

Common Variants in 40 Genes Assessed for Diabetes Incidence and Response to Metformin and Lifestyle Intervention in the Diabetes Prevention Program

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OBJECTIVE—Genome-wide association studies have begun to elucidate the genetic architecture of type 2 diabetes. We examined whether single nucleotide polymorphisms (SNPs) identified through targeted complementary approaches affect diabetes incidence in the at-risk population of the Diabetes Prevention Program (DPP) and whether they influence a response to preventive interventions.

RESEARCH DESIGN AND METHODS—We selected SNPs identified by prior genome-wide association studies for type 2 diabetes and related traits, or capturing common variation in 40 candidate genes previously associated with type 2 diabetes, implicated in monogenic diabetes, encoding type 2 diabetes drug targets or drug-metabolizing/transporting enzymes, or involved in relevant physiological processes. We analyzed 1,590 SNPs for association with incident diabetes and their interaction with response to metformin or lifestyle interventions in 2,994 DPP participants. We controlled for multiple hypothesis testing by assessing false discovery rates.

RESULTS—We replicated the association of variants in the metformin transporter gene *SLC47A1* with metformin response and detected nominal interactions in the AMP kinase (AMPK)

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gene *STK11*, the AMPK subunit genes *PRKAA1* and *PRKAA2*, and a missense SNP in *SLC22A1*, which encodes another metformin transporter. The most significant association with diabetes incidence occurred in the AMPK subunit gene *PRKAG2* (hazard ratio 1.24, 95% CI 1.09–1.40, $P = 7 \times 10^{-4}$). Overall, there were nominal associations with diabetes incidence at 85 SNPs and nominal interactions with the metformin and lifestyle interventions at 91 and 69 mostly nonoverlapping SNPs, respectively. The lowest P values were consistent with experiment-wide 33% false discovery rates.

CONCLUSIONS—We have identified potential genetic determinants of metformin response. These results merit confirmation in independent samples. *Diabetes* 59:2672–2681, 2010

The number of common genetic variants reproducibly associated with type 2 diabetes is growing (1). Well-powered candidate gene association studies and, more recently, genome-wide association studies (GWASs) have identified over two dozen loci robustly and reproducibly associated with type 2 diabetes or related quantitative glycemic traits. While these discoveries have advanced our understanding of the genetics of type 2 diabetes, they only explain a small fraction of the overall genetic contribution to the disease. Furthermore, in most cases, the genes involved in type 2 diabetes risk have not yet been identified: the majority of the associations detected thus far merely mark genomic regions where a certain variant is overrepresented in diseased cases versus unaffected controls, and subsequent fine-mapping and functional studies are necessary before a molecular mechanism can be ascribed to each locus.

Nevertheless, progress in the translation of genetic discoveries to clinical practice can advance along parallel paths. On the one hand, knowledge of the specific gene or variant causing the molecular phenotype is not needed to determine whether the associated region can aid prediction or affect the response to therapy; and on the other hand, targeted approaches can be applied in vivo in humans that shed light on the function of genes of interest, as a way to narrow the regions to study. These two objectives can be achieved by controlled interventions in randomized clinical trials. Such pharmacogenetic or gene-environmental strategies can fulfill both complementary roles, testing whether genetic variants predict response to therapy and whether a particular pharmacologic or lifestyle intervention affects the mode of action of specific risk loci.

In type 2 diabetes, the field of pharmacogenetics re-

TABLE 1
Baseline characteristics of the DPP participants enrolled in the genetic study

	Overall	Placebo	Metformin	Lifestyle
<i>n</i>	2,994	1,000	990	1,004
Sex				
Male	982 (32.8)	311 (31.1)	344 (34.7)	327 (32.6)
Female	2,012 (67.2)	689 (68.9)	646 (65.3)	677 (67.4)
Race or ethnic group				
White	1,674 (55.9)	557 (55.7)	570 (57.6)	547 (54.5)
African American	612 (20.4)	210 (21.0)	206 (20.8)	196 (19.5)
Hispanic	498 (16.6)	164 (16.4)	158 (16.0)	176 (17.5)
Asian/Pacific Islander	128 (4.3)	39 (3.9)	33 (3.3)	56 (5.6)
American Indian	82 (2.7)	30 (3.0)	23 (2.3)	29 (2.9)
Family history of diabetes	2,089 (69.8)	703 (70.4)	683 (69.0)	703 (70.1)
History of gestational diabetes mellitus	321 (16.0)	105 (15.2)	104 (16.1)	112 (16.5)
Quantitative traits				
Age (years)	50.8 ± 10.7	50.5 ± 10.5	51.0 ± 10.4	50.7 ± 11.4
Weight (kg)	94.6 ± 20.3	94.8 ± 20.2	94.6 ± 20.0	94.5 ± 20.8
BMI (kg/m ²)	34.1 ± 6.7	34.3 ± 6.7	34.0 ± 6.7	34.0 ± 6.8
Waist circumference (cm)	105.2 ± 14.6	105.3 ± 14.5	105.0 ± 14.6	105.3 ± 14.9
Waist-to-hip ratio	0.92 ± 0.09	0.92 ± 0.08	0.93 ± 0.09	0.93 ± 0.09
Plasma glucose (mg/dl)				
In the fasting state	106.7 ± 8.2	107.0 ± 8.4	106.7 ± 8.4	106.5 ± 7.9
2 h after an oral glucose load	164.7 ± 17.1	164.6 ± 17.2	165.1 ± 17.2	164.4 ± 16.9
Glycated hemoglobin (%)	5.91 ± 0.51	5.91 ± 0.50	5.91 ± 0.51	5.91 ± 0.51
Leisure physical activity (MET h/week)	16.1 ± 25.9	16.6 ± 29.1	16.4 ± 25.9	15.4 ± 22.4

Data are *n* (%) or means ± SD.

mains in its infancy. Although pharmacogenetic investigation has yielded clinically actionable results in neonatal diabetes and maturity-onset diabetes of the young, extending these studies to common type 2 diabetes has been more arduous (2). With regard to metformin, intriguing results were obtained by Shu et al. (3) in their investigation of common variants in *SLC22A1*, which encodes the liver-specific organic cation transporter 1 responsible for the absorption of metformin into hepatocytes: in a study of 20 human participants, carriers of reduced-function polymorphisms of *SLC22A1* had a 17% higher area under the glucose curve after an oral glucose tolerance test when treated with metformin, indicating decreased responsiveness. Unfortunately, these results have not been confirmed in the long-term follow-up of a large observational cohort of patients treated with metformin monotherapy (4). Recently, a preliminary association was discovered between a variant in *SLC47A1*, which encodes the multidrug and toxin extrusion protein 1 (involved in the excretion of metformin into the bile and urine) and the glucose-lowering effect of metformin (5).

The Diabetes Prevention Program (DPP) can help answer some of these questions (6). The strengths of this randomized clinical trial include its enrollment of participants at high risk of developing diabetes, multiethnic composition, comprehensive longitudinal measures, and standardized behavioral and pharmacologic interventions. Extensive in-depth phenotyping and the use of behavioral and pharmacologic interventions allow characterization of the effects of known type 2 diabetes variants on diabetes incidence and response to therapy. We therefore designed a large-scale genotyping study by which we tested 1,590 variants identified through prior genetic studies of type 2 diabetes or related traits, as well as those capturing all common variation in 40 biological candidate genes, for association with diabetes incidence or response to preventive interventions (lifestyle modification or metformin) in the DPP.

RESEARCH DESIGN AND METHODS

The DPP. The DPP was a 27-center randomized clinical trial in the U.S. that assessed whether metformin or lifestyle interventions prevent or delay development of diabetes in high-risk individuals. The DPP enrolled 3,234 overweight or obese people without diabetes but with impaired glucose tolerance and elevated fasting glucose and randomized them to placebo, metformin (850 mg twice daily), or a lifestyle intervention program consisting of individual and group counseling sessions conducted by dietary and exercise professionals aimed at ≥7% weight loss and ≥150 min of physical activity per week. A fourth arm of 585 subjects assigned to troglitazone (400 mg daily) was stopped because of hepatotoxicity (7). The primary end point was development of diabetes, ascertained by semi-annual measurement of fasting glucose or an annual 75-g oral glucose tolerance test, either of which was confirmed on a second occasion. The metformin and lifestyle interventions reduced the incidence of diabetes by 31% (95% CI 17–43) and 58% (95% CI 48–66), respectively, versus placebo (6). The 2,994 participants in the placebo, metformin, and lifestyle arms who gave informed consent for genetic investigation are the subjects of this study, which was approved by institutional review boards at each of the 27 participating sites. Their demographic characteristics are shown in Table 1.

