, corticosteroids, and soluble receptor for advanced glycation end product inhibitors (25); of these, PKC inhibition has been shown to be effective in randomized clinical trials (32). However, recent work strongly suggests that diabetic retinopathy involves more than elevated glucose and microvascular lesions, so evidence for alterations of the neural retina and insulin action are presented below.

Evidence for neural retinal involvement in diabetic retinopathy. First, although microvascular changes are undeniably integral to retinopathy, the retina is a vascularized neural tissue, not a network of blood vessels. Histopathologic studies emphasized the loss of neurons in human diabetic retinopathy >40 years ago (33,34). Since then, numerous reports using electroretinography, dark adaptation, contrast sensitivity, and color vision tests have conclusively demonstrated that neuroretinal function is compromised before the onset of vascular lesions in humans (35–38,39). In fact, loss of oscillatory potentials on electroretinograms predicts the onset of proliferative retinopathy better than vascular lesions seen on fundus photographs or capillary nonperfusion visualized by fluorescein angiograms (40). Electroretinograms and psychophysical tests are primarily used in research settings, but recent reports using clinically available visual field test modifications (short-wave automated perimetry and frequency doubling perimetry) reveal field defects in patients with little or no vascular retinopathy (41,42), and visual fields predict the severity of retinopathy better than visual acuity (43). Together, these findings strongly suggest that functional tests are more sensitive indicators of retinal integrity than fundus photographs or optical coherence tomography and may serve as useful end points for clinical trials, but they must be validated first.

At the cellular level, diabetes alters the function and structure of all retinal cell types. Postmortem human diabetic retinas exhibit increased markers of apoptosis in ganglion cells (44). Animal studies show accelerated apoptosis of retinal neurons (45,46), glial activation (47–49), impaired glial cell metabolism (50–52), and microglial cell activation (11,53,54).

Together, these studies leave little doubt that neural retinal defects are among the earliest detectable changes in diabetes. Regardless of whether the initial events begin in blood vessels or neural cells, the clinical stages of diabetic retinopathy manifest cellular, histologic, and functional features of a retinal neuropathy (15,55,56). To the best of our knowledge, there is no evidence that a primary, selective defect in vascular cells is sufficient to cause diabetic retinopathy. Clearly, it is essential to treat both the vascular and neural elements of the retina to preserve vision. This concept permits a new paradigm for understanding the mechanism of vision impairment in diabetes and provides therapeutic targets that are directly linked to vision (57,58).

A feed-forward concept of diabetic retinopathy involving neural and vascular lesions. In general, acute self-limited cellular stresses, such as acute bacterial or viral infections, lead to physiologic adaptive inflammatory responses that permit healing. By contrast, chronic infections, such as hepatitis or tuberculosis, rheumatoid arthritis, or diabetes, induce maladaptive inflammatory responses and culminate in pathology. In this light, which

cellular stresses might initiate diabetic retinopathy and which responses might perpetuate or resolve it? Which cells initially sense the stress and respond? Several possibilities exist and must be considered.

If diabetes exerts its primary damage on vascular cells and increases permeability or vascular occlusion, neuronal and glial cell integrity would be compromised by the entry of circulating macrophages, antibodies, inflammatory cytokines/chemokines, excitotoxic amino acids, or fatty acids into the retina. On the other hand, if diabetes primarily affects the neural retina, it could compromise vascular integrity by loss of normal barrier-inducing functions of glia or increased expression of proinflammatory cytokines or reactive oxygen species that promote vascular leakage or occlusion. At this point, it is not known whether vascular or neural cell defects occur first; most likely they are interdependent. Thus, we propose a feed-forward concept of vascular-neural dysfunction that begins shortly after the onset of diabetes and increases over time, creating a widening vortex of retinal injury (Fig. 3). Eventually, accumulated injury and failing reparative responses lead to the clinically evident features of diabetic retinopathy.

Inflammation in diabetic retinopathy. Inflammation is a prominent component of many diseases, including primary retinal degenerations, insulin resistance, and diabetes (59). Diabetic retinopathy was initially termed, “diabetic retinitis” (60), although this usage fell out of favor in the 1970s. Chronic inflammation is characterized by increased vascular permeability, edema, inflammatory cell infiltration, cytokine and chemokine expression, tissue destruction, neovascularization, and attempts at repair (61), and diabetic retinopathy exhibits most of these features (Table 1). Microglia associate intimately with neurons that express molecules such as CX3CL1 (fractalkine) (62) and CD200 (63) that negatively regulate microglial activation through their respective receptors. As such, perturbation of expression of ligand or receptor during stress would activate microglia to produce proinflammatory cytokines and acquire an activated morphology (11,64). Activated microglia produce chemokines such as monocyte chemoattractant protein-1 (11), inducing expression of adhesion molecules, which can promote the leukostasis of neutrophils, on endothelium (65) and potentially inducing the extravasation of inflammatory macrophages (66). In addition, elevated levels of complement and reduced levels of complement inhibitors (67) and acute-phase proteins (68) are likely key events in the phagocytic clearance of necrotic and apoptotic neurons (12). Recent evidence strongly suggests that inflammation involving vessels and neural tissue occurs early in experimental (11,64,65,67,69) and human (70) retinopathy, involving humoral and cellular components of innate immunity.

Physiologic repair processes that help retinal cells survive stress include increased expression of numerous growth factors and cytokines, including vascular endothelial growth factor (VEGF), IGF-1, interleukin-1 β , and tumor necrosis factor- α (rev. in 71). These proteins, which have been implicated in the development of retinopathy, also provide neurotrophic functions to support retinal cell survival (71–73). Diabetes-induced upregulation of these growth factors may be an attempt to reestablish a pro-survival-state equilibrium while simultaneously altering the blood-retinal barrier and inducing inflammatory responses (Fig. 4). In the short term, increased cytokine/chemokine

