

Distinct Diagnostic Criteria of Fulminant Type 1 Diabetes Based on Serum C-Peptide Response and HbA_{1c} Levels at Onset

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OBJECTIVE— Diagnostic criteria in fulminant type 1 diabetes, a novel subtype of type 1 diabetes, remain unclear.

RESEARCH DESIGN AND METHODS— We analyzed basal and longitudinal changes of serum C-peptide levels during a 75-g oral glucose tolerance test (OGTT) in 125 consecutively recruited patients with type 1 diabetes including fulminant type 1 diabetes ($n = 25$) and acute-onset type 1 diabetes ($n = 100$). Discriminating criteria of fulminant type 1 diabetes were examined using receiver-operating characteristic curve analysis and multiple logistic regression analysis.

RESULTS— The integrated values of serum C-peptide response during OGTT (Σ C-peptide) in fulminant type 1 diabetes at onset, 1 year, and 2 years after onset were markedly lower than those in acute-onset type 1 diabetes. None of the patients with fulminant type 1 diabetes had improvement of C-peptide response to OGTT. Fasting C-peptide values at onset in fulminant type 1 diabetes were significantly lower than those in acute-onset type 1 diabetes. We established diagnostic criteria of serum C-peptide and HbA_{1c} levels at onset that discriminate fulminant type 1 diabetes from acute-onset type 1 diabetes with high sensitivity and specificity: a criterion in which the levels of both the fasting C-peptide is ≤ 0.033 nmol/l and HbA_{1c} is $\leq 8.0\%$ or a criterion in which the levels of both the Σ C-peptide is ≤ 0.540 nmol/l and HbA_{1c} is $\leq 8.0\%$.

CONCLUSIONS— Fulminant type 1 diabetes has extremely low β -cell function at onset that rarely recovers after onset. Sensitive and specific diagnostic criteria were established for detection of fulminant type 1 diabetes based on serum C-peptide and HbA_{1c} levels at onset.

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Abbreviations: AUC, area under receiver-operating characteristic curve; GADAb, GAD autoantibody; IAA, insulin autoantibody; IA-2Ab, insulinoma-associated protein 2/islet cell antigen 512 autoantibody; ICA, islet cell antibody; OGTT, oral glucose tolerance test; ROC, receiver-operating characteristic.

A table elsewhere in this issue shows conventional and Système International (SI) units and conversion factors for many substances.

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In 1988, a study suggested that there might be a novel subtype of type 1 diabetes that was called the “fulminant form of type 1 diabetes” (1,2). In 2000, this subtype, which had the same characteristics as the subtype discussed in the 1988 study, was described and named “fulminant type 1 diabetes” (3–6). The characteristic features at onset of this subtype of type 1 diabetes (1–6) include: 1) abrupt onset of diabetes with fulminant symptoms including marked hyperglycemia, severe diabetic ketoacidotic coma, and normal or near-normal HbA_{1c} levels; 2) absence of autoantibodies against islet cells including islet cell antibodies (ICA), GAD autoantibodies (GADAb), insulinoma-associated protein 2/islet cell antigen 512 autoantibodies (IA-2Ab), and insulin autoantibodies (IAA); and 3) involvement of exocrine pancreas as well as pancreatic islets with elevated serum levels of pancreatic enzymes. The key issue for the diagnosis in this new subtype of type 1 diabetes is absence of autoantibodies against islet cells. However, the measurement of pancreatic autoantibodies including GADAb, ICA, IA-2Ab, and IAA is in the process of being standardized (7), and these autoantibodies can be measured in a limited number of laboratories. Data on longitudinal changes of serum C-peptide levels as well as the basal values in newly diagnosed patients with type 1 diabetes, including fulminant type 1 diabetes and classical type 1 diabetes, were obtained prospectively. We tried to establish sensitive and specific criteria for clinical diagnosis of fulminant type 1 diabetes at onset based on serum C-peptide concentration.

RESEARCH DESIGN AND METHODS

A total of 126 newly diagnosed type 1 diabetic patients (onset age 34 ± 15 years [mean \pm SD]; age range 2–71; 70 men, 56 women) were consecutively recruited in the “Toranomon Prospective Study on Type 1 Diabetes” from

1980 to 2001 (5,8–13). All patients met the American Diabetes Association criteria for type 1 diabetes (14). The duration from onset of diabetes symptoms to hospitalization was within 90 days in all case subjects. The exclusion criteria were the presence of mitochondrial DNA mutation (A/G) at 3,243 bp and/or hepatocyte nuclear factor-1 α gene mutation. One patient was excluded because of the presence of mitochondrial DNA mutation (A/G) at 3,243 bp. According to the tentative criteria in previous studies (3,5), 25 patients who were negative for autoantibodies against pancreatic antigens (including ICA, GADAb, IA-2Ab, and IAA) and had normal or near-normal HbA_{1c} levels ($\leq 8.3\%$) at onset of diabetes were diagnosed as having fulminant type 1 diabetes. The remaining 100 patients were subdivided as acute-onset type 1 diabetes. Residual β -cell function of the total 125 patients, including 25 patients with fulminant type 1 diabetes and 100 patients with acute-onset type 1 diabetes, was analyzed in this study.