SNP selection. We selected SNPs in two ways: 1) SNPs in high-likelihood candidate genes and 2) SNPs identified by ongoing GWASs for type 2 diabetes or related metabolic traits. The 40 candidate genes were tentatively associated with type 2 diabetes, implicated in monogenic forms of diabetes, known to encode type 2 diabetes drug targets or drug-metabolizing/transporting enzymes, or involved in cellular metabolism, hormonal regulation, or response to exercise (Table 2). We used *Tagger* (8) to capture (at $r^2 \geq 0.8$) all common (minor allele frequency >5%) variations in European (CEU) and African (YRI) HapMap populations in these candidate genes. For seven additional genes (*ACE*, *CASQ1*, *GCKR*, *IRS1*, *KCNQ1*, *LIPC*, and *NOS3*), rather than attempting full coverage of genetic variation, we selected a limited number of SNPs previously associated with the phenotypes of interest. As the study evolved, it became obvious that previous reports of genetic association provided an equally compelling—or perhaps even higher—prior probability of true association with type 2 diabetes traits than biological function alone; thus, we also focused on GWASs whose results were available at the time this customized genotyping array was designed: SNPs associated with type 2 diabetes in the Diabetes Genetics Initiative (9), DIAGRAM (10), or three smaller 100K SNP GWASs in which we participated (11–13); SNPs tentatively associated with quantitative glycemic traits (fasting glucose, the insulinogenic index, and insulin resistance by homeostasis model assessment) in the Diabetes Genetics Initiative; or SNPs associated with obesity (14,15) or lipid traits (16–18). For quality control and analytical reasons, we also included some SNPs previously genotyped in these samples, as well as

TABLE 2
Number of SNPs analyzed per selection category

Category	<i>n</i>
Candidate genes (<i>n</i> SNPs)	1,256
Comprehensive coverage	1,241
Monogenic diabetes	317
<i>ABCC8</i> (84), <i>GCK</i> (37), <i>HNF1A</i> (20), <i>HNF1B</i> (74), <i>HNF4A</i> (66), <i>KCNJ11</i> (9), <i>NEUROD1</i> (14), <i>PDX1</i> (13)	
Monogenic obesity	21
<i>MC4R</i> (21)	
Previously associated with type 2 diabetes	60
<i>CAPN10</i> (33), <i>PTPN1</i> (27)	
Drug targets	190
<i>PPARG</i> (59), <i>PRKAA1</i> (9), <i>PRKAA2</i> (18), <i>PRKABI</i> (10), <i>PRKAB2</i> (13), <i>PRKAG1</i> (7), <i>PRKAG2</i> (53), <i>PRKAG3</i> (11), <i>STK11</i> (10)	
Drug-metabolizing enzymes and transporters	135
<i>CYP3A4</i> (15), <i>SLC22A1</i> (47), <i>SLC22A2</i> (44), <i>SLC47A1</i> (29)	
Hormonal regulation	117
<i>ADIPOQ</i> (25), <i>ADIPOR1</i> (22), <i>ADIPOR2</i> (31), <i>GCG</i> (13), <i>ITLN1</i> (11), <i>ITLN2</i> (15)	
Cellular energy	334
<i>FOXO1A</i> (35), <i>PCK1</i> (37), <i>PCK2</i> (14), <i>PKLR</i> (10), <i>PPARA</i> (63), <i>PPARGC1A</i> (79), <i>PPARGC1B</i> (96)	
Response to exercise	67
<i>CREB1</i> (13), <i>MEF2A</i> (38), <i>MEF2D</i> (16)	
Select SNPs in candidate genes	15
<i>ACE</i> (1), <i>CASQ1</i> (2), <i>GCKR</i> (2), <i>IRS1</i> (2), <i>KCNQ1</i> (1), <i>LIPC</i> (1), <i>NOS3</i> (6)	
Ancillary studies	26
<i>ALOX5</i> (15), <i>IL6</i> (1), renin-angiotensin system (7), <i>USF1</i> (2), <i>TNF</i> (1)	
GWASs	248
Amish 100K GWAS (13): Type 2 diabetes	46
Framingham Heart Study 100K GWAS (11): Type 2 diabetes	38
Pima 100K GWAS (12): Type 2 diabetes	12
Obesity (14,15)	11
Diabetes Genetics Initiative (9)	
Fasting glucose	37
Insulin resistance by homeostasis model assessment	16
Insulinogenic index	17
Type 2 diabetes	30
DIAGRAM (10): Type 2 diabetes	6
Lipids (16–18)	35
Quality control	60
African American ancestry informative markers (19)	29
Hispanic ancestry informative markers (20)	29
Previously genotyped in DPP	2
Total	1,590

ancestry-informative markers to derive a global proportion of geographic ancestry in African American (19) or Hispanic (20) participants. Finally, we included a small number of SNPs provided by investigators leading ancillary studies approved by the DPP ancillary studies and genetics subcommittees. The total number of SNPs analyzed for each category is shown in Table 2.

Genotyping. We initially designed a 1,536-SNP oligonucleotide pool array for the Illumina BeadArray platform (Illumina, San Diego, CA). In the 1,445 SNPs that passed quality control metrics, the sample pass rate was 99.8% and the average genotyping call rate per SNP was 98.5%. Because 91 SNPs failed genotyping on the oligonucleotide pool array, we assessed the adequacy of the coverage afforded by the successfully genotyped SNPs in each region. To rescue relevant SNPs, we used linkage disequilibrium (LD) to select proxy SNPs highly correlated to those that had failed and genotyped them on a Sequenom iPLEX platform. After quality control, 1,590 SNPs were available for analysis.

Statistical analysis. We tested the effect of each SNP on diabetes incidence under an additive genetic model by Cox proportional hazards models, using age, sex, ethnicity, and treatment arm as covariates and including treatment (metformin or lifestyle) \times genotype interaction terms. In secondary analyses, we stratified participants by treatment arm; if the interaction *P* value was nominally significant, only stratified analyses were considered. We used the MACH software (21) and the HapMap CEU population to impute allelic calls at SNPs not directly genotyped in the DPP. Because of concerns regarding the accuracy of imputation methods in admixed populations, we restricted this procedure to individuals of self-described non-Hispanic white ethnicity. Genotype-phenotype correlations on imputed data were considered confirmatory of prior associations, as well as an initial fine-mapping exploration. Using the program STRUCTURE (22), we applied these markers trained on the HapMap populations to assign a proportion of global European ancestry to each DPP participant.

We considered two sequential approaches to correct for multiple hypothesis testing based on the number of SNPs examined (23). We first ran 1,000 permutations in which diabetes outcome was randomly assigned to an individual's genotype within each ethnicity and treatment group (keeping sex and age together with genotype, and BMI with diabetes outcome). The *P* value for the overall null hypothesis is the fraction of permutations ($n/1,000$) for which the scalar statistic is at least as extreme as that observed for the data (24). To estimate the expected proportion of type I errors among the rejected hypotheses, we also computed false discovery rates (FDRs) as in Benjamini and Hochberg (25).

RESULTS

Supplementary Table 1 (available in an online appendix at <http://diabetes.diabetesjournals.org/cgi/content/full/db10-0543/DC1>) shows that we achieved adequate coverage of all 40 genes in the two targeted populations, with 37 genes reaching at least 80% of common variants captured at $r^2 \geq 0.8$ in Europeans and all 40 reaching at least 70% of common variants captured at that level (comparable numbers were obtained in Africans). The average proportion of European ancestry among the DPP self-described white participants, as determined by ancestry-informative markers, was 98.9%, and the average proportion of West-African ancestry among DPP self-described African American participants was 89.3%. Given these results, we used self-described ethnicity as a covariate for these analyses. The full set of results is available in supplementary Table 2.