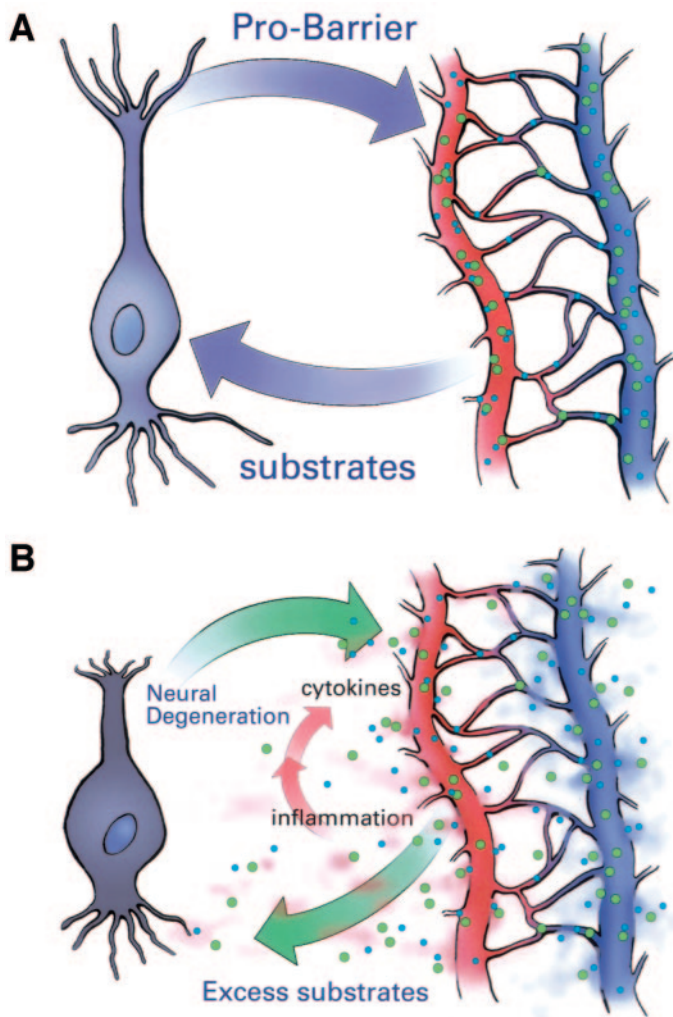


FIG. 3. A feed-forward concept of diabetic retinopathy. Under normal conditions (A) blood vessels supply neural tissue with nutrients. In turn, neural cells produce factors that induce the blood-retinal barrier so the retina can control the local biochemical environment and protect the retina from circulating antibodies, inflammatory cells, and amino acids. In diabetes (B) normal probarrier stimuli from the neural retina may be compromised, allowing blood-borne elements to enter the retina and damage neural cells through a chronic inflammatory process. Neurodegeneration leads to induction of growth factors (e.g., VEGF) and loss of probarrier factors, aggravating vascular damage, including capillary occlusion and permeability. Thus, a feed-forward cycle of combined vascular and neural damage ultimately leads to impaired vision.

TABLE 1
Features of inflammation in diabetic retinopathy

Increased blood flow and vascular permeability
Tissue (macular) edema
Accelerated cell death (45,100)
Macrophage infiltration (70), microglial cell activation (11,53,54)
Increased leucocyte adhesion (31)
Increased cytokine expression (VEGF, IGF-1, IL-1 β , others) (71,137)
Complement activation, FAS ligand upregulation (67,69)
Acute phase response protein expression (68)
Neovascularization
Glial cell proliferation

Reference numbers are in parentheses. FAS, fatty acid synthase; IL, interleukin.

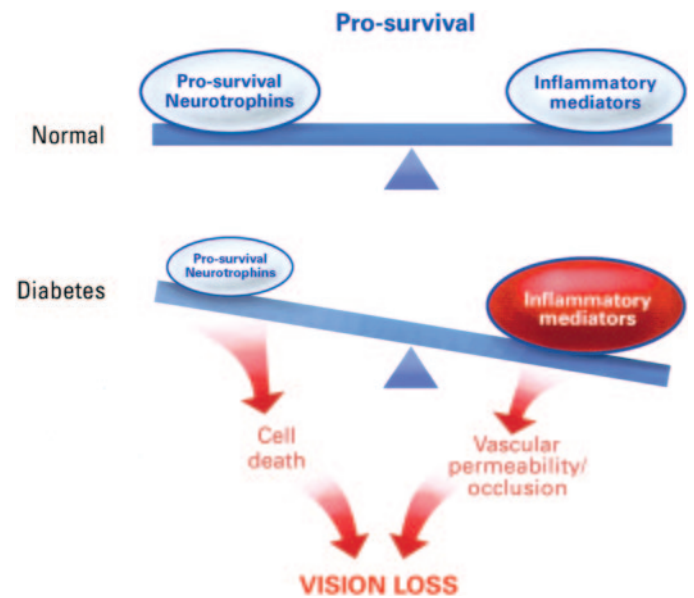


FIG. 4. Diabetes disturbs the homeostatic equilibrium of the retina. Under normal conditions, there is an equilibrium in which prosurvival and anti-inflammatory stimuli maintain retinal cell survival and function. In diabetes, prosurvival (neurotrophic) inputs may be reduced and proinflammatory cytokines, chemokines, and cellular responses increased. Together, these processes accelerate retinal cell death and increase vascular permeability and occlusion, thus impairing vision. Treatments may be directed at augmenting neurotrophic inputs and decreasing proinflammatory responses so that repair processes can predominate.

expression may serve an adaptive function to maintain neuronal function but, over time, becomes maladaptive, with progressive vascular damage, ultimately resulting in macular edema and neovascularization. The feed-forward process thereby perpetuates both vascular and neuronal damage and culminates in the clinical features of diabetic retinopathy.

In this light, the issue is not simply whether diabetes incites a retinal inflammatory process but whether the inflammatory response contributes to initiation, propagation, or resolution of the injury. Impaired wound healing is a long-recognized feature of diabetes, and recent evidence indicates that diabetic animals have impaired reparative responses to central nervous system metabolic stress. For example, hypoxic/ischemic stress in normal mice brain increases expression of the antiapoptotic genes *bcl-2* and *blf-1* in microglial cells. However, diabetic *db/db* mice do not mount this reparative response and have larger brain infarcts than control animals (74). Thus, the normal inflammatory response is designed to limit tissue injury, and diabetes may disrupt the ability of tissues to respond appropriately and recover. Further characterization of the temporal sequence and balance of these injurious and reparative responses in the retina is required to understand how to minimize tissue injury. Therefore, if the upregulation of growth factors and cytokines, such as VEGF, serves to support cell survival during diabetes, it is possible that their inhibition by pharmacologic means could aggravate retinal neuronal or vascular cell death while apparently improving the clinical picture if the retinal cells that express the growth factors/cytokines are dying (75). Nevertheless, from a therapeutic point, it is likely that identifying and halting key steps of the inflammatory process contributing to tissue destruction will help to control retinopathy.

Taken together, recognition of how diabetes disrupts the entire retina also illustrates how diabetes alters the parenchyma of other tissues, including adipose, skeletal muscle, kidney, heart, brain, peripheral nerve, and bone. In fact, the concept of “microvascular” complications does not adequately describe diabetic retinopathy, nephropathy, or neuropathy and should be reevaluated.

Evidence for impaired insulin action in diabetic retinopathy. Excess glucose is generally considered to be the primary culprit in the development and progression of diabetic retinopathy. Diabetes mellitus (honey urine) is literally and clinically defined as a disorder of carbohydrate metabolism, so glucose is a logical culprit to consider. However, disordered lipid and protein metabolism are also integral to and causally linked to the central biochemical abnormality in all forms of impaired insulin action. The experimental case for a central role of glucose is based largely on studies of retinal pathology in experimental galactosemia in rodents and dogs (76,77). These animals have normal plasma insulin levels yet develop retinal vascular lesions similar to those in diabetes; however, neural retinal changes have not been examined in this model.

The clinical case for a central role of glucose is based on the strong statistical association between HbA_{1c} (A1C) levels and retinopathy (78–80). However, data from the Diabetes Control and Complications Trial (DCCT) also show a lower risk of retinopathy for the same A1C levels in patients who received intensive control compared with those who received conventional treatment (81). Thus, either the consistency of euglycemia and/or fewer periods of relative insulin deficiency may account for the benefits of intensive therapy. In the DCCT, the reduced risk of retinopathy was achieved by giving insulin more frequently to patients in the intensive-treatment groups (82). Hence, the DCCT might reasonably be viewed as an insulin dose-response trial (83). Further clinical evidence for a role of insulin is that systemic insulin resistance is a risk factor for retinopathy in patients with type 1 diabetes in the DCCT (84), the EURODIAB study (85), and the Pittsburgh Epidemiology of Diabetes Complications study (86). In type 2 diabetes, insulin deficiency is also an independent risk factor for the presence of retinopathy (87,88). These data strongly suggest a clinical role for systemically administered insulin and raise the question of whether the retina, like other tissues, is a direct target of insulin.