At onset of overt diabetes, all patients were hospitalized. Their clinical characteristics were recorded, and plasma glucose, HbA_{1c}, arterial pH, serum elastase 1, serum amylase, and serum lipase were measured within 2 days after initial diagnosis. HbA_{1c} levels were measured by a high-performance liquid chromatography method calibrated with Japan Diabetes Society Calibrator Lot 2 (15). This method for HbA_{1c} assay was standardized by the Japan Diabetes Society/Japanese Society of Clinical Chemistry and established a firm and reproducible link to the method of the International Federation of Clinical Chemistry (15). In all patients with type 1 diabetes, blood samples for the assay for pancreatic autoantibodies were obtained at least twice within 10 and 30 days after the diagnosis of diabetes. All serum samples for autoantibody and C-peptide assays were kept at -80°C until assay.

Diabetes-related autoantibodies (including ICA, GADAb, IA-2Ab, and IAA) were assayed as previously described (8,16–18). Our laboratory participated in the second through fifth International Workshop on Standardization of the ICA Assay, in which we established the quality of our ICA assay as follows: a cutoff point of 5 Juvenile Diabetes Foundation units, sensitivity of 90%, and specificity of 92% (16). The sensitivity and specificity of the

GADAb assay were 80 and 100%, respectively, in the second International GADAb Workshop (16). The IA-2Ab assay was evaluated in the third proficiency IA-2Ab test organized by the Research Institute for Children, and the results showed 100% sensitivity and 100% specificity (17). The sensitivity and specificity of the IAA assay were both 100% in the fourth International IAA Workshop (18). After obtaining written informed consent, detection of mitochondrial DNA mutation (A/G) at 3,243 bp and hepatocyte nuclear factor-1 α gene mutation was performed as previously described (19,20). Serum C-peptide levels were measured as previously described (9). The assay quality was checked using recombinant human C-peptide of World Health Organization standards (ampoules coded 84/510) (21) and two control samples (Daiichi Pharmaceutical, Tokyo, Japan). At the C-peptide concentration of 0.760 and 0.056 nmol/l, the intra-assay coefficient of variation (CV) values (10 replicates) were 3.4 and 9.5%, respectively. The interassay CV values (10 occasions) at concentrations of C-peptide of 0.760 and 0.056 nmol/l were, respectively, 8.1 and 14.2%. The minimal detection limit of C-peptide concentration was 0.017 nmol/l (9). C-peptide levels < 0.017 nmol/l were regarded as 0.000 nmol/l in this study. Samples from each subject were in the same assay run. Patients were tested using a 75-g oral glucose tolerance test (OGTT) at onset ($n = 125$), at 1 year ($n = 66$), and at 2 years ($n = 60$) after onset. OGTT at onset was performed for each patient after obtaining steady glycemic control by insulin therapy (3–4 weeks). The response of serum C-peptide during OGTT was quantified as the integrated value of the serum C-peptide levels obtained during OGTT at 0, 30, 60, 90, and 120 min: Σ C-peptide. The improvement of residual β -cell function was defined as increments of Σ C-peptide values > 0.331 nmol/l (1.0 ng/ml) when compared with the values at onset during the follow-up period.

Statistical analysis

The Mann-Whitney *U* test and the Fisher's exact test were applied to compare the values of clinical features among the different subgroups of type 1 diabetes. The Wilcoxon's signed-rank test was used to compare the longitudinal changes of the Σ C-peptide values of each subtype of type 1 diabetes. Frequencies of subjects who

had their improvement of residual β -cell function in different subgroups of type 1 diabetes were compared using Fisher's exact test. Receiver-operating characteristic (ROC) curves were analyzed to define the optimal cutoff values of clinical and laboratory findings at onset for discrimination of fulminant type 1 diabetes from acute-onset type 1 diabetes (22). Each discriminating criterion of clinical or laboratory finding at onset was devised based on the corresponding cutoff value. The areas under ROC curves (AUCs), which indicate the accuracy of the tests, were calculated and compared (23). A multiple logistic regression analysis was performed to identify independent factors for prediction of fulminant type 1 diabetes. The following variables of clinical and laboratory findings at onset were included in the multiple regression analysis: age at onset, sex, BMI, duration of hyperglycemic symptoms before diagnosis, plasma glucose levels, arterial pH, serum amylase levels, serum lipase levels, serum elastase 1 levels, Σ C-peptide values, and fasting C-peptide levels. Continuous variables were dichotomized based on whether they met or did not meet the discriminating criteria, which were devised by the optimal cutoff values as mentioned above, and then these variables were included in multiple regression analysis. A multiple regression model for predicting diagnosis of fulminant type 1 diabetes was devised and evaluated on our study population. The model was constructed by stepwise selection with $P = 0.05$ used as the criterion for both entry into and retention in the model. We used JMP software (version 5) and MedCalc software (version 7) for statistical analyses. All data except for AUCs were expressed as mean \pm SD. The protocol of the Toranomon Prospective Study on Type 1 Diabetes was approved by the Ethical Committee of Toranomon Hospital.