Table 3 shows the candidate gene regions harboring variants nominally associated with diabetes incidence in the treatment-adjusted models for the full study (i.e., there was no evidence for interaction with either intervention); only the top SNP within each gene region (out of 85 nominal associations) is given. The most significant associations occurred at SNPs in the AMP kinase (AMPK) subunit gene *PRKAG2* (hazard ratio [HR] 1.24, 95% CI 1.09–1.40, $P = 7.0 \times 10^{-4}$ for the top SNP rs5017427, which is consistent with an experiment-wide 34% FDR). Twelve other *PRKAG2* SNPs were nominally associated with diabetes (five in the top ten). Although most of them are in moderate to high LD with the index SNP (r^2 ranging from 0.49 to 1.0 in HapMap CEU), at least two of them (rs954482 and rs2727537) are only weakly correlated with rs5017427 (r^2 0.07 and 0.05, respectively). Nevertheless, the consistency of the association signal in this region provides reassurance with regard to the absence of genotyping artifacts in our dataset. Of SNPs previously associated with type 2 diabetes in the 100K Amish, Framingham, or Pima GWASs, three (rs1422930 in *ODZ2*, rs1859441 near *COL2A1* and *SENPI1*, and rs385909 near *SH3YL1*) had consistent nominal associations with diabetes incidence in the DPP, and two had nominally significant associations (rs10520926 and rs3136279) in the opposite direction. On the other hand, none of the six SNPs selected from the

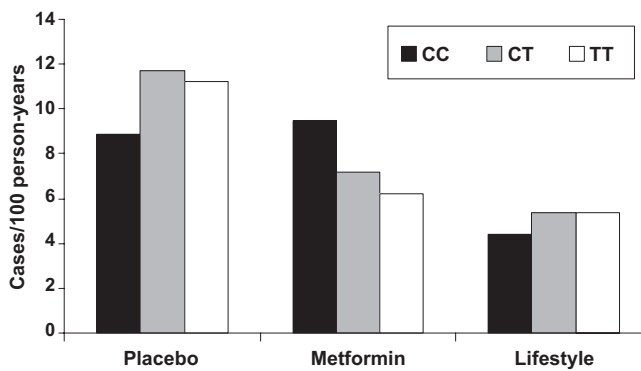


FIG. 1. Diabetes incidence in the DPP, by genotype at rs8065082 in the *SLC47A1* gene. This SNP is in tight LD with rs2289669 ($r^2 \sim 0.8$), whose major allele predicts a poorer response to metformin (5). In the DPP, major allele homozygotes at rs8065082 did not benefit from metformin with regard to diabetes prevention, whereas minor allele carriers did ($P < 0.001$).

DIAGRAM meta-analysis (original odds ratio [OR] ranging from 1.05 to 1.15) were nominally significant in the DPP. Fifteen SNPs in genes that cause either maturity-onset diabetes of the young or neonatal diabetes were nominally associated with diabetes incidence; one of them, rs11868513 in *HNF1B* (not in LD with the previously type 2 diabetes-associated SNP rs757210), was strongly associated with diabetes incidence in the placebo arm (HR 1.69, 95% CI 1.36–2.10, $P = 2 \times 10^{-6}$). Finally, 14 SNPs in genes that encode metformin transporters (*SLC22A1*, *SLC22A2*, and *SLC47A1*) were nominally associated with diabetes incidence. Of the 85 nominal associations with diabetes incidence in DPP, only two SNPs (rs651164 in *SLC22A1* and rs3736265 in *PPARGC1A*) were nominally associated with type 2 diabetes in DIAGRAM in a consistent direction (OR 1.08, 95% CI 1.02–1.16, $P = 0.01$, and OR 1.15, 95% CI 1.01–1.31, $P = 0.04$, respectively), with 60 other SNPs not being nominally significantly associated in DIAGRAM and 23 SNPs not captured in that dataset.

Table 4 shows the candidate gene regions harboring variants that have a nominally significant genotype \times metformin interaction; only the top SNP within each gene region is given (out of 91 nominal associations). The best result was consistent with a study-wide 33% FDR. At rs8065082 in *SLC47A1*, there was a nominal interaction with metformin ($P = 0.006$), with the minor allele associated with lower diabetes incidence in the metformin arm (HR 0.78, 95% CI 0.64–0.96, $P = 0.02$) but not in the placebo arm (1.15, 0.97–1.37, $P = 0.11$). At this locus, major allele homozygotes did not benefit from metformin with regard to diabetes prevention (HR 1.07, 95% CI 0.77–1.50, vs. placebo, $P = 0.68$), whereas minor allele carriers did (0.58, 0.46–0.73, vs. placebo, $P < 0.001$; Fig. 1). We also noted a nominally significant interaction of a missense SNP in *SLC22A1* (rs683369, encoding L160F) with metformin, with the major allele protecting from diabetes in the metformin arm (HR 0.69, 95% CI 0.53–0.89, $P = 0.004$) but not the placebo arm (1.01, 0.79–1.30, $P = 0.91$); the major allele is therefore associated with 31% risk reduction in diabetes incidence but only under the action of metformin. In this arm, the likelihood of developing diabetes depended on the number of phenylalanine alleles (HR 0.72, 95% CI 0.59–0.88, vs. placebo for LL homozygotes; 0.92, 0.66–1.28, for heterozygotes; and

1.44, 0.56–3.67, for FF homozygotes). There were five nominally significant interactions at SNPs encoding putative drug targets for metformin, in the gene encoding the AMPK kinase *STK11* and the AMPK subunit genes *PRKAA1*, *PRKAA2*, and *PRKAB2*, respectively. A total of 22 SNPs in the *ABCC8-KCNJ11* region also had nominally significant interactions with metformin, including rs5215, which is tightly linked to the widely replicated type 2 diabetes-associated missense SNP rs5219 (E23K) in *KCNJ11*.

Table 5 shows the candidate gene regions harboring variants that have a nominally significant interaction with the lifestyle intervention; only the top SNP within each gene region is given (out of 69 nominal associations). The best result was consistent with an experiment-wide 84% FDR. Twelve of the top findings were in four AMPK subunit genes (*PRKAA2*, *PRKAB2*, *PRKAG1*, and *PRKAG2*), and 11 SNPs clustered around the peroxisome proliferator-associated receptor γ coactivators 1 α and 1 β (*PPARGC1A* and *PPARGC1B*, respectively).

Review of 1,609 SNPs imputed in non-Hispanic white DPP participants (supplemental Table 3) revealed the nominal association of other *PRKAG2* SNPs with diabetes incidence (best $P = 5 \times 10^{-5}$). Imputed SNPs in the *PRKAA1*, *PRKAA2*, and *ABCC8-KCNJ11* regions also had nominally significant interactions with metformin.

DISCUSSION

We conducted a large-scale genotyping study in the DPP, with the aim to test whether common variants in candidate genes involved in major spheres of human physiology predict diabetes incidence or response to preventive interventions in a multiethnic at-risk population. Our secondary purpose was to characterize the mechanism of action of previously associated variants. We provide evidence supporting a previously reported association of variants in the metformin transporter gene *SLC47A1* with weaker metformin response, here defined as the reduced ability of metformin to lower diabetes incidence (5). We identified a number of nominal associations with diabetes incidence or metformin response in several compelling candidate genes; however, none stand strict statistical correction for multiple hypothesis testing by FDRs.

Correction for multiple tests requires careful consideration in genetic association studies (26). When large numbers of SNPs are tested, methods that are valid in the presence of correlations due to LD, such as permutation methods or evaluation of FDR, are preferred over those that assume independence of SNPs. The scope of the present analysis is guided by technological convenience, and it might be argued that the number of distinct scientific hypotheses formulated, rather than the physical size of the genotyping array, is most relevant to the interpretation of results. However, what constitutes a single hypothesis (e.g., an SNP, a gene, an entire pathway, or a constellation of phenotypes) is subjective. On the other hand, correcting for the equivalent of the universe of independent common variants in the human genome (empirically estimated at ~ 1 million [27]) is gaining increasing favor among genetic statisticians. In this context, the novel findings reported here should be viewed as hypothesis-generating.