Constitutive retinal insulin receptor signaling is reduced in diabetes. In liver and skeletal muscle, insulin binds to its receptor tyrosine kinase and increases the phosphorylation of a series of protein and lipid kinases to produce tissue-specific biologic responses. The liver and muscle lack a blood-tissue barrier and are exposed to fluctuating insulin levels through normal feeding/fasting cycles. We have reported (89) that retinal insulin signaling is substantially different from that in liver and muscle. The retina, with its blood-retinal barrier, possesses a constitutively active insulin receptor signaling system that has basal tyrosine kinase activity equivalent to that in postprandial liver and that does not vary with feeding and fasting. The activities of the insulin receptor tyrosine kinase and the linked prosurvival enzymes, phosphatidylinositol (PI) 3-kinase, Akt, and p70S6 kinase, are all several-fold greater in the retina than in skeletal muscle or liver of normal rats. This high constitutive activity is consistent with the high metabolic requirements of the normal retina.

Diabetes disrupts insulin signaling in peripheral tissues. Recent experimental diabetes causes rapid and progressive loss of the constitutive retinal insulin receptor kinase activity and downstream PI 3-kinase → p70S6 kinase signaling cascade (90–92). A key distinction between retina and peripheral tissues is that the former suffers from loss of high basal activity rather than from reduction of plasma insulin-stimulated responses. Therefore, it appears that diabetes per se contributes to primary biochemical defects in the retina via loss of constitutive insulin signaling. Moreover, incubation of retinal neuronal cell cultures in high-glucose-containing medium, coupled with the stress of with serum deprivation, reduces insulin-stimulated Akt phosphorylation and cell survival (93). Hence, the retina is clearly an insulin-sensitive tissue, and excess glucose or lipids may exert their noxious effects in part by downregulating insulin signaling in retina as in peripheral tissues.

Reduced insulin action leads to neurodegeneration. In skeletal muscle, insulin stimulates anabolic functions and prevents tissue breakdown, and loss of insulin action in diabetes causes muscle wasting. Evidence is now accumulating suggesting that the insulin receptor also provides trophic functions in the brain via the PI 3-kinase/Akt pathway (94). In the retina, insulin stimulates insulin receptor substrate (IRS)-2 tyrosine phosphorylation (89); retinal IRS-2 content is decreased in diabetic rats (90), and IRS-2 deletion leads to degeneration of inner retinal neurons and photoreceptors (95). Hence, it is reasonable to hypothesize that decreased insulin-stimulated prosurvival stimuli, at least in part via loss of PI 3-kinase/Akt-mediated effects, may contribute to retinal cell death. Indeed, disruption of insulin receptor signaling has a profound impact on retinal cell growth and development in *Drosophila*, chickens, and mice (96–98). Retinal neurons (73) and vascular cells (99) depend on insulin receptor activity for survival, and both types of cells die by apoptosis in both human and rat models of diabetes (45,100). It is reasonable to predict that long-term disturbances in retinal insulin signaling may accelerate cell death and impair insulin-dependent anabolic activities, such as protein synthesis (101). Therefore, insulin signaling appears to provide neurotrophic actions in the retina, and retinopathy may result in part from neurotrophin deficiency, as shown for peripheral neuropathy (102,103). In this light, the persistent benefit of initial intensive control in DCCT/EDIC (Epidemiology of Diabetes Interventions and Complications) patients despite subsequent deterioration of A1C values (104,105) could result from enhanced neurotrophic input from insulin on retinal and peripheral nerve cells.

The findings that diabetes impairs insulin receptor signaling in retina, brain (106–109), and peripheral nerve (110), as well as in classic insulin-sensitive tissues, suggest that the metabolic impact of diabetes on “complications”-prone tissues and peripheral tissues forms a continuum. Whereas muscle and adipose respond acutely to fluctuating insulin levels and change rapidly after diabetes onset, retina and brain insulin action has a higher set point and responds less rapidly after diabetes onset. Plasma insulin penetrates the eye and brain more slowly than peripheral tissues, and with saturable kinetics (111–114). Thus, it is important to consider the role of impaired insulin action in the development of complications, even in tissues where insulin does not regulate glucose uptake. “Complica-

TABLE 2
Possible mechanisms of vision impairment in diabetes

Cellular defects	Clinical features	Effects on visual function
Increased vascular permeability, capillary nonperfusion	Symptoms: decreased central acuity; signs: retinal thickening, cystoid macular edema, lipid exudates	Light scattering within retina blurs image; cysts compress neurons; loss of glutamate; glutamine cycling between glial cells and neurons; increased susceptibility of neurons to plasma-derived toxic factors; possible ischemia of neurons
Primary neuronal impairment	Symptoms: decreased night and color vision; signs: nerve fiber layer defects, retinal depression sign, normal-appearing retina	Reduced contrast sensitivity, dark adaptation, color vision, ERG responses

ERG, electroretinogram.

tions," then, are the manifestations of altered diabetes metabolism in organs that are associated with clinical impairment.

Retinopathy in the absence of or before onset of hyperglycemia. Further clues that suggest excess glucose may not explain all aspects of retinal pathogenesis come from numerous well-documented cases of retinopathy in the absence of overt hyperglycemia (115–121). The patients described in these reports had normal or impaired glucose tolerance but did not have overt diabetes, yet some had typical proliferative retinopathy and diabetic nephropathy. In the Diabetes Prevention Program, 8% of patients with impaired glucose tolerance (pre-diabetes), but without overt hyperglycemia, had retinal microaneurysms (Diabetes Prevention Program, unpublished data). Collectively, these reports from many investigators warrant an explanation and lead to the question of whether hyperglycemia per se is necessary or sufficient to initiate or perpetuate retinopathy. These findings further suggest that the available data on the effects of excess plasma glucose per se do not causally account for the range of cellular and functional changes in diabetic retinopathy.

Overall, the data support the idea that impaired insulin action, the primary defect of diabetes, directly impacts the retina and may initiate retinal dysfunction. This leads us to question 1) whether this change results from loss of pancreatic insulin and 2) whether retinopathy is due to this alteration alone or if retinopathy results from combinatorial insults. Potential factors that may alter the retinal homeostatic equilibrium include glucose, lipids, hypertension, counterregulatory hormones (glucagon, glucocorticoids, adipokines), or inflammation due to insulin resistance.

HOW DOES DIABETES IMPAIR VISION?

Impaired vision in patients with diabetes is most frequently associated with macular edema, macular ischemia, epiretinal membranes that distort or elevate the macula, or vitreous hemorrhages that obscure the ocular media. For example, leakage of retinal capillaries contributes to macular edema, and it is generally thought that the clinically apparent vascular leakage accounts for the impaired vision. However, the specific cellular mechanisms by which macular edema reduces visual acuity have not been well defined. From an optical perspective, macular cysts scatter light within the retina (T.W.G., personal observation) such that it is not focused on photoreceptors, decreasing the image quality. From a cellular standpoint, visual function could decline if fluid accumulation within the retina 1)

alters extracellular ionic concentrations required for action potentials, 2) physically compresses retinal neurons, 3) compromises the normal interchange of glutamate and glutamine between glial cells and neurons required for neurotransmission, or 4) neurons have increased susceptibility to excitotoxic amino acids, antibodies, or inflammatory cells that reach the retina via leaking vessels. Vascular leakage could thus impair neuronal function and vision in the absence of clinically detectable macular edema. Occluded capillaries near the fovea may also cause retinal neurons to suffer ischemic damage. Table 2 describes possible mechanisms of vision impairment in diabetes.