RESULTS

Clinical features at onset of diabetes in patients with fulminant type 1 diabetes

Twenty-five patients with fulminant type 1 diabetes had significantly shorter duration of hyperglycemic symptoms before diagnosis (mean 4 ± 3 days, range 1–10, $P < 0.0001$) versus acute-onset type 1 diabetes (mean 48 ± 32 days, range 1–90, $n = 100$). Plasma glucose levels at onset of

diabetes in patients with fulminant type 1 diabetes (47.4 ± 20.4 mmol/l [16.9–102.0]) were significantly higher than those in patients with acute-onset type 1 diabetes (26.7 ± 12.8 mmol/l [11.3–71.1]) ($P < 0.0001$). In contrast, the levels of HbA_{1c} at onset of diabetes in patients with fulminant type 1 diabetes ($6.5 \pm 1.1\%$ [4.3–8.3]) were significantly lower than those in patients with acute-onset type 1 diabetes ($11.3 \pm 2.7\%$ [5.1–19.1]) ($P < 0.0001$). The degree of acidosis in patients with fulminant type 1 diabetes (arterial blood pH 7.11 ± 0.14 [6.91–7.34]) was more severe than that in patients with acute-onset type 1 diabetes (7.28 ± 0.09 [7.02–7.39]) ($P < 0.0001$). BMI at onset of patients with fulminant type 1 diabetes (20.5 ± 2.5 kg/m² [16.7–28.7]) was significantly higher than that of patients with acute-onset type 1 diabetes (18.7 ± 2.4 kg/m² [13.8–26.7]) ($P = 0.0009$). Ninety-six percent (24/25) of patients with fulminant type 1 diabetes had elevated levels of serum pancreatic enzymes. Age at onset (37 ± 14 years [21–65]) and sex (16 men, 9 women) in patients with fulminant type 1 diabetes were not significantly different from those in patients with acute-onset type 1 diabetes (age at onset 33 ± 15 years [2–71], 54 men, 46 women). Among 100 patients with acute-onset type 1 diabetes, ICA was positive in 74% (74/100), GADAb in 69% (69/100), IA-2Ab in 61% (61/100), and IAA in 15% (15/100). All patients with fulminant type 1 diabetes were negative for ICA, GADAb, IA-2Ab, and IAA. All features of patients with fulminant type 1 diabetes are compatible with those in previously reported studies (1–6).

Longitudinal changes of residual β -cell function and diagnostic criteria in patients with fulminant type 1 diabetes

Residual β -cell function. The values of Σ C-peptide in patients with fulminant type 1 diabetes were markedly lower than those in patients with acute-onset type 1 diabetes at onset ($P < 0.0001$), 1 year after onset ($P < 0.0001$), and 2 years after onset ($P < 0.0001$) (Fig. 1). The Σ C-peptide values in patients with fulminant type 1 diabetes remained unchanged during a 2-year period after onset (Fig. 1). None of the patients with fulminant type 1 diabetes had improvement of their Σ C-peptide values during a 2-year period after onset. In contrast, in patients with

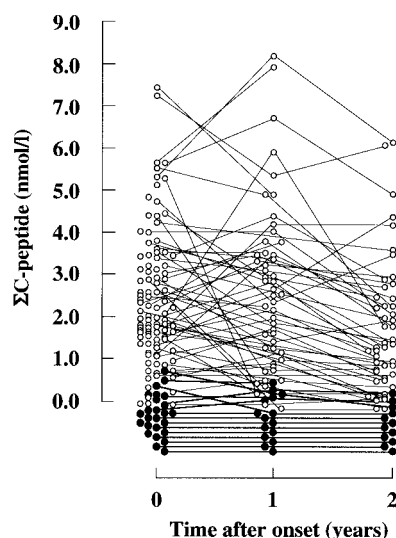


Figure 1—Longitudinal changes of residual β -cell function in type 1 diabetes. The mean integrated values of serum C-peptide levels obtained during OGTT (Σ C-peptide) in patients with fulminant type 1 diabetes (●) were 0.087 ± 0.190 nmol/l at onset ($n = 25$), 0.083 ± 0.157 nmol/l at 1 year after onset ($n = 15$), and 0.025 ± 0.078 nmol/l at 2 years after onset ($n = 12$). The mean values in patients with acute-onset type 1 diabetes (○) were 2.443 ± 1.550 nmol/l at onset ($n = 100$), 2.611 ± 1.970 nmol/l at 1 year after onset ($n = 51$), and 1.791 ± 1.548 nmol/l at 2 years after onset ($n = 48$).