We previously quantified the power of the DPP to detect modest genetic effects on diabetes incidence (28). Assuming there are no gene-treatment interactions, these calcu-

TABLE 3
Candidate gene variants nominally associated with diabetes incidence in the DPP

SNP	Chromosome	Position (NCBI 36)	Gene	Alleles (m/M)	Overall, treatment-adjusted			FDR Q
					HR (95% CI)	Observed P	Permuted P	
rs5017427	7	150886136	<i>PRKAG2</i>	T/C	1.24 (1.09–1.40)	0.0007	0.001	0.34
rs2453583	17	19382628	<i>SLC47A1</i>	T/A	0.81 (0.72–0.92)	0.001	0.003	0.34
rs315978	6	160572848	<i>SLC22A2</i>	T/C	0.73 (0.60–0.89)	0.002	0.002	0.34
rs4273018	15	97910079	<i>MEF2A</i>	T/C	1.20 (1.07–1.35)	0.002	0.005	0.50
rs9551419	13	27378356	<i>PDX1</i>	T/C	0.77 (0.65–0.92)	0.004	0.003	0.34
rs1042531	20	55574386	<i>PCK1</i>	C/A	0.84 (0.74–0.94)	0.004	0.002	0.34
rs1342387	1	201180979	<i>ADIPOR1</i>	A/G	1.17 (1.05–1.31)	0.006	0.009	0.55
rs1388332	4	23438412	<i>PPARGC1A</i>	G/A	1.32 (1.08–1.61)	0.006	0.007	0.53
rs6093976	20	42469194	<i>HNF4A</i>	A/G	0.78 (0.65–0.94)	0.008	0.008	0.53
rs651164	6	160501364	<i>SLC22A1</i>	T/C	1.18 (1.04–1.33)	0.008	0.007	0.53
rs12330015	22	44968942	<i>PPARA</i>	C/T	0.79 (0.66–0.95)	0.01	0.01	0.58
rs2755209	13	40035804	<i>FOXO1</i>	C/A	1.16 (1.04–1.31)	0.01	0.01	0.58
rs10875552	5	149169682	<i>PPARGC1B</i>	C/T	0.84 (0.74–0.96)	0.01	0.008	0.53
rs739690	11	17373454	<i>KCNJ11</i>	C/G	1.87 (1.15–3.05)	0.01	0.01	0.66
rs7811022	7	99223619	<i>CYP3A4</i>	G/C	1.33 (1.06–1.67)	0.01	0.02	0.67
rs832646	2	182266649	<i>NEUROD1</i>	C/T	0.57 (0.35–0.91)	0.02	0.02	0.68
rs11836547	12	1756708	<i>ADIPOR2</i>	C/G	1.37 (1.05–1.78)	0.02	0.02	0.67
rs916829	11	17397049	<i>ABCC8</i>	A/G	0.80 (0.66–0.97)	0.03	0.04	0.78
rs12951345	17	33151976	<i>HNF1B</i>	G/T	1.17 (1.02–1.34)	0.03	0.03	0.73
rs709159	3	12456203	<i>PPARG</i>	C/A	1.16 (1.01–1.33)	0.04	0.04	0.82
rs12058717	1	159108473	<i>ITLN1</i>	T/C	1.28 (1.01–1.63)	0.04	0.049	0.87

SNPs in or near biological candidate genes showing nominal association with diabetes incidence in the DPP are shown. HRs are estimated for the minor allele (m) vs. the major allele (M) under an additive genetic model. Only the top SNP within each gene region is shown; the full set of results (including allele frequencies) is available in supplementary Table 2.

lifestyle modification, and metformin arms have 53, 34, and 44% power, respectively. The DPP has inadequate power for detecting an effect size of <10%. Thus, it is not

TABLE 4
Candidate gene variants showing a nominally significant interaction with the metformin intervention in the DPP

SNP	Chromosome	Position (NCBI 36)	Gene	Alleles (m/M)	Interaction genotype * metformin		FDR Q
					Observed P	Permuted P	
rs11868513	17	33126805	<i>HNF1B</i>	A/G	0.0007	0.001	0.33
rs4148609	11	17441307	<i>ABCC8</i>	A/G	0.002	0.002	0.33
rs11086926	20	42492111	<i>HNF4A</i>	G/T	0.002	0.001	0.33
rs10213440	4	23475437	<i>PPARGC1A</i>	C/T	0.002	0.002	0.33
rs4424892	15	97921908	<i>MEF2A</i>	G/A	0.003	0.006	0.38
rs8065082	17	19405783	<i>SLC47A1</i>	T/C	0.006	0.008	0.41
rs6666307	1	154719358	<i>MEF2D</i>	T/A	0.009	0.005	0.38
rs3792269	2	241180152	<i>CAPN10</i>	G/A	0.01	0.01	0.41
rs758027	12	1662034	<i>ADIPOR2</i>	C/T	0.01	0.002	0.33
rs7124355	11	17369536	<i>KCNJ11</i>	A/G	0.01	0.01	0.41
rs662301	6	160616909	<i>SLC22A2</i>	T/C	0.02	0.02	0.47
rs2908289	7	44190467	<i>GCK</i>	A/G	0.02	0.02	0.47
rs741765	19	1172545	<i>STK11</i>	T/C	0.02	0.02	0.47
rs6701920	1	159181541	<i>ITLN2</i>	A/G	0.02	0.01	0.41
rs741579	5	149165563	<i>PPARGC1B</i>	G/A	0.02	0.01	0.38
rs4810083	20	55553677	<i>PCK1</i>	T/C	0.03	0.03	0.53
rs9803799	1	56952660	<i>PRKAA2</i>	G/T	0.03	0.02	0.49
rs17367421	1	153553865	<i>PKLR</i>	C/G	0.03	0.03	0.53
rs683369	6	160471194	<i>SLC22A1</i>	G/C	0.03	0.03	0.53
rs249429	5	40817996	<i>PRKAA1</i>	C/T	0.04	0.03	0.53
rs6733736	2	162704406	<i>GCG</i>	G/A	0.04	0.01	0.43
rs4253652	22	44947503	<i>PPARA</i>	G/A	0.04	0.02	0.47
rs6690158	1	145123867	<i>PRKAB2</i>	T/C	0.04	0.03	0.53

SNPs in or near biological candidate genes showing a nominally significant interaction with the metformin intervention in the DPP are shown. HRs are estimated for the minor allele (m) vs. the major allele (M) under an additive genetic model. Only the top SNP within each gene region is shown; the full set of results (including allele frequencies) is available in supplementary Table 2.