Direct diabetes damage to glial cell or neuronal metabolism would directly impact neurotransmission (38,122) and may lead to apoptosis of retinal neurons and visual field defects. Indeed, retinal axons are lost before the onset of visible vascular lesions (123,124). Recent reports also demonstrate that impaired local responses on multifocal electroretinograms predict subsequent development of vascular lesions (125). Vision depends on neuronal function, so in the final analysis, all forms of vision impairment with clear ocular media (macular edema, macular ischemia, traction retinal detachment) must, by definition, include neuronal dysfunction. Further work is needed to determine how alterations in vascular, glial, microglial, and neuronal cell interactions reduce the quality of vision.

WHEN DOES RETINOPATHY BEGIN?

Retinopathy is diagnosed clinically with the onset of ophthalmoscopic signs such as microaneurysms, hemorrhages, and cotton-wool spots, but as discussed above, functional defects often precede these signs. If diabetic peripheral neuropathy is diagnosed on the basis of subtle electrophysiologic and sensory defects before the onset of clinical symptoms, should retinopathy not be categorized before the onset of vascular lesions? This interval should provide the best window of opportunity for treatment while visual function remains intact. Individuals who have not developed clinically evident retinopathy represent the greatest therapeutic opportunity to preserve vision because they constitute the majority of the patients and respond better to intensive therapy (79). Therefore, development of sensitive, predictive measures of retinal function that could be employed in practice and clinical trials would have great utility.

OBSTACLES AND OPPORTUNITIES FOR PREVENTION AND TREATMENT

Diabetic retinopathy remains a plague of modern society. Advanced vascular lesions of diabetic retinopathy have been treated by destructive panretinal photocoagulation and vitrectomy surgery for the last 5 decades. In comparison, peptic ulcer disease was once a common cause of disability, at a time when the diagnosis depended on radiographic studies and treatment was surgical repair of bleeding ulcers. Endoscopy now allows early diagnosis, and research has elucidated the role of gastric *Helicobacter pylori* infections and acid overproduction; antibiotic and acid-blocking medications have substantially reduced the mortality and morbidity of peptic ulcers. We are looking for opportunities to make detection and treatment paradigms as effective as those for peptic ulcer disease. Of the major blinding retinal diseases (macular degeneration, retinitis pigmentosa, and diabetic retinopathy), retinopathy is the only one for which a specific medical intervention (intensive insulin therapy) has been shown to retard both its development and progression. Improved medical care over the past 3 decades has reduced the risk of vision-threatening retinopathy (126); therefore, there is ample reason for optimism that retinopathy can be prevented.

Nevertheless, several factors limit the progress toward prevention or cure of retinopathy in the face of imperfectly controlled diabetes. A key step includes development of clinically practical means to diagnose retinal dysfunction before the onset of vascular lesions and symptomatic vision loss. Retinal function tests that can be utilized clinically, such as short-wave automated or frequency-doubling perimetry or dark adaptation, might predict disease onset and progression. Development of such tests could shorten the duration of therapeutic clinical trials by providing end points acceptable to the U.S. Food and Drug Administration, unlike the current threshold of three lines of visual acuity change or three-step change on the Early Treatment of Diabetic Retinopathy Study retinopathy scale.

The models available for *in vivo* retinopathy studies also restrain progress. Although the retina is easily visible, it is, ironically, the only major tissue affected by diabetes that cannot be biopsied in humans. Vitreous samples from patients with proliferative retinopathy reflect late-stage changes but are not readily available in the early stages of disease. Rodent models of diabetes, mostly streptozotocin-induced rats and mice (127) and *Ins2^{Akita}* mice (92), reveal retinal vascular and neural alterations. These models are relatively inexpensive and can provide detailed molecular information, including acute and chronic changes in gene expression and cell signaling. Conditional gene knockout studies in mice could provide substantial insights by mimicking or altering disease progression. However, evolutionary distance from humans and inbreeding may not accurately predict disease in genetically diverse humans. For example, the roles of the polyol pathway and aldose reductase inhibitors predicted by rodent studies were not borne out by human studies of sorbinil or tolrestat. The small eyes of rodents also do not have maculas or permit validation of pharmacokinetic data needed to develop drug delivery devices for patients. Therefore, investment to determine the potential of large animals such as pigs or nonhuman primates for retinopathy research may be efficient and cost-effective over the long term. Clinical and histopathologic signs of retinopathy develop in type 2

diabetic monkeys (128–131), but further investment is needed to develop large animal models of type 1 and type 2 diabetes for complications research (132). Parallel studies in humans should permit correlation of the systemic metabolic profiles and retinal phenotypes of animal models so the information gained in animals can be translated more readily to human studies.

The development of new treatments for retinopathy is also hampered by the slow course of the clinical vascular changes. However, evidence that a ruboxistaurin, a PKC inhibitor (32), and pancreatic transplants (133) can modify the course of established retinopathy indicates that progression of the disease can be modified. Intensive insulin treatment is the ideal means to prevent and treat mild retinopathy, but hypoglycemia limits the ability of many patients to achieve the degree of control needed to prevent retinopathy (134). Most patients are not candidates for pancreatic transplantation, and islet cell or stem cell transplants are not widely available. The key to future treatments lies in elucidating the biological processes that lead to retinopathy. Therapeutic strategies may involve dual arms that 1) identify and augment deficient neurotrophin pathways and 2) inhibit key proinflammatory/proapoptotic pathways to restore normal vascular and neuronal function (Fig. 4). This dual approach may lead to improved management of retinopathy in the face of continued diabetes.

SUMMARY

The intent of this perspective is to synthesize recent clinical and laboratory findings and to engender new approaches to the diagnosis and treatment of diabetic retinopathy, with increased emphasis on prevention and early intervention. Better understanding of the biology of diabetic retinopathy will require genetic dissection of the molecular pathways in rodents, determination of the interrelationships between vascular and neural cells, and proof-of-principle studies in large animal models. Additional work is required to understand how the early cellular and biochemical mechanisms of retinal injury and stress responses impair vision, which molecular components can be targeted for therapeutic intervention, and how best to introduce drugs to the retina with minimal systemic impact. Enhanced collaboration between groups of laboratory and clinical investigators with better-defined criteria for moving drugs tested in animals into clinical trials will accelerate preservation of vision in people with diabetes.

