

Differences in prognostic factors and outcomes in African Americans and whites with acute myeloid leukemia

Mikkael A. Sekeres, Bercedis Peterson, Richard K. Dodge, Robert J. Mayer, Joseph O. Moore, Edward J. Lee, Jonathan Kolitz, Maria R. Baer, Charles A. Schiffer, Andrew J. Carroll, James W. Vardiman, Frederick R. Davey, Clara D. Bloomfield, Richard A. Larson, and Richard M. Stone, for the Cancer and Leukemia Group B (CALGB)

Whites have a more favorable prognosis than African Americans for a number of cancers. The relationship between race and outcome is less clear in acute myeloid leukemia (AML). Using data from 7 Cancer and Leukemia Group B studies initiated from 1985 to 1997, we conducted a retrospective cross-sectional analysis of 2570 patients (270 African American and 2300 white) with de novo AML who received induction chemotherapy. African Americans were younger than whites

(48 versus 54 years, $P < .001$). African Americans also had different cytogenetic risk group distributions than whites ($P < .001$): they were more commonly classified in the favorable (23% versus 14%) and unfavorable (31% versus 23%) groups, and less commonly classified in the intermediate group (47% versus 63%). African American men had a lower complete remission (CR) rate (54%, compared with 64% for white men, 65% for white women, and 70% for African American

women, $P = .001$) and a worse overall survival compared with all other patients ($P = .004$), when known risk factors are taken into account. African Americans and whites with AML differ with respect to important prognostic factors. African American men have worse CR rates and overall survival than whites and African American women, and should be considered a poor-risk group. (Blood. 2004;103:4036-4042)

© 2004 by The American Society of Hematology

Introduction

Acute myeloid leukemia (AML) is a malignant disorder arising through the acquisition of genetic mutations in hematopoietic stem or progenitor cells,² resulting in impairment of hematopoiesis and unrestrained proliferation of an immature clone. In the United States, the annual population incidence of AML in whites is 3.7 cases per 100 000 people, whereas in African Americans it is 2.9 cases per 100 000 people.³

For a number of cancers, particularly cancers of the colon, breast, and prostate, whites have a more favorable prognosis than African Americans in the United States.⁴⁻¹¹ The relationship between race and outcome is less clear in leukemia. African American children with acute lymphoblastic leukemia (ALL) are less likely to achieve remission and have an inferior disease-free and overall survival compared with white children with the same disease.^{12,13} One recent study from the Children's Cancer Group examined racial and ethnic differences in children with ALL and found that black children were overrepresented in high-risk characteristic groups and had a worse outcome compared with white children.¹⁴ Although data concerning incidence and outcome in AML

according to race or ethnic group are sparse, Latinos appear to have a higher prevalence of one of the better-risk subtypes, acute promyelocytic leukemia (APL) (French-American-British [FAB] classification M3).¹⁵⁻¹⁷ It is unclear whether racial differences in subtypes and outcome in leukemia result from environmental or cultural influences,¹⁷ differences in genetics, or variability in reporting at diagnosis and/or follow-up.¹⁴

Surveillance, Epidemiology, and End Result (SEER) Program data indicate that the age-adjusted mortality rate for AML is slightly higher in whites compared with African Americans (2.5 versus 2.0 per 100 000 US population).³ The age adjustment is important, as whites are diagnosed at a median of 67 years, while the median for African Americans is 60 years. It is not known whether AML-specific mortality differences between the 2 races exist nationwide.¹⁸ In addition, race-specific death rates do not control for other prognostic factors, including cytogenetics and AML subtype.

Disparate cancer outcomes between races often are attributed to differential access to care, varying treatment aggressiveness and

From the Department of Hematology and Medical Oncology, The Cleveland Clinic Foundation, Cleveland, OH; The Cancer and Leukemia Group B Statistical Center, Durham, NC; Department of Adult Oncology, The Dana-Farber Cancer Institute, Harvard Medical School, Boston, MA; Duke University Medical Center, Durham, NC; Sinai Hospital of Baltimore, Baltimore, MD; North Shore University Hospital, New York University School of Medicine, Manhasset, NY; Roswell Park Cancer Institute, Buffalo, NY; Karmanos Cancer Institute, Wayne State University, Detroit, MI; University of Alabama at Birmingham, Birmingham, AL; University of Chicago, Chicago, IL; SUNY Upstate Medical University, Syracuse, NY; and The Ohio State University, Columbus, OH.