acute-onset type 1 diabetes, the Σ C-peptide values at 2 years after onset were significantly lower than those at onset ($P < 0.0001$) and 1 year after onset ($P < 0.0001$), although there was no significant change between the Σ C-peptide values at onset and 1 year after onset. Furthermore, 39% (20/51) of patients with acute-onset type 1 diabetes had improvement of their Σ C-peptide values at 1 year after onset ($P = 0.0030$ vs. fulminant type 1 diabetes). Fasting serum C-peptide values at onset (mean 0.009 ± 0.021 nmol/l, $n = 25$) as well as Σ C-peptide values in fulminant type 1 diabetes were significantly lower than those in acute-onset type 1 diabetes (0.243 ± 0.139 nmol/l, $n = 100$, $P < 0.0001$).

ROC curve analysis. The AUCs for Σ C-peptide values (0.974 ± 0.013 [mean \pm SE]) and fasting C-peptide levels (0.973 ± 0.013) were significantly greater than AUCs for serum amylase levels (0.877 ± 0.046 , $P = 0.034$ and $P = 0.038$, respectively), arterial pH (0.841 ± 0.037 , $P < 0.001$ and $P = 0.001$, respec-

tively), plasma glucose levels (0.827 ± 0.053 , both $P = 0.006$), serum lipase levels (0.797 ± 0.056 , $P = 0.002$ and $P = 0.001$, respectively), BMI (0.715 ± 0.062 , both $P < 0.001$), and onset age (0.555 ± 0.066 , both $P < 0.001$) (Fig. 2A–D). The AUCs for HbA_{1c} levels (0.969 ± 0.014) and duration of hyperglycemic symptoms (0.944 ± 0.020) were significantly greater than AUCs for arterial pH ($P = 0.001$ and $P = 0.004$, respectively), plasma glucose levels ($P = 0.009$ and $P = 0.021$, respectively), serum lipase levels ($P = 0.003$ and $P = 0.011$, respectively), BMI (both $P < 0.001$), and onset age (both $P < 0.001$). The AUC for serum elastase 1 levels (0.918 ± 0.039) was significantly greater than AUCs for serum lipase levels ($P = 0.041$), BMI ($P = 0.006$), and onset age ($P < 0.001$). There was no significant difference among AUCs for Σ C-peptide values, fasting C-peptide levels, HbA_{1c} levels, duration of hyperglycemic symptoms, and serum elastase 1 levels.

Table 1 indicated discriminating criteria devised from the optimum cutoff values based on ROC curve analysis of clinical and laboratory findings at onset of diabetes and sensitivities, specificities, positive predictive values, and negative predictive values for these criteria. Sensitivities and specificities of the discriminating criteria of Σ C-peptide values and fasting C-peptide levels were $>90\%$. Positive predictive values of the discriminating criteria of Σ C-peptide values and fasting C-peptide levels were both 80% , whereas those of the discriminating criteria of the other variables were $<70\%$.

Multiple logistic regression analysis. Based on a multiple logistic regression analysis, Σ C-peptide values at onset ≤ 0.540 nmol/l and HbA_{1c} levels at onset $\leq 8.0\%$ were recognized as independent variables for discriminating fulminant type 1 diabetes from acute-onset type 1 diabetes; the odds ratio of Σ C-peptide was 16.2 (95% CI 4.0–65.1; $P < 0.0001$), and the odds ratio of HbA_{1c} was 11.5 (2.8–47.4; $P = 0.0007$). Using the criterion where patients whose Σ C-peptide values at onset were ≤ 0.540 nmol/l and HbA_{1c} levels at onset were $\leq 8.0\%$ were diagnosed as fulminant type 1 diabetes, sensitivity and specificity were 92.0% (23/25 [95% CI 74.0–99.0]) and 99.0% (99/100 [94.6–100.0]), respectively, and positive and negative predictive values were 95.8% (23/24 [78.9–