TABLE 3
Continued

Placebo		Metformin		Lifestyle	
HR (95% CI)	Observed <i>P</i>	HR (95% CI)	Observed <i>P</i>	HR (95% CI)	Observed <i>P</i>
1.26 (1.04–1.53)	0.02	1.15 (0.93–1.42)	0.20	1.34 (1.05–1.71)	0.02
0.84 (0.70–1.02)	0.08	0.68 (0.54–0.86)	0.001	0.94 (0.73–1.21)	0.65
0.75 (0.57–1.00)	0.05	0.67 (0.47–0.96)	0.03	0.74 (0.49–1.11)	0.14
1.20 (1.00–1.44)	0.05	1.37 (1.11–1.67)	0.003	0.96 (0.76–1.23)	0.76
0.77 (0.59–1.00)	0.05	0.85 (0.63–1.14)	0.27	0.67 (0.45–1.00)	0.05
0.83 (0.69–1.00)	0.05	0.98 (0.80–1.20)	0.85	0.64 (0.49–0.84)	0.001
1.22 (1.02–1.45)	0.03	1.07 (0.88–1.30)	0.51	1.24 (0.99–1.56)	0.07
1.17 (0.85–1.60)	0.33	1.33 (0.94–1.88)	0.10	1.51 (1.03–2.21)	0.03
0.76 (0.57–1.00)	0.05	0.85 (0.62–1.16)	0.31	0.75 (0.51–1.10)	0.14
1.08 (0.90–1.30)	0.39	1.14 (0.92–1.42)	0.23	1.40 (1.09–1.78)	0.01
0.82 (0.63–1.07)	0.15	0.78 (0.57–1.08)	0.14	0.72 (0.49–1.06)	0.10
1.05 (0.89–1.25)	0.56	1.29 (1.05–1.58)	0.02	1.25 (0.97–1.60)	0.08
0.91 (0.75–1.12)	0.37	0.89 (0.72–1.11)	0.32	0.65 (0.48–0.86)	0.003
1.16 (0.48–2.79)	0.74	2.66 (1.33–5.31)	0.01	2.23 (0.76–6.56)	0.15
1.16 (0.81–1.65)	0.42	1.99 (1.38–2.87)	0.0002	0.95 (0.56–1.60)	0.84
0.59 (0.31–1.13)	0.11	0.79 (0.37–1.72)	0.56	0.24 (0.06–1.01)	0.05
1.60 (1.09–2.34)	0.02	0.92 (0.56–1.52)	0.75	1.72 (1.00–2.95)	0.05
0.76 (0.56–1.02)	0.06	0.89 (0.63–1.25)	0.50	0.78 (0.52–1.16)	0.21
1.13 (0.91–1.40)	0.27	1.20 (0.95–1.52)	0.12	1.20 (0.91–1.59)	0.20
1.06 (0.86–1.31)	0.56	1.21 (0.95–1.56)	0.13	1.27 (0.96–1.68)	0.09
1.24 (0.89–1.74)	0.20	1.35 (0.87–2.09)	0.18	1.32 (0.77–2.28)	0.31

surprising that the DPP does not replicate all GWAS-derived findings in that range or that it fails to reach genome-wide significance in discovery efforts. Our null results on diabetes incidence for truly associated variants may be due to the high-risk population at baseline, the short time of follow-up (3.2 years on average), and/or the use of interventions effective in reducing diabetes incidence. On the other hand, considering the number of

variants likely to influence the phenotypes under study, even submaximal power is likely to provide a number of true positive associations. In this context, genotyped and imputed SNPs in the gene encoding the AMPK $\gamma 2$ subunit (*PRKAG2*) merit further consideration. While the association of SNPs in genes that encode metformin transporters with type 2 diabetes in the entire DPP cohort (if real) requires explanation, this could be due to a sufficiently

TABLE 4
Continued

Placebo		Metformin		Lifestyle	
HR (95% CI)	Observed <i>P</i>	HR (95% CI)	Observed <i>P</i>	HR (95% CI)	Observed <i>P</i>
1.69 (1.36–2.10)	0.000002	0.87 (0.65–1.16)	0.33	1.08 (0.79–1.47)	0.64
1.24 (1.04–1.48)	0.02	0.79 (0.63–0.98)	0.03	1.06 (0.83–1.37)	0.64
0.82 (0.61–1.11)	0.20	1.81 (1.35–2.43)	0.0001	1.29 (0.91–1.83)	0.15
0.76 (0.59–0.97)	0.03	1.31 (1.03–1.66)	0.03	1.07 (0.80–1.43)	0.65
0.88 (0.72–1.09)	0.24	1.43 (1.14–1.80)	0.002	0.78 (0.58–1.05)	0.10
1.15 (0.97–1.37)	0.11	0.78 (0.64–0.96)	0.02	1.09 (0.86–1.38)	0.46
0.54 (0.26–1.09)	0.08	2.15 (1.22–3.80)	0.01	1.11 (0.44–2.77)	0.83
1.61 (1.28–2.02)	0.00005	0.95 (0.70–1.28)	0.73	0.81 (0.55–1.18)	0.27
0.34 (0.15–0.75)	0.01	1.31 (0.76–2.29)	0.33	0.75 (0.26–2.15)	0.59
0.84 (0.68–1.04)	0.10	1.25 (0.99–1.59)	0.06	0.89 (0.68–1.17)	0.41
0.78 (0.49–1.22)	0.27	1.57 (1.09–2.27)	0.02	0.60 (0.31–1.17)	0.13
0.86 (0.69–1.08)	0.19	1.29 (1.02–1.64)	0.04	0.97 (0.73–1.31)	0.86
1.17 (0.96–1.43)	0.13	0.82 (0.64–1.04)	0.10	0.95 (0.72–1.26)	0.73
1.86 (1.19–2.91)	0.01	0.49 (0.17–1.40)	0.19	0.69 (0.27–1.78)	0.45
2.41 (1.08–5.37)	0.03	0.23 (0.03–1.62)	0.14	0.68 (0.17–2.71)	0.58
1.14 (0.95–1.37)	0.15	0.84 (0.69–1.02)	0.08	1.14 (0.90–1.44)	0.27
1.17 (0.85–1.62)	0.34	0.66 (0.43–1.02)	0.06	1.13 (0.72–1.78)	0.59
0.47 (0.23–0.96)	0.04	1.21 (0.73–1.99)	0.46	1.36 (0.77–2.38)	0.29
0.99 (0.77–1.27)	0.91	1.45 (1.12–1.88)	0.004	0.82 (0.58–1.17)	0.27
1.22 (1.01–1.46)	0.04	0.89 (0.71–1.13)	0.34	1.24 (0.97–1.59)	0.09
0.65 (0.20–2.13)	0.48	3.70 (1.56–8.80)	0.003	0.59 (0.08–4.45)	0.61
0.60 (0.27–1.33)	0.20	2.29 (1.10–4.76)	0.03	1.07 (0.44–2.65)	0.88
0.58 (0.33–1.03)	0.06	1.42 (0.85–2.36)	0.18	1.56 (0.87–2.81)	0.14

TABLE 5
Candidate gene variants showing a nominally significant interaction with the lifestyle intervention in the DPP

SNP	Chromosome	Position (NCBI 36)	Gene	Alleles (m/M)	Interaction genotype * lifestyle		FDR Q
					Observed P	Permuted P	
rs3792269	2	241180152	<i>CAPN10</i>	G/A	0.005	0.008	0.84
rs2425640	20	42461451	<i>HNF4A</i>	A/G	0.01	0.01	0.84
rs1342514	1	56959948	<i>PRKAA2</i>	G/C	0.01	0.005	0.84
rs4725408	7	150881542	<i>PRKAG2</i>	C/T	0.02	0.005	0.84
rs7599142	2	162701737	<i>GCG</i>	A/G	0.02	0.01	0.84
rs17367421	1	153553865	<i>PKLR</i>	C/G	0.02	0.01	0.84
rs12374408	4	23463581	<i>PPARGC1A</i>	T/C	0.02	0.03	0.84
rs3751151	12	119926582	<i>HNF1A</i>	T/A	0.02	0.02	0.84
rs11024298	11	17448407	<i>ABCC8</i>	T/G	0.02	0.01	0.84
rs6008306	22	45018756	<i>PPARA</i>	T/C	0.02	0.01	0.84
rs1422429	5	149146627	<i>PPARGC1B</i>	A/G	0.02	0.02	0.84
rs1054442	12	47675587	<i>PRKAG1</i>	C/A	0.03	0.02	0.84
rs9965495	18	56184656	<i>MC4R</i>	T/C	0.03	0.02	0.84
rs1008284	17	33136571	<i>HNF1B</i>	T/C	0.03	0.03	0.84
rs2018675	17	19382123	<i>SLC47A1</i>	T/C	0.03	0.02	0.84
rs8032587	15	98068933	<i>MEF2A</i>	C/T	0.03	0.02	0.84
rs7626560	3	12450088	<i>PPARG</i>	A/G	0.04	0.04	0.84
rs17161829	7	99187763	<i>CYP3A4</i>	A/G	0.04	0.01	0.84
rs6701920	1	159181541	<i>ITLN2</i>	A/G	0.04	0.02	0.84
rs461473	6	160463552	<i>SLC22A1</i>	T/C	0.04	0.04	0.84
rs17159890	1	145094998	<i>PRKAB2</i>	C/A	0.046	0.05	0.84
rs11904814	2	208135043	<i>CREB1</i>	C/A	0.046	0.05	0.84
rs17373414	3	188068221	<i>ADIPOQ</i>	T/C	0.049	0.06	0.84

SNPs in or near biological candidate genes showing a nominally significant interaction with the lifestyle intervention in the DPP are shown. HRs are estimated for the minor allele (m) vs. the major allele (M) under an additive genetic model. Only the top SNP within each gene region is shown; the full set of results (including allele frequencies) is available in supplementary Table 2.

strong effect in the metformin arm alone. Alternatively, SNPs in this region could be capturing variants in other nearby genes: for instance, immediately upstream of *SLC22A1* and *SLC22A2* in chromosome 6 lies the gene encoding the insulin-like growth factor 2 receptor (*IGF2R*), an excellent biological candidate.