Submitted September 23, 2003; accepted February 8, 2004. Prepublished online as *Blood* First Edition Paper, February 19, 2004; DOI 10.1182/blood-2003-09-3118.

A complete list of the members of the Cancer and Leukemia Group B appears in the "Appendix."

The research for CALGB 10401 was supported, in part, by grants from the National Cancer Institute (CA3946) to the Cancer and Leukemia Group B. The

CALGB Statistical Office, Durham, NC, is supported by CA33601. Support for cytogenetics data and for C.D.B. comes from National Institutes of Health grants CA16058, CA101140, CA31946, and CA77658, and from the Coleman Leukemia Research Fund.

Richard K. Dodge died on August 24, 2002.

Presented in part at the 44th annual meeting of the American Society of Hematology, Philadelphia, PA, December 9, 2002.¹

Reprints: Mikkael A. Sekeres, Cleveland Clinic Lerner College of Medicine of Case Western Reserve University, Department of Hematology and Medical Oncology, The Cleveland Clinic Foundation, Desk R35, 9500 Euclid Ave, Cleveland, OH 44195; e-mail: sekerem@ccf.org.

The publication costs of this article were defrayed in part by page charge payment. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 U.S.C. section 1734.

© 2004 by The American Society of Hematology

compliance, and biologic variability.^{5,6,8,11,13,19-21} Even if differential access to care and treatment aggressiveness are eliminated as potential confounders in describing racial differences in outcome for AML (as should occur within a cooperative group study), biologic variability may still play a role.^{22,23}

Using data from Cancer and Leukemia Group B (CALGB) AML studies initiated from 1985 to 1997, we compared baseline prognostic factors, treatment-related complications, and outcome in African American and white patients receiving similar therapy for AML.

Patients, materials, and methods

Study population

We conducted a retrospective cross-sectional study of 2570 patients with de novo AML who were eligible to receive intensive chemotherapy as part of 1 of 7 CALGB studies: CALGB 8525, 8923, 9022, 9222, 9420, and 9621, and the initial 120 patients of CALGB 9720, the only study that allowed patients with prior myelodysplastic syndrome (MDS) or patients who had received prior cytotoxic chemotherapy to be included.²⁴⁻³¹ Patients identified their own race, and those who were classified as being neither white nor African American (221 patients) were excluded from our analyses. Patients with APL were included in CALGB 8525, 9022, 9222, and the majority of the enrollment period for 8923, but excluded from the other, more recent studies.

Treatment and clinical data

Patients were treated with the use of protocol guidelines for study entry, treatment compliance, and therapy. Induction therapy included cytarabine (Ara-C) and daunorubicin; patients in some studies also received etoposide (VP-16) as a third agent. Postremission therapy varied by protocol (Table 1). Clinical data collected at the CALGB Statistical Center (Durham, NC) on most patients included age; sex; CALGB performance status; evidence of extramedullary leukemia; baseline complete blood counts; FAB type; immunophenotype (available on 554 patients [22%]); number of induction courses; response to induction; complete remission (CR) rates (which followed accepted criteria)³²; disease-free survival (DFS) (defined as the interval from attainment of a CR or randomization to postremission therapy to the date of relapse, the date of death from any cause, or the date the patient was last known to be in remission); and overall survival (OS), which included patients who did not attain a CR. Median follow-up time for patients was 7.3 years (interquartile range, 3.0-10.4 years).

Cytogenetic data

Centrally reviewed cytogenetic data were available on 1580 patients (61%) in the study sample who were enrolled on CALGB 8461, a prospective cytogenetics study initiated in 1984.³³ Specimens were obtained at diagnosis from all patients and were processed with the use of unstimulated short-term cultures, a direct method, or both. G-banding was usually done, although Q-banding was also acceptable. At least 20 marrow metaphase cells were analyzed in patients designated as having a normal karyotype.