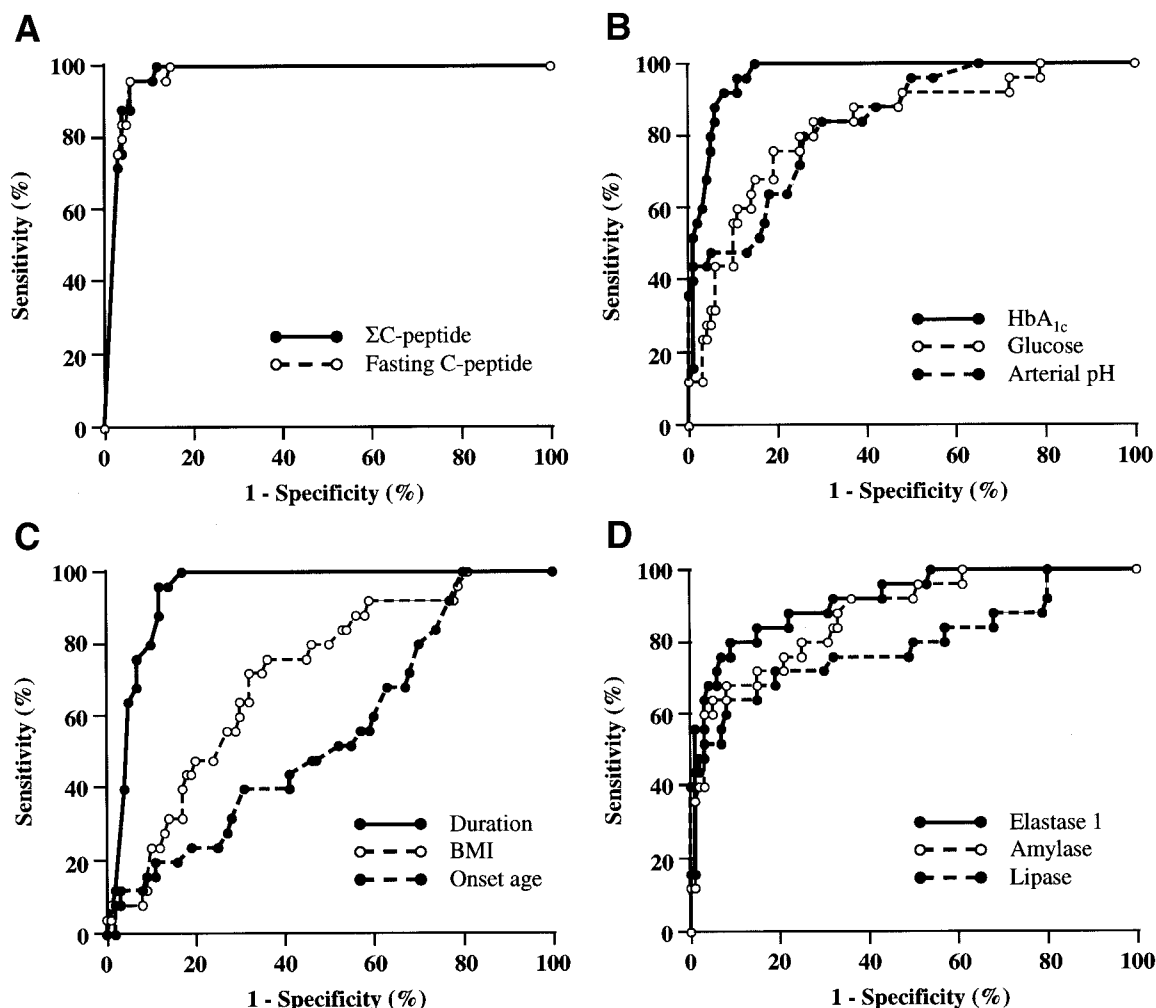


Figure 2—ROC curves for clinical and laboratory findings at the onset of diabetes, discriminating between fulminant type 1 diabetes and acute-onset type 1 diabetes. A: ROC curves for Σ C-peptide values and fasting serum C-peptide levels. B: ROC curves for HbA_{1c} levels, plasma glucose levels, and arterial pH. C: ROC curves for duration of hyperglycemic symptoms before diagnosis, BMI, and onset age of diabetes. D: ROC curves for serum elastase 1 levels, serum amylase levels, and serum lipase levels.

99.9]) and 98.0% (99/101 [93.0–99.7]), respectively.

To assess which variables are significant independent ones in the absence of Σ C-peptide values, which were sometimes hard to obtain in clinical practice, a further multiple logistic regression analysis was done after excluding Σ C-peptide values. In this case, fasting C-peptide levels at onset ≤ 0.033 nmol/l and HbA_{1c} levels at onset $\leq 8.0\%$ were recognized as independent variables; the odds ratio of fasting C-peptide was 11.4 (95% CI 3.4–38.4; $P = 0.0001$), and the odds ratio of HbA_{1c} was 7.7 (2.2–26.7; $P = 0.0013$). Using the criterion where patients whose fasting C-peptide levels at onset were ≤ 0.033 nmol/l and HbA_{1c} levels $\leq 8.0\%$ were diagnosed as fulminant type 1 dia-

betes, sensitivity, specificity, positive predictive value, and negative predictive value were 92.0% (23/25 [95% CI 74.0–99.0]), 98.0% (98/100 [93.0–99.8]), 92.0% (23/25 [74.0–99.0]), and 98.0% (98/100 [93.0–99.8]), respectively.