This study constitutes the first large-scale prospective pharmacogenetic evaluation of metformin action in a controlled clinical trial. The UK Prospective Diabetes Study (29) and A Diabetes Outcome Progression Trial (ADOPT) (30) investigators independently showed that a substantial proportion of patients with type 2 diabetes eventually fail metformin therapy, defined by a need for additional pharmacotherapy to control hyperglycemia. Given the higher prior probability afforded by the known biological role of *SLC47A1* in disposing of metformin and the previously reported genetic association of the major allele at SNP rs2289669 with poorer metformin response (5), validation in the DPP can be convincing without achieving the levels of statistical significance required for novel findings. Our index SNP (rs8065082) is in tight LD with rs2289669 ($r^2 \sim 0.8$ in HapMap CEU) and the direction of effect is consistent in DPP, a cohort nearly 10-fold larger than the one documented in the original report from Rotterdam (5). Thus, our findings confirm those of Becker et al. (5) and suggest that major allele homozygotes at this locus ($\sim 30\%$ of the European population) may experience suboptimal responses to metformin treatment.

Our findings on the *SLC22A1* locus and metformin response are less robust. While our noted association with a missense SNP appears compelling, it is not among the most functional human variants described by

Shu et al. (3), and it is in weak LD with rs622342 ($r^2 \sim 0.14$ in HapMap CEU), a *SLC22A1* SNP associated with metformin response in another report from Rotterdam (31). SNP rs622342 was included among our tag SNPs but showed no evidence of an interaction with metformin (nominal $P = 0.69$) or an effect on diabetes incidence in any arm, raising the possibility that the original finding may have been spurious. Similarly, the *SLC22A2* missense SNP rs316019 (A270S), reported to influence metformin renal excretion and affect its plasma concentrations (32), did not significantly interact with metformin in the DPP (nominal $P = 0.35$). Our novel findings in the putative metformin drug targets *STK11* and AMPK require confirmation, as do those in *MEF2A* and *MEF2D*, themselves regulated by AMPK (33). One of the most significant interactions with metformin occurred at an SNP in *HNF4A*; given its role in hepatic gluconeogenesis (34), this intriguing result deserves further exploration. In contrast, the multiple interactions noted in the *ABCC8-KCNJ11* locus reported previously (35) do not offer a clear mechanism of action. Finally, nominal associations with response to lifestyle modification should be replicated in cohorts that underwent a similar intervention.

In summary, we have conducted a large-scale genetic association study in the DPP and replicated the association of a polymorphism in a metformin transporter with metformin response. Other hypothesis-generating results require more detailed characterization in the DPP and follow-up in independent samples. A focus on likely functional variants may uncover loci with stronger effects.

TABLE 5
Continued

Placebo		Metformin		Lifestyle	
HR (95% CI)	Observed <i>P</i>	HR (95% CI)	Observed <i>P</i>	HR (95% CI)	Observed <i>P</i>
1.61 (1.28–2.02)	0.00005	0.95 (0.70–1.28)	0.73	0.81 (0.55–1.18)	0.27
0.83 (0.69–1.01)	0.06	0.93 (0.75–1.16)	0.51	1.23 (0.96–1.58)	0.11
1.07 (0.90–1.29)	0.44	1.14 (0.92–1.41)	0.25	0.71 (0.54–0.92)	0.01
0.82 (0.39–1.73)	0.61	1.02 (0.41–2.55)	0.97	2.90 (1.52–5.56)	0.001
1.02 (0.84–1.23)	0.86	1.00 (0.81–1.22)	0.97	0.66 (0.51–0.86)	0.002
0.47 (0.23–0.96)	0.04	1.21 (0.73–1.99)	0.46	1.36 (0.77–2.38)	0.29
1.13 (0.94–1.36)	0.20	0.94 (0.76–1.17)	0.60	0.77 (0.59–1.02)	0.07
0.89 (0.73–1.09)	0.26	0.82 (0.65–1.04)	0.10	1.30 (1.01–1.67)	0.04
1.17 (0.82–1.67)	0.39	1.01 (0.65–1.58)	0.95	0.44 (0.21–0.92)	0.03
0.52 (0.25–1.08)	0.08	0.96 (0.49–1.91)	0.91	2.05 (0.97–4.33)	0.06
0.83 (0.69–0.99)	0.04	0.96 (0.79–1.17)	0.70	1.19 (0.94–1.50)	0.15
1.22 (1.02–1.45)	0.03	1.08 (0.88–1.32)	0.47	0.87 (0.68–1.12)	0.27
1.09 (0.90–1.31)	0.39	1.20 (0.97–1.49)	0.09	0.75 (0.58–0.98)	0.04
1.30 (1.08–1.58)	0.01	0.91 (0.71–1.15)	0.43	0.89 (0.68–1.18)	0.42
1.18 (0.99–1.40)	0.06	1.13 (0.92–1.38)	0.24	0.88 (0.69–1.13)	0.33
1.19 (0.72–1.95)	0.50	1.07 (0.56–2.05)	0.83	0.31 (0.10–1.00)	0.05
1.16 (0.93–1.45)	0.19	1.12 (0.88–1.44)	0.36	0.76 (0.55–1.06)	0.10
1.05 (0.69–1.60)	0.82	1.24 (0.77–1.98)	0.38	0.32 (0.11–0.93)	0.04
1.86 (1.19–2.91)	0.01	0.49 (0.17–1.40)	0.19	0.69 (0.27–1.78)	0.45
0.92 (0.64–1.33)	0.67	0.93 (0.64–1.36)	0.72	1.50 (1.01–2.22)	0.04
0.84 (0.56–1.26)	0.40	1.01 (0.62–1.63)	0.98	1.88 (1.13–3.14)	0.02
1.03 (0.86–1.23)	0.78	1.15 (0.93–1.42)	0.18	0.74 (0.57–0.96)	0.03
0.82 (0.57–1.18)	0.28	0.90 (0.63–1.29)	0.57	1.34 (0.90–1.98)	0.15