Cytogenetic risk groups were classified by the schema developed by CALGB for overall survival,³³ as follows: favorable risk cytogenetics included t(8;21), inv(16), or t(16;16); intermediate risk cytogenetics included a normal karyotype, del(9q), -Y, del(5q), loss of 7q, t(9;11), +11, del(11q), abn(12p), +13, del(20q), and +21. Unfavorable cytogenetics included complex (3 or more abnormalities) karyotypes, inv(3) or t(3;3), t(6;9), t(6;11), -7, +8, and t(11;19)(q23;p13.1). Of the 1580 patients, 89 individuals with t(15;17) (indicative of APL) or t(9;22) (suggestive of the blastic phase of chronic myelogenous leukemia) were analyzed separately, as these were thought to represent distinct AML subtypes. An additional 108 patients with karyotypes that included single nonrecurring structural abnormalities, or cytogenetic abnormalities that were found in fewer than 5 patients, were also excluded from the cytogenetic risk stratification, leaving 1383 patients stratified into 1 of the 3 cytogenetic risk groups.

Immunophenotyping was performed by multiparameter flow cytometry, as previously described.³⁴

Statistical analysis

Descriptive statistics, including proportions, medians, and 5-year time-to-event probabilities, were used to summarize the distribution of variables within subgroups. The Fisher exact test was used to describe the associations in 2 × 2 and 2 × 3 contingency tables. Kaplan-Meier plots³⁵ and the log-rank test were used to examine difference in overall survival and DFS. The proportional hazards model was used to model both OS and DFS as a function of demographic, clinical, and cytogenetic risk group (ie, the 3-level variable described in Table 4). Likewise, the logistic regression model was used to model CR rate as a function of this same set of variables.

Our major interest was race differences in prognostic factors and clinical outcomes; this focus was narrowed when large clinical differences between African American men and others emerged. The covariates used in the tests of African American male effects included age, M3 status, white blood cell (WBC) count, and cytogenetic risk group. As slightly more than 1000 patients did not have cytogenetic information, we first controlled only for age, M3 status, and WBC count. Then cytogenetic risk group was added to the model, and we observed if there was any change in the race or African American male regression coefficient. As the addition of cytogenetic risk group had minimal impact on the regression coefficient, or the regression coefficient became even larger, covariate-adjusted results are reported for the control of only age, M3, and WBC count. Controlling for individual cytogenetic abnormalities had no impact on the association between race/sex and the clinical endpoints. A 2-sided alpha level of 0.05 was used for all tests; as a large number of tests were performed, those of borderline significance should be interpreted with caution. Statistical analyses were performed at the CALGB Statistical Center.

Results

Clinical and biologic characteristics at study entry

Of the 2570 patients included in our analysis, 270 (10.5%) were African American and 2300 (89.5%) were white. The distribution of patients by study is summarized in Table 1. There were no significant differences in the percentage of African American men and women represented in each study.

Overall, baseline characteristics were similar in the 2 groups (Table 2). However, African Americans were younger, with a median age of 48 years compared with 54 years for whites ($P < .001$), and were more likely to be female (58% versus 46%, $P < .001$). African Americans also had more leukemic involvement of the central nervous system (CNS) at diagnosis (2% versus 1%, $P = .04$), but were less likely to have skin involvement (4% versus 8%, $P = .02$) than whites.

The FAB classification at study entry also differed between races ($P = .04$). African Americans were more likely than whites to have a diagnosis of M3 (acute promyelocytic) leukemia (11% versus 6%, $P = .003$), but less likely to have a diagnosis of M5 (monocytic) leukemia, with a trend toward significance (7% versus 11%, $P = .06$). These differences persisted when age was controlled for.

Flow cytometry data were available for 62 African Americans (23%) and 492 whites (21%). Human leukocyte antigen DR (HLA-DR) was expressed on myeloblasts from African Americans less frequently than on blasts from whites (73% versus 85%, $P = .02$), as would be expected from the FAB classification data. Other cell surface antigen expression was similar in the 2 groups.