CONCLUSIONS— In previous studies (3,5), the diagnosis of fulminant type 1 diabetes was based on negative findings of type 1 diabetes–related autoantibodies, including ICA, GADAb, IA-2Ab, and IAA, and aggressive mode of onset. However, type 1 diabetes–related autoantibodies in classical acute-onset type 1 diabetes are sometimes negative even at onset of diabetes (24–26). Very few hospital laboratories can assay diabetes-related autoantibodies, including ICA, GADAb,

IA-2Ab, and IAA. Measurement of C-peptide levels can be easily carried out during routine practice. In the present study, we clearly demonstrate that the measurement of serum C-peptide values at onset is highly effective for diagnosis of fulminant type 1 diabetes without measuring pancreatic autoantibodies. The serum C-peptide values at onset were also highly predictive for further change of serum C-peptide levels. Low serum C-peptide levels at onset (fasting C-peptide ≤ 0.033 nmol/l or Σ C-peptide ≤ 0.540 nmol/l) can discriminate fulminant type 1 diabetes from acute-onset type 1 diabetes with high sensitivity and specificity (Table 1). Because acute-onset type 1 diabetes is much more common than fulminant type 1 diabetes, the possibility cannot be

Table 1—Discriminating criteria of fulminant type 1 diabetes from acute-onset type 1 diabetes and the sensitivity, specificity, positive predictive value, and negative predictive value for each discriminating criterion

Discriminating criterion*	Sensitivity	Specificity	Positive predictive value	Negative predictive value
Σ C-peptide ≤ 0.540 nmol/l	96.0 (79.7–99.9)	94.0 (87.4–97.8)	80.0 (61.4–92.3)	98.9 (94.3–100.0)
Fasting serum C-peptide ≤ 0.033 nmol/l	96.0 (79.7–99.9)	94.0 (87.4–97.8)	80.0 (61.4–92.3)	98.9 (94.3–100.0)
Age at onset >20 years	100.0 (86.2–100.0)	20.0 (12.7–29.2)	23.8 (16.0–33.1)	100.0 (83.2–100.0)
BMI >19.1 kg/m ²	76.0 (54.9–90.6)	64.0 (53.8–73.4)	34.5 (22.2–48.6)	91.4 (82.3–96.8)
Duration ≤ 8.0 days	96.0 (79.7–99.9)	88.0 (80.0–93.6)	66.7 (49.0–81.4)	98.9 (93.9–100.0)
Glucose >33.6 mmol/l	76.0 (54.9–90.6)	81.0 (71.9–88.2)	50.0 (33.9–66.6)	93.1 (85.6–97.4)
HbA _{1c} $\leq 8.0\%$	96.0 (79.7–99.9)	89.0 (81.2–94.4)	68.6 (50.7–83.2)	98.9 (94.0–100.0)
Arterial pH ≤ 7.21	84.0 (63.9–95.4)	74.0 (64.3–82.3)	44.7 (30.2–59.9)	94.9 (87.4–98.6)
Amylase >345 IU/l	68.0 (46.5–85.1)	92.0 (84.8–96.5)	68.0 (46.5–85.1)	92.0 (84.8–96.5)
Lipase >173 U/l	64.0 (42.5–82.0)	92.0 (84.8–96.5)	66.7 (44.7–84.4)	91.1 (83.8–95.8)
Elastase one >231 ng/dl	80.0 (59.3–93.2)	91.0 (83.6–95.8)	69.0 (49.2–84.7)	94.8 (88.3–98.3)

Data are percent (95% CI). *Discriminating criterion of fulminant type 1 diabetes ($n = 25$) from acute-onset type 1 diabetes ($n = 100$) was devised from the optimum cutoff value obtained using ROC curve analysis of each clinical or laboratory finding at onset of diabetes. Normal range of amylase, 111–336; normal range of lipase, 25–170; normal range of elastase 1, 22–221.

ruled out that our criteria based only on serum C-peptide give a fair number of false positives in acute-onset type 1 diabetes. Recently, Maldonado et al. (27) reported an association of ketosis-prone and ICA-negative diabetes with diminished fasting C-peptide levels at mean values of 0.08 nmol/l in the U.S. population. This value is similar to the cutoff value of fulminant type 1 diabetes in our study (Table 1).

Our study, using multiple logistic regression analysis, demonstrates that the diagnostic criteria consisting of the combination of serum C-peptide levels at onset and HbA_{1c} levels at onset (a criterion where the level of fasting C-peptide is ≤ 0.033 nmol/l and HbA_{1c} $\leq 8.0\%$ or where the level of Σ C-peptide is ≤ 0.540 nmol/l and HbA_{1c} $\leq 8.0\%$) increased positive predictive values without changing other properties of the criteria, including sensitivities, specificities, and negative predictive values, when compared with those of criteria consisting only of serum C-peptide levels. Therefore, criteria taking into account both serum C-peptide levels at onset and HbA_{1c} levels at onset improve the diagnostic accuracy of fulminant type 1 diabetes without the measurement of islet cell autoantibodies.