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REFERENCES

- Meyer-Schwickerath G: Licktkoagulation. *Buech Augenarzt* 33:1-96, 1959 [article in German]
- Wild S, Roglic G, Green A, Sicree R, King H: Global prevalence of diabetes: estimates for the year 2000 and projections for 2030. *Diabetes Care* 27:1047-1053, 2004
- Bito LZ, Salvador EV, Petrinovic L: Intraocular fluid dynamics. IV. Intraocular sites of solute utilization and transport as revealed by studies on aphakic eyes. *Exp Eye Res* 26:47-55, 1978
- Reeves A: Visual adaptation. In *The Visual Neurosciences*. Chalupa LM, Werner JS, Eds. Cambridge, MA, MIT Press, 2004, p. 851
- Curcio CA, Sloan KR, Kalina RE, Hendrickson AE: Human photoreceptor topography. *J Comp Neurol* 292:497-523, 1990
- Masland RH: The fundamental plan of the retina. *Nat Neurosci* 4:877-886, 2001
- Gardner TW, Lieth E, Khin SA, Barber AJ, Bonsall DJ, Leshner T, Rice K, Brennan WA Jr: Astrocytes increase barrier properties and ZO-1 expression in retinal vascular endothelial cells. *Invest Ophthalmol Vis Sci* 38:2423-2427, 1997
- Newman EA: New roles for astrocytes: regulation of synaptic transmission. *Trends Neurosci* 26:536-542, 2003
- Lieth E, LaNoue KF, Berkich DA, Xu BY, Ratz M, Taylor C, Hutson SM: Nitrogen shuttling between neurons and glial cells during glutamate synthesis. *J Neurochem* 76:1712-1723, 2001
- Strauss O: The retinal pigment epithelium in visual function. *Physiol Rev* 85:845-881, 2005
- Krady JK, Basu A, Allen CM, Xu Y, Lanoue KF, Gardner TW, Levison SW: Minocycline reduces proinflammatory cytokine expression, microglial activation, and caspase-3 activation in a rodent model of diabetic retinopathy. *Diabetes* 54:1559-1565, 2005
- Elward K, Gasque P: "Eat me" and "don't eat me" signals govern the innate immune response and tissue repair in the CNS: emphasis on the critical role of the complement system. *Mol Immunol* 40:85-94, 2003
- Fetler L, Amigorena S: Neuroscience: brain under surveillance: the microglia patrol. *Science* 309:392-393, 2005
- Nimmerjahn A, Kirchhoff F, Helmchen F: Resting microglial cells are highly dynamic surveillants of brain parenchyma in vivo. *Science* 308:1314-1318, 2005
- Gardner TW, Antonetti DA, Barber AJ, LaNoue KF, Levison SW: Diabetic retinopathy: more than meets the eye. *Surv Ophthalmol* 47 (Suppl. 2):S253-S262, 2002
- Bristow EA, Griffiths PG, Andrews RM, Johnson MA, Turnbull DM: The distribution of mitochondrial activity in relation to optic nerve structure. *Arch Ophthalmol* 120:791-796, 2002
- Wangsa-Wirawan ND, Linsenmeier RA: Retinal oxygen: fundamental and clinical aspects. *Arch Ophthalmol* 121:547-557, 2003
- Pournaras CJ: Retinal oxygen distribution: its role in the pathophysiology of vasoproliferative microangiopathies. *Retina* 15:332-347, 1995
- Germer A, Biedermann B, Wolburg H, Schuck J, Grosche J, Kuhrt H, Reichelt W, Schousboe A, Paasche G, Mack AF, Reichenbach A: Distribution of mitochondria within Muller cells. I. Correlation with retinal vascularization in different mammalian species. *J Neurocytol* 27:329-345, 1998
- Bentmann A, Schmidt M, Reuss S, Wolfrum U, Hankeln T, Burmester T: Divergent distribution in vascular and avascular mammalian retinae links neuroglobin to cellular respiration. *J Biol Chem* 280:20660-20665, 2005
- Ahmed J, Braun RD, Dunn R, Linsenmeier RA: Oxygen distribution in the macaque retina. *Invest Ophthalmol Vis Sci* 34:516-521, 1993
- Cohen LH, Noell WK: *Relationships Between Visual Function and Metabolism*. Orlando, FL, Academic Press, 1965
- Ames AI: Energy requirements of CNS cells as related to their function and their vulnerability to ischemia: a commentary based on studies on retina. *Can J Physiol Pharmacol* 70 (Suppl.):S158-S164, 1992
- Reichenbach A, Stolzenburg JU, Eberhardt W, Chao TI, Dettmer D, Hertz L: What do retinal muller (glial) cells do for their neuronal 'small siblings'? *J Chem Neuroanat* 6:201-213, 1993
- Hudson BI, Schmidt AM: RAGE: a novel target for drug intervention in diabetic vascular disease. *Pharm Res* 21:1079-1086, 2004
- Stitt AW: The role of advanced glycation in the pathogenesis of diabetic retinopathy. *Exp Mol Pathol* 75:95-108, 2003
- Barile GR, Pachydaki SI, Tari SR, Lee SE, Donmoyer CM, Ma W, Rong LL, Buciarelli LG, Wendt T, Horig H, Hudson BI, Qu W, Weinberg AD, Yan SF, Schmidt AM: The RAGE axis in early diabetic retinopathy. *Invest Ophthalmol Vis Sci* 46:2916-2924, 2005
- Aiello LP: The potential role of PKC beta in diabetic retinopathy and macular edema. *Surv Ophthalmol* 47:S263-S269, 2002