Baseline cytogenetic data were available for 168 African Americans (62%) and 1412 whites (61%) (Table 3). African Americans were more likely than whites to have a t(15;17), the

Table 1. Summary of included trials

	Treatment protocol						
	8525 ^a	8923 ^b	9022 ^c	9222 ^d	9420 ^e	9621 ^f	9720 ^g
Years of patient accrual	10/85-10/90	2/90-11/93	10/90-3/92	7/92-12/95	1/95-7/97	2/97-3/00	1/98-3/99
Trial phase	3	3	2	3	1	1/2	3
Patients^h							
Total	1036	360	213	407	103	340	111
White, no. (%)	920 (89)	334 (93)	192 (90)	354 (87)	94 (91)	301 (89)	105 (95)
African American, no. (%)	116 (11)	26 (7)	21 (10)	53 (13)	9 (9)	39 (11)	6 (5)
Cytogenetics available, no. (%)	563 (54)	210 (58)	124 (58)	269 (66)	62 (60)	270 (79)	82 (74)
Induction 1 ⁱ	Yes	Yes ⁱ	Yes	Yes	Yes ⁱ	Yes ⁱ	Yes ⁱ
Etoposide used in inductions 1 (and 2, if needed)	No	No	No	No	Yes: 60 or 100 mg/m ² /d × 3 days	Yes: 60-150 mg/m ² /d × 3 days	Yes: 60 or 100 mg/m ² /d × 3 days
Induction 2, if needed	Yes	Yes	Yes	Yes	Yes ^k	Yes ^k	Yes ^k
Intensification or consolidation therapy randomized, by arm	Yes, arms A, B, C	Yes, arms A, B	No	Yes, arms A, B	No	Yes, risk-adapted, arms A, B	No
Intensification or consolidation therapy 1, by arm	A: SDAC; B: IDAC; C: HDAC	A: SDAC; B: M + IDAC	HDAC	A: HDAC; B: HDAC	5 + 2 + 2	A: HDAC; B: HDAC + E	5 + 2 + 2
Intensification or consolidation therapy 2, by arm	A: SDAC; B: IDAC; C: HDAC	A: SDAC; B: M + IDAC	E + C	A: HDAC; B: E + C	—	A: HDAC; B: SCT	—
Intensification or consolidation therapy 3, by arm	A: SDAC; B: IDAC; C: HDAC	A: SDAC; B: None	M + D	A: HDAC; B: M + D	—	A: HDAC; B: None	—
Intensification or consolidation therapy 4, by arm	A: SDAC; B: IDAC; C: HDAC	A: SDAC; B: None	—	—	—	—	—
Maintenance therapy	Yes	No	No	No	Yes	Yes	Yes

SDAC indicates 100 mg/m² cytarabine by ci every day × 5 days; IDAC, 400 mg/m² cytarabine by ci every day × 5 days; HDAC, 3 g/m² cytarabine by intravenous bolus (ivb) over 3 hours every 12 hours on days 1, 3 and 5; HDAC + E, 2 mg/m² cytarabine by ivb over 2 hours every 12 hours on days 1-4 and 40 mg/kg etoposide by ci every day for 4 days; M + IDAC, 5 mg/m² mitoxantrone every 12 hours and 500 mg/m² cytarabine every 12 hours for 6 doses each; E + C, 1800 mg/m² etoposide by ci day 1 plus 50 mg/kg cyclophosphamide ivb on days 2 and 3; M + D, 12 mg/m² mitoxantrone ivb plus 24 mg/m² diaziquone by ci days 1, 2, and 3 plus 5 μg/kg filgrastim subcutaneously days 4 through 28; 5 + 2 + 2, 100 mg/m² cytarabine by continuous infusion every day × 5 days plus 2 days of daunorubicin and etoposide at doses equivalent to those received in induction 1, ± PSC 833. Maintenance therapy for study 8525 consisted of 45 mg/m² daunorubicin ivb on day 1 plus 100 mg/m² cytarabine subcutaneously every 12 hours on days 1 through 5 repeated monthly × 4; for studies 9420, 9621, and 9720, maintenance therapy consisted of interleukin-2 as specified by Farag et al³⁰ and Baer et al.³¹ ivp indicates intravenous push; SCT, stem cell transplantation; and —, not applicable.

^aMayer et al.²⁴

^bStone et al.^{25,36}

^cMoore et al.²⁶

^dMoore et al.²⁷

^eLee et al.²⁸

^fKolitz et al.²⁹

^gBaer et al.³¹

^hPatient population data are as follows: total, N = 2570; white, n = 2300 (89.5% of total); African American, n = 270 (10.5% of total); and cytogenetics available, n = 1580 (61% of total).