We have prospectively demonstrated that patients with fulminant type 1 diabetes had extremely low C-peptide response during OGTT at onset, at 1 year after onset, and at 2 years after onset when compared with patients with acute-onset type 1 diabetes. Characteristically, none of patients with fulminant type 1 diabetes had

improvement of Σ C-peptide values, i.e., >0.331 -nmol/l (1.0 ng/ml) increase of Σ C-peptide values after onset (1 year and 2 years) when compared with the values at onset. In contrast, in more than one-third of acute-onset type 1 diabetic patients, improvement of Σ C-peptide values as demonstrated by OGTT at 1 year after onset was recognized. These findings are in line with previous reports that a high C-peptide level at diagnosis has been associated with a higher C-peptide level during the first year of follow-up (28,29). No recovery of residual β -cell function after onset is another characteristic feature of fulminant type 1 diabetes. An inverse correlation has been reported between residual β -cell function and the degree of glycemic instability (30). We previously demonstrated that a negative relationship between preserved β -cell function and progression of diabetes complications exists (12). This indicates that patients with fulminant type 1 diabetes have higher risks of developing diabetes complications than patients with acute-onset type 1 diabetes. Therefore, fulminant type 1 diabetic patients might have a need for accurate diagnosis followed by intensive insulin therapy to attain good and stable glycemic control.

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References

1. Nakanishi K, Kobayashi T, Sugimoto T, Itoh T, Murase T, Kosaka K: Does pancreatic involvement occur in IDDM? *Diabetes Care* 11:100–101, 1988
2. Kobayashi T: Immunology and immunogenetics of type 1 diabetes in Japan. *IDF Bull* 35:34–37, 1990
3. Imagawa A, Hanafusa T, Miyagawa J, Matsuzawa Y: A novel subtype of type 1 diabetes characterized by a rapid onset and an absence of diabetes-related antibodies. *N Engl J Med* 342:301–307, 2000
4. Tanaka S, Kobayashi T, Momotsu T: A novel subtype of type 1 diabetes mellitus. *N Engl J Med* 342:1835–1837, 2000
5. Tanaka S, Kobayashi T, Nakanishi K, Koyama R, Okubo M, Murase T, Odawara M, Inoko H: Association of HLA-DQ genotype in autoantibody-negative and rapid-onset type 1 diabetes mellitus. *Diabetes Care* 25:2302–2307, 2002
6. Imagawa A, Hanafusa T, Uchigata Y, Kanatsuka A, Kawasaki E, Kobayashi T, Shimada A, Shimizu I, Toyoda T, Maruyama T, Makino H: Fulminant type 1 diabetes: a nationwide survey in Japan. *Diabetes Care* 26:2345–2352, 2003
7. Bingley PJ, Bonifacio E, Mueller PW: Diabetes Antibody Standardization Program: first assay proficiency evaluation. *Diabetes* 52:1128–1136, 2003
8. Kobayashi T, Sugimoto T, Itoh T, Kosaka K, Tanaka T, Suwa S, Satoh K, Tsuji K: The prevalence of islet cell antibodies in Japanese insulin-dependent and non-insulin-dependent diabetic patients studied by indirect immunofluorescence and by a new method. *Diabetes* 35:335–340, 1986
9. Nakanishi K, Kobayashi T, Miyashita H,