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REFERENCES

1. Prokopenko I, McCarthy MI, Lindgren CM. Type 2 diabetes: new genes, new understanding. *Trends Genet* 2008;24:613–621

2. Pearson ER. Pharmacogenetics in diabetes. *Curr Diab Rep* 2009;9:172–181
3. Shu Y, Sheardown SA, Brown C, Owen RP, Zhang S, Castro RA, Ianculescu AG, Yue L, Lo JC, Burchard EG, Brett CM, Giacomini KM. Effect of genetic variation in the organic cation transporter 1 (OCT1) on metformin action. *J Clin Invest* 2007;117:1422–1431
4. Zhou K, Donnelly LA, Kimber CH, Donnan PT, Doney AS, Leese G, Hattersley AT, McCarthy MI, Morris AD, Palmer CN, Pearson ER. Reduced-function SLC22A1 polymorphisms encoding organic cation transporter 1 and glycemic response to metformin: a GoDARTS study. *Diabetes* 2009;58:1434–1439
5. Becker ML, Visser LE, van Schaik RH, Hofman A, Uitterlinden AG, Stricker BH. Genetic variation in the multidrug and toxin extrusion 1 transporter protein influences the glucose-lowering effect of metformin in patients with diabetes: a preliminary study. *Diabetes* 2009;58:745–749
6. Knowler WC, Barrett-Connor E, Fowler SE, Hamman RF, Lachin JM, Walker EA, Nathan DM, Diabetes Prevention Program Research Group. Reduction in the incidence of type 2 diabetes with lifestyle intervention or metformin. *N Engl J Med* 2002;346:393–403
7. Diabetes Prevention Program Research Group. The Diabetes Prevention Program: Design and methods for a clinical trial in the prevention of type 2 diabetes. *Diabetes Care* 1999;22:623–634
8. de Bakker PI, Yelensky R, Pe'er I, Gabriel SB, Daly MJ, Altshuler D. Efficiency and power in genetic association studies. *Nat Genet* 2005;37:1217–1223
9. Diabetes Genetics Initiative of Broad Institute of Harvard and MIT, Lund University, and Novartis Institutes of BioMedical Research, Saxena R, Voight BF, Lyssenko V, Burtt NP, de Bakker PI, Chen H, Roix JJ, Kathiresan S, Hirschhorn JN, Daly MJ, Hughes TE, Groop L, Altshuler D, Almgren P, Florez JC, Meyer J, Ardlie K, Bengtsson Boström K, Isomaa B, Lettre G, Lindblad U, Lyon HN, Melander O, Newton-Cheh C, Nilsson P, Orho-Melander M, Råstam L, Speliotes EK, Taskinen MR, Tuomi T, Guiducci C, Berglund A, Carlson J, Gianniny L, Hackett R, Hall L, Holmkvist J, Laurila E, Sjögren M, Sterner M, Surti A, Svensson M, Svensson M, Tewhey R, Blumenstiel B, Parkin M, Defelice M, Barry R, Brodeur W, Camarata J, Chia N, Fava M, Gibbons J, Handsaker B, Healy C, Nguyen K, Gates C, Sougnez C, Gage D, Nizzari M, Gabriel SB, Chim GW, Ma Q, Parikh H, Richardson D, Ricke D, Purcell S. Genome-wide association analysis identifies loci for type 2 diabetes and triglyceride levels. *Science* 2007;316:1331–1336
10. Zeggini E, Scott LJ, Saxena R, Voight BF, Marchini JL, Hu T, de Bakker PI, Abecasis GR, Almgren P, Andersen G, Ardlie K, Boström KB, Bergman RN, Bonnycastle LL, Borch-Johnsen K, Burtt NP, Chen H, Chines PS, Daly MJ, Deodhar P, Ding CJ, Doney AS, Duren WL, Elliott KS, Erdos MR, Frayling TM, Freathy RM, Gianniny L, Grallert H, Grarup N, Groves CJ, Guiducci C, Hansen T, Herder C, Hitman GA, Hughes TE, Isomaa B, Jackson AU, Jørgensen T, Kong A, Kubalanza K, Kuruvilla FG, Kuusisto J, Langenberg C, Lango H, Lauritzen T, Li Y, Lindgren CM, Lyssenko V, Marville AF, Meisinger C, Midtjell K, Mohlke KL, Morken MA, Morris AD, Narisu N, Nilsson P, Owen KR, Palmer CN, Payne F, Perry JR, Pettersen E, Platou C, Prokopenko I, Qi L, Qin L, Rayner NW, Rees M, Roix JJ, Sandbaek A, Shields B, Sjögren M, Steinthorsdottir V, Stringham HM, Swift AJ, Thorleifsson G, Thorsteinsdottir U, Timpson NJ, Tuomi T, Tuomilehto J, Walker M, Watanabe RM, Weedon MN, Willer CJ, Wellcome Trust Case Control Consortium, Illig T, Hveem K, Hu FB, Laakso M, Stefansson K, Pedersen O, Wareham NJ, Barroso I, Hattersley AT, Collins FS, Groop L, McCarthy MI, Boehnke M, Altshuler D. Meta-analysis of genome-wide association data and large-scale replication identifies additional susceptibility loci for type 2 diabetes. *Nat Genet* 2008;40:638–645
11. Florez JC, Manning AK, Dupuis J, McAteer J, Irenze K, Gianniny L, Mirel DB, Fox CS, Cupples LA, Meigs JB. A 100K genome-wide association scan for diabetes and related traits in the Framingham Heart Study: replication and integration with other genome-wide datasets. *Diabetes* 2007;56:3063–3074
12. Hanson RL, Bogardus C, Duggan D, Kobes S, Knowlton M, Infante AM, Marovich L, Benitez D, Baier LJ, Knowler WC. A search for variants associated with young-onset type 2 diabetes in American Indians in a 100K genotyping array. *Diabetes* 2007;56:3045–3052
13. Rampersaud E, Damcott CM, Fu M, Shen H, McArdle P, Shi X, Shelton J, Yin J, Chang YP, Ott SH, Zhang L, Zhao Y, Mitchell BD, O'Connell J, Shuldiner AR. Identification of novel candidate genes for type 2 diabetes from a genome-wide association scan in the Old Order Amish: evidence for replication from diabetes-related quantitative traits and from independent populations. *Diabetes* 2007;56:3053–3062
14. Willer CJ, Speliotes EK, Loos RJ, Li S, Lindgren CM, Heid IM, Berndt SI, Elliott AL, Jackson AU, Lamina C, Lettre G, Lim N, Lyon HN, McCarrroll SA, Papadakis K, Qi L, Randall JC, Rocaecca RM, Sanna S, Scheet P, Weedon MN, Wheeler E, Zhao JH, Jacobs LC, Prokopenko I, Soranzo N, Tanaka T, Timpson NJ, Almgren P, Bennett A, Bergman RN, Bingham SA, Bonnycastle LL, Brown M, Burtt NP, Chines P, Coin L, Collins FS, Connell JM, Cooper C, Smith GD, Dennison EM, Deodhar P, Elliott P, Erdos MR, Estrada K, Evans DM, Gianniny L, Gieger C, Gillson CJ, Guiducci C, Hackett R, Hadley D, Hall AS, Havulinna AS, Hebebrand J, Hofman A, Isomaa B, Jacobs KB, Johnson T, Jousilahti P, Jovanovic Z, Khaw KT, Kraft P, Kuokkanen M, Kuusisto J, Laitinen J, Lakatta EG, Luan J, Luben RN, Mangino M, McArdle WL, Meitinger T, Mulas A, Munroe PB, Narisu N, Ness AR, Northstone K, O'Rahilly S, Purmann C, Rees MG, Ridderstråle M, Ring SM, Rivadeneira F, Ruokonen A, Sandhu MS, Saramies J, Scott LJ, Scuteri A, Silander K, Sims MA, Song K, Stephens J, Stevens S, Stringham HM, Tung YC, Valle TT, Van Duijn CM, Vimalaswaran KS, Vollenweider P, Waeber G, Wallace C, Watanabe RM, Waterworth DM, Watkins N, Wellcome Trust Case Control Consortium, Witteman JC, Zeggini E, Zhai G, Zillikens MC, Altshuler D, Caulfield MJ, Chanock SJ, Farooqi IS, Ferrucci L, Guralnik JM, Hattersley AT, Hu FB, Jarvelin MR, Laakso M, Mooser V, Ong KK, Ouwehand WH, Salomaa V, Samani NJ, Spector TD, Tuomi T, Tuomilehto J, Uda M, Uitterlinden AG, Wareham NJ, Deloukas P, Frayling TM, Groop LC, Hayes RB, Hunter DJ, Mohlke KL, Peltonen L, Schlessinger D, Strachan DP, Wichmann HE, McCarthy MI, Boehnke M, Barroso I, Abecasis GR, Hirschhorn JN, Genetic Investigation of ANthropometric Traits Consortium. Six new loci associated with body mass index highlight a neuronal influence on body weight regulation. *Nat Genet* 2009;41:25–34
15. Thorleifsson G, Walters GB, Gudbjartsson DF, Steinthorsdottir V, Sulem P, Helgadóttir A, Styrkarsdóttir U, Gretarsdóttir S, Thorlacius S, Jonsdóttir I, Jonsdóttir T, Olafsdóttir EJ, Olafsdóttir GH, Jonsson T, Jonsson F, Borch-Johnsen K, Hansen T, Andersen G, Jørgensen T, Lauritzen T, Aben KK, Verbeek AL, Roeleveld N, Kampman E, Yanek LR, Becker LC, Tryggvadóttir L, Rafnar T, Becker DM, Gulcher J, Kiemeny LA, Pedersen O, Kong A, Thorsteinsdóttir U, Stefansson K. Genome-wide association yields new sequence variants at seven loci that associate with measures of obesity. *Nat Genet* 2009;41:18–24
16. Kathiresan S, Melander O, Guiducci C, Surti A, Burtt NP, Rieder MJ, Cooper GM, Roos C, Voight BF, Havulinna AS, Wahlstrand B, Hedner T, Corella D, Tai ES, Ordovas JM, Berglund G, Vartiainen E, Jousilahti P, Hedblad B, Taskinen MR, Newton-Cheh C, Salomaa V, Peltonen L, Groop L, Altshuler DM, Orho-Melander M. Six new loci associated with blood low-density lipoprotein cholesterol, high-density lipoprotein cholesterol or triglycerides in humans. *Nat Genet* 2008;40:189–197
17. Willer CJ, Sanna S, Jackson AU, Scuteri A, Bonnycastle LL, Clarke R, Heath SC, Timpson NJ, Najjar SS, Stringham HM, Strait J, Duren WL, Maschio A, Busonero F, Mulas A, Albai G, Swift AJ, Morken MA, Narisu N, Bennett D, Parish S, Shen H, Galan P, Meneton P, Herberg S, Zelenika D, Chen WM, Li Y, Scott LJ, Scheet PA, Sundvall J, Watanabe RM, Nagaraja R, Ebrahim S, Lawlor DA, Ben-Shlomo Y, Davey-Smith G, Shuldiner AR, Collins R, Bergman RN, Uda M, Tuomilehto J, Cao A, Collins FS, Lakatta E, Lathrop GM, Boehnke M, Schlessinger D, Mohlke KL, Abecasis GR. Newly identified loci that influence lipid concentrations and risk of coronary artery disease. *Nat Genet* 2008;40:161–169
18. Kathiresan S, Willer CJ, Peloso GM, Demissie S, Musunuru K, Schadt EE, Kaplan L, Bennett D, Li Y, Tanaka T, Voight BF, Bonnycastle LL, Jackson AU, Crawford G, Surti A, Guiducci C, Burtt NP, Parish S, Clarke R, Zelenika D, Kubalanza KA, Morken MA, Scott LJ, Stringham HM, Galan P, Swift AJ, Kuusisto J, Bergman RN, Sundvall J, Laakso M, Ferrucci L, Scheet P, Sanna S, Uda M, Yang Q, Lunetta KL, Dupuis J, de Bakker PI, O'Donnell CJ, Chambers JC, Kooner JS, Herberg S, Meneton P, Lakatta EG, Scuteri A, Schlessinger D, Tuomilehto J, Collins FS, Groop L, Altshuler D, Collins R, Lathrop GM, Melander O, Salomaa V, Peltonen L, Orho-Melander M, Ordovas JM, Boehnke M, Abecasis GR, Mohlke KL, Cupples LA. Common variants at 30 loci contribute to polygenic dyslipidemia. *Nat Genet* 2009;41:56–65
19. Smith MW, Patterson N, Lautenberger JA, Truelove AL, McDonald GL, Waliszewska A, Kessing BD, Malasky MJ, Scafe C, Le E, De Jager PL, Mignault AA, Yi Z, De The G, Essex M, Sankale JL, Moore JH, Poku K, Phair JP, Goedert JJ, Vlahov D, Williams SM, Tishkoff SA, Winkler CA, De La Vega FM, Woodage T, Sninsky JJ, Hafler DA, Altshuler D, Gilbert DA, O'Brien SJ, Reich D. A high-density admixture map for disease gene discovery in African Americans. *Am J Hum Genet* 2004;74:1001–1013
20. Price AL, Patterson N, Yu F, Cox DR, Waliszewska A, McDonald GJ, Tandon A, Schirmer C, Neubauer J, Bedoya G, Duque C, Villegas A, Bortolini MC, Salzano FM, Gallo C, Mazzotti G, Tello-Ruiz M, Riba L, Aguilar-Salinas CA, Canizales-Quintero S, Menjivar M, Klitz W, Henderson B, Haiman CA, Winkler C, Tusie-Luna T, Ruiz-Linares A, Reich D. A genomewide admixture map for Latino populations. *Am J Hum Genet* 2007;80:1024–1036
21. Li Y, Abecasis GR. Mach 1.0: Rapid haplotype reconstruction and missing genotype inference. *Am J Hum Genet* 2006;79:2290