ⁱ50% of enrolled patients received granulocyte-macrophage colony-stimulating factor (GM-CSF) during induction.

^jPatients in 9420 received daunorubicin at doses ranging from 30 to 60 mg/m²/d × 3 days and also received the multidrug resistance modulator PSC 833 at a loading dose of 1.5 mg/kg and a continuous infusion of 10 mg/kg/d for 3 days; patients in 9621 received daunorubicin at doses ranging from 40 to 90 mg/m²/d × 3 days depending on their randomization to PSC 833 at a loading dose of 2.8 mg/kg and a continuous infusion of 10 mg/kg/d for 3 days; patients in 9720 received daunorubicin at doses of 40 or 60 mg/m²/d × 3 days depending on their randomization to PSC 833 at a loading dose of 2.8 mg/kg and a continuous infusion of 10 mg/kg/d for 3 days.

^kDoses were equivalent to those received in induction 1. Patients receiving PSC 833 for induction 1 received it for induction 2 over 2 days.

^lInduction 1 consisted of daunorubicin 45 mg/m² ivp × 3 days and cytarabine 100 or 200 mg/m² by continuous infusion (ci) × 7 days. Induction 2, which was administered if needed, consisted of daunorubicin 45 mg/m² ivp × 2 days and cytarabine 100 or 200 mg/m² by ci × 5 days.

translocation associated with APL (8.3% versus 4.7%, respectively, $P = .06$) and a t(9;22) (1.8% versus 0.4%, $P = .06$), though numbers were small. There were 1383 patients (141 African Americans and 1242 whites) who were classifiable into favorable, intermediate, or unfavorable cytogenetic risk score groups. African Americans had significantly different cytogenetic risk group distributions than whites ($P < .001$): they were more commonly classified in the favorable (23% versus 14%) and unfavorable (31% versus 23%) risk groups, and less commonly classified in the intermediate group (47% versus 63%).

Table 4 shows cytogenetic abnormalities for the 1383 patients classified into 1 of the 3 risk groups. African Americans were more likely than whites to have the favorable t(8;21) (17.0% versus 5.8%, $P < .0001$). Whites, on the other hand, were more likely to have a normal karyotype (51.9% versus 37.6%, $P = .0014$). African Americans were more likely to have t(6;9) (2.1% versus 0.6%, $P = .07$) and t(6;11) (2.8% versus

0.6%, $P = .02$), both unfavorable cytogenetic abnormalities. There was no difference in the distribution of cytogenetic abnormalities between African American men and women.

Clinical outcome

African American and white AML patients received a similar number of courses of remission induction therapy (Table 5) and had similar treatment-related mortality rates (18% for whites and 16% for African Americans), which did not differ by sex. Induction toxicity did not differ in the 2 races. Whites were more likely to have grade 3 or 4 anemia, however ($P = .03$).

African American men had a significantly lower CR rate than all other patients when age, M3, and WBC count are controlled for ($P = .001$). African American men had a CR rate of 54% ($n = 114$) as compared with the much higher CR rates of 64% ($n = 1233$) for white men, 65% ($n = 1067$) for white women, and 70% ($n = 156$)