- Ohkubo M, Sugimoto T, Murase T, Kosaka K, Inouye K, Kono M: Relationships among islet cell antibodies, residual β -cell function, and metabolic control in patients with insulin-dependent diabetes mellitus of long duration: use of sensitive C-peptide radioimmunoassay. *Metabolism* 39:925–930, 1990
10. Nakanishi K, Kobayashi T, Murase T, Nakatsuji T, Inoko H, Tsuji K, Kosaka K: Association of HLA-A24 with complete β -cell destruction in IDDM. *Diabetes* 42: 1086–1093, 1993
 11. Kajio H, Kobayashi T, Nakanishi K, Okubo M, Tsukada T, Nakayama T, Yamada N, Murase T, Yazaki Y, Kosaka K: Relationship between insulin-dependent diabetes mellitus (IDDM) and non-insulin-dependent diabetes mellitus: β -cell function, islet cell antibody, and haptoglobin in parents of IDDM patients. *Metabolism* 44:869–875, 1995
 12. Nakanishi K, Kobayashi T, Inoko H, Tsuji K, Murase T, Kosaka K: Residual β -cell function and HLA-A24 in IDDM: markers of glycemic control and subsequent development of diabetic retinopathy. *Diabetes* 44:1334–1339, 1995
 13. Nakanishi K, Kobayashi T, Murase T, Naruse T, Nose Y, Inoko H: Human leukocyte antigen-A24 and -DQA*0301 in Japanese insulin-dependent diabetes mellitus: independent contributions to susceptibility to the disease and additive contributions to acceleration of β -cell destruction. *J Clin Endocrinol Metab* 84: 3721–3725, 1999
 14. The Expert Committee on the Diagnosis and Classification of Diabetes Mellitus: Report of the Expert Committee on the Diagnosis and Classification of Diabetes Mellitus. *Diabetes Care* 20:1183–1197, 1997
 15. Hoelzel W, Weykamp C, Jeppsson JO, Miedema K, Barr JR, Goodall I, Hoshino T, John WG, Kobold U, Little R, Mosca A, Mauri P, Paroni R, Susanto F, Takei I, Thienpont L, Umemoto M, Wiedmeyer HM, IFCC Working Group on HbA_{1c} Standardization: IFCC reference system for measurement of hemoglobin A_{1c} in human blood and the national standardization schemes in the United States, Japan, and Sweden: a method-comparison study. *Clin Chem* 50:166–174, 2004
 16. Kobayashi T, Nakanishi K, Murase T, Kosaka K: Small dose of subcutaneous insulin as a strategy for preventing slowly progressive β -cell failure in islet cell antibody-positive patients with clinical features of NIDDM. *Diabetes* 45:622–626, 1996
 17. Masuda M, Powell M, Chen S, Beer C, Fichna P, Rees Smith B, Furmaniak J: Autoantibodies to IA-2 in insulin-dependent diabetes mellitus: measurements with a new immunoprecipitation assay. *Clin Chim Acta* 291:53–66, 2000
 18. Maruyama T, Kasuga A, Ozawa Y, Nagata A, Abiko F, Suzuki Y, Saruta T: Glutamic acid decarboxylase65 (GAD65) antibodies and insulin auto-antibodies in Japanese patients with non-insulin-dependent diabetes mellitus. *Endocr J* 44:43–51, 1997
 19. Kobayashi T, Nakanishi K, Nakase H, Kajio H, Okubo M, Murase T, Kosaka K: In situ characterization of islets in diabetes with a mitochondrial DNA mutation at nucleotide position 3243. *Diabetes* 46: 1567–1571, 1997
 20. Tanaka S, Kobayashi T, Tomura H, Okubo M, Nakanishi K, Takeda J, Murase T: A novel dominant-negative mutation of the hepatocyte nuclear factor-1 gene in Japanese early-onset type 2 diabetes. *Horm Metab Res* 32:373–377, 2000
 21. Bristow AF, Das RE: WHO international reference reagents for human proinsulin and human insulin C-peptide. *J Biol Stand* 16:179–186, 1988
 22. Zweig MH, Campbell G: Receiver-operating characteristic (ROC) plots: a fundamental evaluation tool in clinical medicine. *Clin Chem* 39:561–577, 1993
 23. Hanley JA, McNeil BJ: A method of comparing the area under receiver operating characteristic curves derived from the same cases. *Radiology* 148:839–843, 1983
 24. Landin-Olsson M, Arnqvist HJ, Blohme G, Littorin B, Lithner F, Nystrom L, Schersten B, Sundkvist G, Wibell L, Ostman J, Lernmark A: Appearance of islet cell autoantibodies after clinical diagnosis of diabetes mellitus. *Autoimmunity* 29:57–63, 1999
 25. Decochez K, Tits J, Coolens JL, Van Gaal L, Krzentowski G, Winnock F, Anckaert E, Weets I, Pipeleers DG, Gorus FK: High frequency of persisting or increasing islet-specific autoantibody levels after diagnosis of type 1 diabetes presenting before 40 years of age: the Belgian Diabetes Registry. *Diabetes Care* 23:838–844, 2000
 26. Borg H, Gottsater A, Fernlund P, Sundkvist G: A 12-year prospective study of the relationship between islet antibodies and β -cell function at and after the diagnosis in patients with adult-onset diabetes. *Diabetes* 51:1754–1762, 2002
 27. Maldonado M, Hampe CS, Gaur LK, D'Amico S, Iyer D, Hammerle LP, Bolgiano D, Rodriguez L, Rajan A, Lernmark A, Balasubramanyam A: Ketosis-prone diabetes: dissection of a heterogeneous syndrome using an immunogenetic and β -cell functional classification, prospective analysis, and clinical outcomes. *J Clin Endocrinol Metab* 88: 5090–5098, 2003
 28. Marner B, Agner T, Binder C, Lernmark A, Nerup J, Mandrup-Poulsen T, Walldorff S: Increased reduction in fasting C-peptide is associated with islet cell antibodies in type 1 (insulin-dependent) diabetic patients. *Diabetologia* 28:875–880, 1985
 29. Martin S, Pawlowski B, Greulich B, Ziegler AG, Mandrup-Poulsen T, Mahon J: Natural course of remission in IDDM during first year after diagnosis. *Diabetes Care* 15:66–74, 1992
 30. Fukuda M, Tanaka A, Tahara Y, Ikegami H, Yamamoto Y, Kumahara Y, Shima K: Correlation between minimal secretory capacity of pancreatic β -cells and stability of diabetic control. *Diabetes* 37:81–88, 1988