- Caldwell RB, Bartoli M, Behzadian MA, El-Remessy AE, Al-Shabraway M, Platt DH, Liou GI, Caldwell RW: Vascular endothelial growth factor and diabetic retinopathy: role of oxidative stress. *Curr Drug Targets* 6:511-524, 2005
- Antonetti DA, Lieth E, Barber AJ, Gardner TW: Molecular mechanisms of vascular permeability in diabetic retinopathy. *Sem Ophthalmol* 14:240-248, 1999
- Miyamoto K, Khosrof S, Bursell SE, Rohan R, Murata T, Clermont AC, Aiello LP, Ogura Y, Adamis AP: Prevention of leukostasis and vascular leakage in streptozotocin-induced diabetic retinopathy via intercellular adhesion molecule-1 inhibition. *Proc Natl Acad Sci U S A* 96:10836-10841, 1999
- The PKC-DRS Study Group: The effect of ruboxistaurin on visual loss in patients with moderately severe to very severe nonproliferative diabetic retinopathy: initial results of the Protein Kinase C β inhibitor Diabetic Retinopathy Study (PKC-DRS) multicenter randomized clinical trial. *Diabetes* 54:2188-2197, 2005
- Wolter J: Diabetic retinopathy. *Am J Ophthalmol* 51:1123-1139, 1961
- Bloodworth JMB: Diabetic retinopathy. *Diabetes* 2:1-22, 1962
- Parisi V, Uccioli L: Visual electrophysiological responses in persons with type 1 diabetes. *Diabetes Metab Res Rev* 17:12-18, 2001
- Ghirlanda G, Di Leo MA, Caputo S, Cercone S, Greco AV: From functional to microvascular abnormalities in early diabetic retinopathy. *Diabetes Metab Res Rev* 13:15-35, 1997
- Bresnick GH: Diabetic retinopathy viewed as a neurosensory disorder. *Arch Ophthalmol* 104:989-990, 1986
- Greenstein VC, Shapiro A, Zaidi Q, Hood DC: Psychophysical evidence for post-receptor sensitivity loss in diabetics. *Invest Ophthalmol Vis Sci* 33:2781-2790, 1992
- Bearse MA Jr, Han Y, Schneck ME, Barez S, Jacobsen C, Adams AJ: Local multifocal oscillatory potential abnormalities in diabetes and early diabetic retinopathy. *Invest Ophthalmol Vis Sci* 45:3259-3265, 2004
- Bresnick GH, Palta M: Predicting progression to severe proliferative diabetic retinopathy. *Arch Ophthalmol* 105:810-814, 1987
- Han Y, Adams AJ, Bearse MA Jr, Schneck ME: Multifocal electroretinogram and short-wavelength automated perimetry measures in diabetic eyes with little or no retinopathy. *Arch Ophthalmol* 122:1809-1815, 2004
- Realini T, Lai MQ, Barber L: Impact of diabetes on glaucoma screening using frequency-doubling perimetry. *Ophthalmology* 111:2133-2136, 2004
- Bengtsson B, Heijl A, Agardh E: Visual fields correlate better than visual acuity to severity of diabetic retinopathy. *Diabetologia* 48:2494-2500, 2005
- Abu El-Asrar AM, Dralands L, Missotten L, Al-Jadaan IA, Geboes K: Expression of apoptosis markers in retinas of human subjects with diabetes. *Invest Ophthalmol Vis Sci* 45:2760-2766, 2004
- Barber AJ, Lieth E, Khin SA, Antonetti DA, Buchanan AG, Gardner TW: Neural apoptosis in the retina during experimental and human diabetes: early onset and effect of insulin. *J Clin Invest* 102:783-791, 1998
- Park SH, Park JW, Park SJ, Kim KY, Chung JW, Chun MH, Oh SJ: Apoptotic death of photoreceptors in the streptozotocin-induced diabetic rat retina. *Diabetologia* 46:1260-1268, 2003
- Mizutani M, Gerhardinger C, Lorenzi M: Muller cell changes in human diabetic retinopathy. *Diabetes* 47:445-449, 1998
- Lieth E, Barber AJ, Xu B, Dice C, Ratz MJ, Tanase D, Strother JM, the Penn State Retina Research Group: Glial reactivity and impaired glutamate metabolism in short-term experimental diabetic retinopathy. *Diabetes* 47:815-820, 1998
- Barber AJ, Antonetti DA, Gardner TW: Altered expression of retinal occludin and glial fibrillary acidic protein in experimental diabetes: the Penn State Retina Research Group. *Invest Ophthalmol Vis Sci* 41:3561-3568, 2000
- Lieth E, LaNoue KF, Antonetti DA, Ratz M, the Penn State Retina Research Group: Diabetes reduces glutamate oxidation and glutamine synthesis in the retina. *Exp Eye Res* 70:723-730, 2000
- Puro DG: Diabetes-induced dysfunction of retinal Muller cells. *Trans Am Ophthalmol Soc* 100:339-352, 2002
- Li Q, Puro DG: Diabetes-induced dysfunction of the glutamate transporter in retinal Muller cells. *Invest Ophthalmol Vis Sci* 43:3109-3116, 2002
- Rungger-Brandle E, Dosso AA, Leuenberger PM: Glial reactivity, an early feature of diabetic retinopathy. *Invest Ophthalmol Vis Sci* 41:1971-1980, 2000
- Zeng XX, Ng YK, Ling EA: Neuronal and microglial response in the retina of streptozotocin-induced diabetic rats. *Visual Neurosci* 17:463-471, 2000
- Lorenzi M, Gerhardinger C: Early cellular and molecular changes induced by diabetes in the retina. *Diabetologia* 44:791-804, 2001
- Layton CJ, Chidlow G, Casson RJ, Wood JP, Graham M, Osborne NN: Monocarboxylate transporter expression remains unchanged during the