Table 2. Baseline characteristics

Characteristic	Whites, N = 2300	African Americans, N = 270	P
Median age, y	54	48	< .001
Minimum	16	17	
Maximum	86	86	
Sex, n (%)			< .001
Men	1233 (54)	114 (42)	
Women	1067 (46)	156 (58)	
Performance status, n (%)			.21
0	673 (30)	67 (25)	
1	1015 (45)	117 (44)	
2	431 (19)	59 (22)	
3	111 (5)	16 (6)	
4	28 (1)	6 (2)	
Extramedullary leukemia, n (%)			
Liver			.91
No	2041 (90)	239 (91)	
Yes	215 (10)	24 (9)	
Spleen			.73
No	2051 (91)	244 (92)	
Yes	203 (9)	22 (8)	
CNS			.04
No	2236 (99)	261 (98)	
Yes	14 (≤ 1)	5 (2)	
PNS			.72
No	2255 (99)	267 (> 99)	
Yes	14 (≤ 1)	1 (< 1)	
Nodes			.44
No	1968 (87)	226 (85)	
Yes	291 (13)	39 (15)	
Skin			.02
No	2086 (92)	258 (96)	
Yes	170 (8)	10 (4)	
Gum			.76
No	2002 (89)	235 (88)	
Yes	255 (11)	32 (12)	
Mass			.73
No	2237 (99)	265 (99)	
Yes	20 (1)	3 (1)	
Median WBC count, × 10⁹/L	13 700	16 300	.39
Minimum	100	300	
Maximum	560 000	301 000	
Median platelet level, × 10⁹/L	54 000	57 000	.52
Minimum	4000	2000	
Maximum	1 200 000	426 000	
Median Hgb level, g/L	92	90	.59
Minimum	28	35	
Maximum	149	150	
Histology, FAB, n (%)			.04
M0	49 (2)	9 (3)	
M1	497 (22)	52 (19)	
M2	678 (29)	84 (31)	
M3	144 (6)	31 (11)	
M4	539 (23)	61 (23)	
M5	251 (11)	19 (7)	
M6	70 (3)	6 (2)	
M7	13 (1)	1 (< 1)	
Other	24 (1)	3 (1)	
Missing	35 (2)	4 (1)	

CNS indicates central nervous system; PNS, peripheral nervous system; and Hgb, hemoglobin.

for African American women. Differences persisted when cytogenetic risk group is controlled for.

While there was a significant difference between the races in OS when age, M3, and WBC count are controlled for ($P = .009$), this

Table 3. Cytogenetic characteristics among 1580 patients with adequate karyotypes

Cytogenetic classification	Whites, n (%)	African Americans, n (%)	Total, n (%)
t(15;17)*	66 (4.7)	14 (8.3)	80 (5.1)
t(9;22)*	6 (0.4)	3 (1.8)	9 (0.6)
Not included in risk group	98 (7)	10 (6)	108 (6.8)
Included in risk group	1242 (88.0)	141 (83.9)	1383 (87.5)
Total	1412 (100)	168 (100)	1580 (100)
Risk group†			
Favorable	173 (14)	32 (23)	205 (15)
Intermediate	786 (63)	66 (47)	852 (62)
Unfavorable	283 (23)	43 (31)	326 (24)
Total	1242 (100)	141 (100)	1383 (100)

* $P = .06$.

† $P = < .001$. Two-degrees-of-freedom test of difference between races in risk group distribution.

difference was due almost entirely to the poorer survival of African American men compared with all other patients ($P = .004$). Figure 1 shows Kaplan-Meier curves demonstrating survival by race and sex, and predicted survival curves after age, M3, and WBC count are controlled for. There was also a small, but significant difference between the races in DFS ($P = .02$). Controlling for cytogenetic risk group had negligible impact on OS or DFS.

Survival within subgroups favored whites. Within FAB subgroups, this survival difference was statistically significant only within the M4 subtype of AML, for which African Americans had a median survival of 0.8 years, compared with the 1.3 year median survival for whites ($P = .004$). Within cytogenetic risk groups, African Americans with favorable cytogenetics had a median

Table 4. Percentage of whites and African Americans with each cytogenetic abnormality