22. Pritchard JK, Stephens M, Donnelly P. Inference of population structure using multilocus genotype data. *Genetics* 2000;155:945–959
23. Dudoit S, Shaffer JP, Boldrick JC. Multiple hypothesis testing in microarray experiments. *Statistical Science* 2003;18:71–103
24. Potter DM. Omnibus permutation tests of the association of an ensemble of genetic markers with disease in case-control studies. *Genet Epidemiol* 2006;30:438–446
25. Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. *J R Stat Soc* 1995;57:289–300
26. Hattersley AT, McCarthy MI. What makes a good genetic association study? *Lancet* 2005;366:1315–1323
27. Pe'er I, Yelensky R, Altshuler D, Daly MJ. Estimation of the multiple testing burden for genomewide association studies of nearly all common variants. *Genet Epidemiol* 2008;32:381–385
28. Moore AF, Jablonski KA, McAteer JB, Saxena R, Pollin TI, Franks PW, Hanson RL, Shuldiner AR, Knowler WC, Altshuler D, Florez JC, Diabetes Prevention Program Research Group. Extension of type 2 diabetes genome-wide association scan results in the Diabetes Prevention Program. *Diabetes* 2008;57:2503–2510
29. Turner RC, Cull CA, Frighi V, Holman RR. Glycemic control with diet, sulfonylurea, metformin, or insulin in patients with type 2 diabetes mellitus: progressive requirement for multiple therapies (UKPDS 49): UK Prospective Diabetes Study (UKPDS) Group. *JAMA* 1999;281:2005–2012
30. Kahn SE, Haffner SM, Heise MA, Herman WH, Holman RR, Jones NP, Kravitz BG, Lachin JM, O'Neill MC, Zinman B, Viberti G, ADOPT Study Group. Glycemic durability of rosiglitazone, metformin, or glyburide monotherapy. *N Engl J Med* 2006;355:2427–2443
31. Becker ML, Visser LE, van Schaik RH, Hofman A, Uitterlinden AG, Stricker BH. Genetic variation in the organic cation transporter 1 is associated with metformin response in patients with diabetes mellitus. *Pharmacogenomics J* 2009;9:242–247
32. Song IS, Shin HJ, Shim EJ, Jung IS, Kim WY, Shon JH, Shin JG. Genetic variants of the organic cation transporter 2 influence the disposition of metformin. *Clin Pharmacol Ther* 2008;84:559–562
33. Holmes BF, Sparling DP, Olson AL, Winder WW, Dohm GL. Regulation of muscle GLUT4 enhancer factor and myocyte enhancer factor 2 by AMP-activated protein kinase. *Am J Physiol Endocrinol Metab* 2005;289:E1071–E1076
34. Rhee J, Inoue Y, Yoon JC, Puigserver P, Fan M, Gonzalez FJ, Spiegelman BM. Regulation of hepatic fasting response by PPAR γ coactivator-1 α (PGC-1): requirement for hepatocyte nuclear factor 4 α in gluconeogenesis. *Proc Natl Acad Sci U S A* 2003;100:4012–4017
35. Florez JC, Jablonski KA, Kahn SE, Franks PW, Dabelea D, Hamman RF, Knowler WC, Nathan DM, Altshuler D. Type 2 diabetes-associated missense polymorphisms *KCNJ11* E23K and *ABCC8* A1369S influence progression to diabetes and response to interventions in the Diabetes Prevention Program. *Diabetes* 2007;56:531–536