- development of diabetic retinal neuropathy in the rat. *Invest Ophthalmol Vis Sci* 46:2878–2885, 2005
57. Antonetti DA, Barber AJ, LaNoue KF, Nakamura M: New insights into the pathophysiology of diabetic retinopathy: potential cell-specific therapeutic targets. *Diabetes Technol Ther* 2:601–608, 2001
 58. Barber AJ: A new view of diabetic retinopathy: a neurodegenerative disease of the eye. *Prog Neuropsychopharmacol Biol Psychiatry* 27:283–290, 2003
 59. Wellen KE, Hotamisligil GS: Inflammation, stress, and diabetes. *J Clin Invest* 115:1111–1119, 2005
 60. Dana GW: Type of diabetes mellitus associated with diabetic retinitis. *AMA Arch Ophthalmol* 50:123–124, 1953
 61. Cotran RS, Kumar V, Collins T: *Robbins Pathologic Basis of Disease*. Philadelphia, Saunders, 1999
 62. Harrison JK, Jiang Y, Chen S, Xia Y, Maciejewski D, McNamara RK, Streit WJ, Salafra MN, Adhikari S, Thompson DA, Botti P, Bacon KB, Feng L: Role for neuronally derived fractalkine in mediating interactions between neurons and CX3CR1-expressing microglia. *Proc Natl Acad Sci U S A* 95:10896–10901, 1998
 63. Dick AD, Carter D, Robertson M, Broderick C, Hughes E, Forrester JV, Liversidge J: Control of myeloid activity during retinal inflammation. *J Leukoc Biol* 74:161–166, 2003
 64. Schroder S, Palinski W, Schmid-Schonbein GW: Activated monocytes and granulocytes, capillary nonperfusion, and neovascularization in diabetic retinopathy. *Am J Pathol* 139:81–100, 1991
 65. Adamis AP: Is diabetic retinopathy an inflammatory disease? *Br J Ophthalmol* 86:363–365, 2002
 66. Caicedo A, Espinosa-Heidmann DG, Pina Y, Hernandez EP, Cousins SW: Blood-derived macrophages infiltrate the retina and activate Muller glial cells under experimental choroidal neovascularization. *Exp Eye Res* 81:38–47, 2005
 67. Zhang J, Gerhardinger C, Lorenzi M: Early complement activation and decreased levels of glycosylphosphatidylinositol-anchored complement inhibitors in human and experimental diabetic retinopathy. *Diabetes* 51:3499–3504, 2002
 68. Gerhardinger C, Costa MB, Coulombe MC, Toth I, Hoehn T, Grosu P: Expression of acute-phase response proteins in retinal muller cells in diabetes. *Invest Ophthalmol Vis Sci* 46:349–357, 2005
 69. Joussen AM, Poulaki V, Mitsiades N, Cai WY, Suzuma I, Pak J, Ju ST, Rook SL, Esser P, Mitsiades CS, Kirchhof B, Adamis AP, Aiello LP: Suppression of Fas-FasL-induced endothelial cell apoptosis prevents diabetic blood-retinal barrier breakdown in a model of streptozotocin-induced diabetes. *FASEB J* 17:76–78, 2003
 70. Cusick M, Chew EY, Chan CC, Kruth HS, Murphy RP, Ferris FL 3rd: Histopathology and regression of retinal hard exudates in diabetic retinopathy after reduction of elevated serum lipid levels. *Ophthalmology* 110:2126–2133, 2003
 71. Gariano RF, Gardner TW: Retinal angiogenesis in development and disease. *Nature* 438:960–966, 2005
 72. Seigel GM, Chiu L, Paxhia A: Inhibition of neuroretinal cell death by insulin-like growth factor-1 and its analogs. *Mol Vis* 6:157–163, 2000
 73. Barber AJ, Nakamura M, Wolpert EB, Reiter CEN, Seigel GM, Antonetti DA, Gardner TW: Insulin rescues retinal neurons from apoptosis by a phosphatidylinositol 3-kinase/Akt-mediated mechanism that reduces the activation of caspase-3. *J Biol Chem* 276:32814–32821, 2001
 74. Zhang L, Nair A, Krady K, Corpe C, Bonneau RH, Simpson IA, Vannucci SJ: Estrogen stimulates microglia and brain recovery from hypoxia-ischemia in normoglycemic but not diabetic female mice. *J Clin Invest* 113:85–95, 2004
 75. Greenberg DA, Jin K: From angiogenesis to neuropathology. *Nature* 438:954–959, 2005
 76. Kern TS, Engerman RL: A mouse model of diabetic retinopathy. *Arch Ophthalmol* 114:986–990, 1996
 77. Frank RN: The galactosemic dog: a valid model for both early and late stages of diabetic retinopathy (Letter). *Arch Ophthalmol* 113:275–276, 1995
 78. Klein R, Klein BE: Relation of glycemic control to diabetic complications and health outcomes. *Diabetes Care* 21 (Suppl. 3):C39–C43, 1998
 79. Diabetes Control and Complications Trial Research Group: The effect of intensive treatment of diabetes on the development and progression of long-term complications in insulin-dependent diabetes mellitus. *N Engl J Med* 329:977–986, 1993
 80. Stratton IM, Kohner EM, Aldington SJ, Turner RC, Holman RR, Manley SE, Matthews DR: UKPDS 50: risk factors for incidence and progression of retinopathy in type II diabetes over 6 years from diagnosis. *Diabetologia* 44:156–163, 2001
 81. The Diabetes Control and Complications Trial Research Group: The relationship of glycemic exposure (HbA_{1c}) to the risk of development and progression of retinopathy in the Diabetes Control and Complications Trial. *Diabetes* 44:968–983, 1995
 82. The Diabetes Control and Complications Trial Research Group: Implementation of treatment protocols in the Diabetes Control and Complications Trial. *Diabetes Care* 18:361–376, 1995
 83. Reiter CEN, Gardner TW: Retinal insulin and insulin signaling: implications for diabetic retinopathy. *Prog Ret Eye Res* 22:545–562, 2003
 84. Zhang L, Krzentowski G, Albert A, Lefebvre PJ: Risk of developing retinopathy in Diabetes Control and Complications Trial type 1 diabetic patients with good or poor metabolic control. *Diabetes Care* 24:1275–1279, 2001
 85. Chaturvedi N, Sjoelie AK, Porta M, Aldington SJ, Fuller JH, Songini M, Kohner EM: Markers of insulin resistance are strong risk factors for retinopathy incidence in type 1 diabetes: the EURODIAB Prospective Complications Study. *Diabetes Care* 24:284–289, 2001
 86. Stuhldreher WL, Becker DJ, Drash AL, Ellis D, Kuller LH, Wolfson SK, Orchard TJ: The association of waist/hip ratio with diabetes complications in an adult IDDM population. *J Clin Epidemiol* 47:447–456, 1994
 87. Kohner EM, Aldington SJ, Stratton IM, Manley SE, Holman RR, Matthews DR, Turner RC: United Kingdom Prospective Diabetes Study, 30: diabetic retinopathy at diagnosis of non-insulin-dependent diabetes mellitus and associated risk factors. *Arch Ophthalmol* 116:297–303, 1998
 88. Maneschi F, Mashiter K, Kohner EM: Insulin resistance and insulin deficiency in diabetic retinopathy of non-insulin-dependent diabetes. *Diabetes* 32:82–87, 1983
 89. Reiter CE, Sandrasegarane L, Wolpert EB, Klinger M, Simpson IA, Barber AJ, Antonetti DA, Kester M, Gardner TW: Characterization of insulin signaling in rat retina in vivo and ex vivo. *Am J Physiol* 285:E763–E774, 2003
 90. Reiter CEN, Wu X, Sandrasegarane L, Nakamura M, Gilbert KA, Singh RSJ, Fort PE, Antonetti DA, Gardner TW: Diabetes induces basal retinal insulin receptor signaling: reversal with systemic and local insulin. *Diabetes* 55:1148–1156, 2006
 91. Kondo T, Kahn CR: Altered insulin signaling in retinal tissue in diabetic states. *J Biol Chem* 279:37997–38006, 2004
 92. Barber AJ, Antonetti DA, Kern TS, Reiter CEN, Soans RS, Krady JK, Levison SW, Gardner TW, Bronson SK: The Ins^{2AKita} mouse as a model of early retinal complications in diabetes. *Invest Ophthalmol Vis Sci* 46:2210–2218, 2005
 93. Nakamura M, Barber AJ, Antonetti DA, LaNoue KF, Robinson KA, Buse MG, Gardner TW: Excessive hexosamines block the neuroprotective effect of insulin and induces apoptosis in retinal neurons. *J Biol Chem* 276:43748–43755, 2001
 94. Plum L, Schubert M, Bruning JC: The role of insulin receptor signaling in the brain. *Trends Endocrinol Metab* 16:59–65, 2005
 95. Yi X, Schubert M, Peachey NS, Suzuma K, Burks DJ, Kushner JA, Suzuma I, Cahill C, Flint CL, Dow MA, Leshan RL, King GL, White MF: Insulin receptor substrate 2 is essential for maturation and survival of photoreceptor cells. *J Neurosci* 25:240–248, 2005
 96. Junger MA, Rintelen F, Stocker H, Wasserman JD, Vegh M, Radimerski T, Greenberg ME, Hafen E: The Drosophila forkhead transcription factor FOXO mediates the reduction in cell number associated with reduced insulin signaling. *J Biol* 2:20, 2003
 97. Bateman JM, McNeill H: Temporal control of differentiation by the insulin receptor/tor pathway in Drosophila. *Cell* 119:87–96, 2004
 98. Pimentel B, Sanz C, Varela-Nieto I, Rapp UR, De Pablo F, de La Rosa EJ: c-Raf regulates cell survival and retinal ganglion cell morphogenesis during neurogenesis. *J Neurosci* 20:3254–3262, 2000
 99. Kondo T, Vicent D, Suzuma K, Yanagisawa M, King GL, Holzenberger M, Kahn CR: Knockout of insulin and IGF-1 receptors on vascular endothelial cells protects against retinal neovascularization. *J Clin Invest* 111:1835–1842, 2003
 100. Mizutani M, Kern TS, Lorenzi M: Accelerated death of retinal microvascular cells in human and experimental diabetic retinopathy. *J Clin Invest* 97:2883–2890, 1996
 101. Chihara E: Impairment of protein synthesis in the retinal tissue in diabetic rabbits: secondary reduction of fast axonal transport. *J Neurochem* 37:247–250, 1981
 102. Brussee V, Cunningham FA, Zochodne DW: Direct insulin signaling of neurons reverses diabetic neuropathy. *Diabetes* 53:1824–1830, 2004
 103. Pierson CR, Zhang W, Murakawa Y, Sima AAF: Insulin deficiency rather than hyperglycemia accounts for impaired neurotrophic responses and nerve fiber regeneration in type 1 diabetic neuropathy. *J Neuropathol Exp Neurol* 62:260–271, 2003
 104. Writing Team for the Diabetes Control and Complications Trial/Epidemiology of Diabetes Interventions and Complications Research Group:

- Effect of intensive therapy on the microvascular complications of type 1 diabetes mellitus. *JAMA* 287:2563–2569, 2002
105. Martin CL, Albers J, Herman WH, Cleary P, Waberski B, Greene DA, Stevens MJ, Feldman EL: Neuropathy among the Diabetes Control and Complications Trial cohort 8 years after trial completion. *Diabetes Care* 29:340–344, 2006
 106. Schubert M, Gautam D, Surjo D, Ueki K, Baudler S, Schubert D, Kondo T, Alber J, Galldik N, Kusterman E, Arndt S, Jacobs AH, Krone W, Kahn CR, Bruning JC: Role for neuronal insulin resistance in neurodegenerative diseases. *Proc Natl Acad Sci U S A* 101:3100–3105, 2004
 107. Fisher SJ, Bruning JC, Lannon S, Kahn CR: Insulin signaling in the central nervous system is critical for the normal sympathoadrenal response to hypoglycemia. *Diabetes* 54:1447–1451, 2005
 108. Li ZG, Zhang W, Grunberger G, Sima AA: Hippocampal neuronal apoptosis in type 1 diabetes. *Brain Res* 946:221–231, 2002
 109. Li ZG, Zhang W, Sima AA: The role of impaired insulin/IGF action in primary diabetic encephalopathy. *Brain Res* 1037:12–24, 2005
 110. Cai F, Helke CJ: Abnormal PI3 kinase/Akt signal pathway in vagal afferent neurons and vagus nerve of streptozotocin-diabetic rats. *Brain Res Mol Brain Res* 110:234–244, 2003
 111. Shires TK, Braddock KJ, Pulido JS: Insulin in the vitreous of the normal and streptozotocin-induced diabetic rat. *Peptides* 13:671–675, 1992
 112. Banks WA: The source of cerebral insulin. *Eur J Pharmacol* 490:5–12, 2004
 113. Banks WA, Jaspan JB, Huang W, Kastin AJ: Transport of insulin across the blood-brain barrier: saturability at euglycemic doses of insulin. *Peptides* 18:1423–1429, 1997
 114. Woods SC, Seeley RJ, Baskin DG, Schwartz MW: Insulin and the blood-brain barrier. *Curr Pharm Des* 9:795–800, 2003
 115. Linner E, Svanborg A, Zelander T: Prediabetic retinal and renal lesions without obvious disturbances in glucose metabolism in a patient with family history of diabetes. *Am J Med* 39:298–304, 1965
 116. Collens WS, Silverstein JN, Dobkin GB: Case of a diabetic with a Kimmelstiel-Wilson syndrome and a normal glucose tolerance. *Ann Intern Med* 50:1282–1288, 1959
 117. Harrower ADB, Clark BF: Diabetic retinopathy with normal glucose tolerance. *Br J Ophthalmol* 60:459–463, 1976
 118. Barnes AJ, Kohner EM, Johnston DG, Alberti KGMM: Severe retinopathy and mild carbohydrate intolerance: possible role of insulin deficiency and elevated circulating growth hormone. *Lancet* 1:1465–1468, 1985
 119. Chan JY, Cole E, Hanna AK: Diabetic nephropathy and proliferative retinopathy with normal glucose tolerance. *Diabetes Care* 8:385–390, 1985
 120. Hutton W: Retinal microangiopathy without associated glucose intolerance. In *Diabetic Retinopathy*. Lynn JR, Snyder WB, Vaiser A, Eds. New York, Grune & Stratton, 1974
 121. Johansen K: Diabetic angiopathy in a patient with normal glucose tolerance and plasma insulin response. *Am J Med* 47:487–491, 1969
 122. Varkonyi TT, Peto T, Degi R, Keresztes K, Lengyel C, Kempler P, Lonovics J: Impairment of visual evoked potentials: an early central manifestation of diabetic neuropathy? (Letter). *Diabetes Care* 25:1661–1662, 2002
 123. Ozdek S, Lonneville YH, Onol M, Yetkin I, Hasanreisoglu BB: Assessment of nerve fiber layer in diabetic patients with scanning laser polarimetry. *Eye* 16:761–765, 2002
 124. Chihara E, Matsuoka T, Ogura Y, Matsumura M: Retinal nerve fiber layer defect as an early manifestation of diabetic retinopathy. *Ophthalmology* 100:1147–1151, 1993
 125. Han Y, Bearse MA Jr, Schneck ME, Barez S, Jacobsen CH, Adams AJ: Multifocal electroretinogram delays predict sites of subsequent diabetic retinopathy. *Invest Ophthalmol Vis Sci* 45:948–954, 2004
 126. Hovind P, Tarnow L, Rossing K, Rossing P, Eising S, Larsen N, Binder C, Parving HH: Decreasing incidence of severe diabetic microangiopathy in type 1 diabetes. *Diabetes Care* 26:1258–1264, 2003
 127. Martin PM, Roon P, Van Ells TK, Ganapathy V, Smith SB: Death of retinal neurons in streptozotocin-induced diabetic mice. *Invest Ophthalmol Vis Sci* 45:3330–3336, 2004
 128. Kim SY, Johnson MA, McLeod DS, Alexander T, Otsuji T, Steidl SM, Hansen BC, Luttly GA: Retinopathy in monkeys with spontaneous type 2 diabetes. *Invest Ophthalmol Vis Sci* 45:4543–4553, 2004
 129. Johnson MA, Luttly GA, McLeod DS, Otsuji T, Flower RW, Sandagar G, Alexander T, Steidl SM, Hansen BC: Ocular structure and function in an aged monkey with spontaneous diabetes mellitus. *Exp Eye Res* 80:37–42, 2005
 130. Kim SY, Johnson MA, McLeod DS, Alexander T, Hansen BC, Luttly GA: Neutrophils are associated with capillary closure in spontaneously diabetic monkey retinas. *Diabetes* 54:1534–1542, 2005
 131. Buchi ER, Kurosawa A, Tso MO: Retinopathy in diabetic hypertensive monkeys: a pathologic study. *Graefes Arch Clin Exp Ophthalmol* 234:388–398, 1996
 132. Penn State College of Medicine: *Workshop on Mechanisms of Vision Impairment in Diabetic Retinopathy*. Hershey, PA, Penn State College of Medicine, 2003
 133. Giannarelli R, Coppelli A, Sartini M, Aragona M, Boggi U, Vistoli F, Rizzo G, Prato SD, Mosca F, Marchetti P: Effects of pancreas-kidney transplantation on diabetic retinopathy. *Transpl Int* 18:619–622, 2005
 134. Cryer PE: Hypoglycemia is the limiting factor in the management of diabetes. *Diabetes Metab Res Rev* 15:42–46, 1999
 135. Gardner TW, Aiello LP: Pathogenesis of diabetic retinopathy. In *Diabetes and Ocular Disease: Past, Present, and Future Therapies*. Flynn HF, Smiddy W, Eds. San Francisco, CA, American Academy of Ophthalmology, 2000, p. 1–17
 136. Magistretti PJ, Pellerin L, Rothman DL, Shulman RG: Energy on demand. *Science* 283:496–497, 1999
 137. Aiello LP, Gardner TW: Proliferative diabetic retinopathy: the growth factor paradigm. In *Diabetic Retinopathy*. Flynn HF, Smiddy W, Eds. San Francisco, CA, American Academy of Ophthalmology, 2000, p. 239–273