Risk group	Whites, n (%)	African Americans, n (%)	Total, n (%)	P
Favorable				
t(8;21)	72 (5.8)	24 (17.0)	96 (7.0)	< .0001
inv(16) or t(16;16)	101 (8.1)	8 (5.7)	109 (7.9)	.40
Intermediate				
Normal	644 (51.9)	53 (37.5)	697 (50.6)	.0014
del(9q)	34 (2.7)	5 (3.5)	39 (2.8)	.59
t(9;11)	31 (2.5)	3 (2.1)	34 (2.5)	1.0
del(5q)	51 (4.1)	8 (5.7)	59 (4.3)	.38
loss of 7q	23 (1.9)	5 (3.5)	28 (2.0)	.20
+11	25 (2.0)	3 (2.1)	28 (2.0)	.76
del(11q)	9 (0.7)	1 (0.7)	10 (0.7)	1.0
abn(12p)	38 (3.1)	6 (4.3)	44 (3.2)	.44
+13	32 (2.6)	5 (3.5)	37 (2.7)	.42
del(20q)	14 (1.1)	2 (1.4)	16 (1.2)	.67
+21	36 (2.9)	1 (0.7)	37 (2.7)	.17
-Y	44 (3.5)	7 (5.0)	51 (3.7)	.35
Unfavorable				
Complex, 3 or more abn	131 (10.5)	18 (12.8)	149 (10.8)	.30
inv(3) or t(3;3)	18 (1.5)	1 (0.7)	19 (1.4)	.71
t(6;9)	7 (0.6)	3 (2.1)	10 (0.7)	.07
t(6;11)	7 (0.6)	4 (2.8)	11 (0.8)	.02
-7	37 (3.0)	5 (3.6)	42 (3.0)	.61
+8	139 (11.2)	22 (15.6)	161 (11.7)	.13
t(11;19) (q23;p13.1)	9 (0.7)	0 (0)	9 (0.7)	.61

Number of patients tested were as follows: whites, $n = 1242$; African Americans, $n = 141$; total, $n = 1383$. Abnormalities are ordered in this table according to the way in which they were used to form cytogenetic risk groups. Percentages do not sum up to 100% because patients could have more than one abnormality.

Table 5. Overall clinical outcome

Characteristic	Whites			African Americans			P unadjusted (adjusted)*	
	Men	Women	Total	Men	Women	Total	Comparing races†	For AA men‡
Induction courses, n (%)								
1	867 (70)	788 (74)	1655 (72)	87 (76)	111 (71)	198 (73)	.67	.34
2	366 (30)	279 (26)	645 (28)	27 (24)	45 (29)	72 (27)	(.35)	(.24)
Response to induction, n (%)								
CR	783 (64)	698 (65)	1481 (64)	61 (54)	109 (70)	170 (63)	.64	.01
NR/death	450 (37)	369 (35)	819 (36)	53 (46)	47 (30)	100 (37)	(.10)	(.001)
Disease-free survival								
5-year probability	0.26	0.27	0.26	0.27	0.21	0.21	.09	.21
Standard error	0.016	0.107	0.123	0.058	0.041	0.033	(.02)	(.08)
Overall survival								
5-year probability	0.24	0.26	0.25	0.16	0.26	0.22	.19	.04
Standard error	0.013	0.014	0.009	0.036	0.036	0.026	(.009)	(.004)

Patient populations are as follows: white men, N = 1233; white women, N = 1067; total whites, N = 2300; African American men, N = 114; African American women, N = 156; and total African Americans, N = 270.

NR indicates no response.

*Adjusted P values control for age, M3 status, and white blood cell count.

†P values for comparison of African Americans to whites.

‡P values for comparison of African American men to others.

survival of 2.1 years, compared with 5.6 years for whites ($P = .42$). Similar patterns were found in those with intermediate (1.0 versus 1.3 years, $P = .10$) and unfavorable (0.3 versus 0.6 years, $P = .96$) risk cytogenetic cohorts. Looking at specific cytogenetic abnormalities within cytogenetic subgroups shows that African Americans with a t(15;17) lived a median of 2.1 years, compared with 2.4 years for whites ($P = .73$), while those with a t(8;21) survived a median of 3.1 years, compared with 5.1 years for whites ($P = .89$). African Americans with complex karyotypes had a dismal survival (a median of 0.2 years), but survival was also quite poor for whites (a median of 0.5 years, $P = .31$).

Discussion

Patients with AML receive similar treatment regimens consisting of an anthracycline or anthracenedione and cytarabine, regardless of race. In general, modifications to this therapy are introduced under the following circumstances: (1) patients with an advanced age or poor health or multiple comorbidities may choose nonintensive or palliative therapy; (2) patients with the FAB subtype M3 AML, associated with the t(15;17), now receive additional all-*trans* retinoic acid; or (3) in patients with a particularly poor prognosis, a hematopoietic stem cell transplantation (SCT) may be considered, age and health permitting, once remission is achieved. The results of our study indicate that African Americans, particularly African American men, are more likely than whites to fall into the latter category and that African Americans have worse overall survival than whites when the 2 groups were given similar initial therapies.

We pooled data from 7 CALGB AML treatment studies conducted over 14 years to compare African American and white patients with respect to prognostic variables and outcome. African Americans represented 10.5% of study patients, a number that approaches the proportion of African Americans 18 years or older in the United States, 11.4%, as determined by US Census Bureau data.³⁷

Prognostic variables differed in the 2 groups. The percentage of African Americans with the favorable FAB subtype M3 AML (APL) was almost twice that of whites (and correlated to the percentage with a t(15;17)). African Americans also were 3 times as likely to have the favorable-risk t(8;21). Whites, on the other hand, were more likely to have a normal karyotype (51.9% of cytogenetic findings, compared with only 37.5% of findings in African Americans). The explanation for this difference is not apparent. Patients with APL were excluded from the more contemporary studies (as the use of differentiation agents became standard practice) and may have been overrepresented in earlier studies (which excluded patients with antecedent MDS, which rarely precedes APL). Thus, the prevalence of APL in this study (6.8%) cannot be thought of as representative of the population, and rates in each race may have been skewed. It is also possible that African Americans with favorable morphology, such as the FAB subtype M3 AML, were preferentially referred to CALGB study sites for therapy. African Americans with “nonnormal” cytogenetics may have more environmental exposures that predispose them to cancer and bone marrow stem cell disorders compared with whites.^{38,39} This may account for the increased incidence of unfavorable cytogenetics in this group, which occurs in pediatric ALL.⁴⁰ There

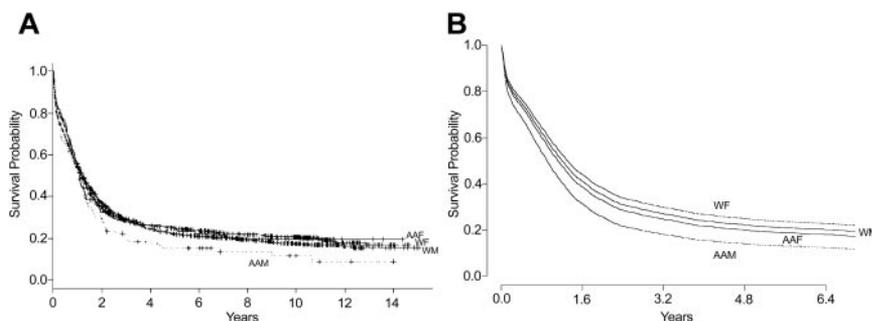


Figure 1. Survival by race and sex. (A) Unadjusted results (Kaplan-Meier plot). $P = .04$ for unadjusted comparison of survival of African American men with all other groups. (B) Adjusted results (log-rank test). $P = .004$ for comparison of survival of African American men with all other groups. AAF indicates African American women (n = 156); AAM, African American men (n = 114); WF, white women (n = 1067); and WM, white men (n = 1233).

may also be a genetic predisposition to certain favorable chromosomal rearrangements.

Perhaps more importantly, a new prognostic variable emerged. In both unadjusted and adjusted analyses, African American men had lower rates of complete remission and overall survival than any other group. This difference cannot be explained by other prognostic factors, which were controlled for; by treatment-related toxicities, which were similar in both ethnic groups; or by differences in therapies, as all patients were treated on CALGB trials and received a similar number of remission induction courses. While it is possible that African American men did not receive similar postremission therapy (eg, they may not have undergone an SCT owing to lack of a donor, as unrelated donors are less common in this population),^{41,42} their CR rates also were significantly lower than those in any other group, which would predict for a worse survival regardless of postremission therapy. Information regarding SCT following CR1 or in first relapse was not routinely collected. However, the number of patients who underwent an SCT in CR1 was small, as few patients were removed from these treatment trials to undergo an SCT, and this procedure would have been highly unlikely in studies focusing on AML in older adults (CALGB studies 8923, 9420, and 9720). Moreover, where such SCT information was collected, we would be concerned about systematic information bias. This finding of a worse outcome in African American men with AML is similar to what has been found in studies of men with colon cancer^{43,44} and may be due to biologic heterogeneity and/or differential drug metabolism and variable efficacy, as would occur with myeloblast expression of *MDR1* or an *FLT3* internal tandem duplication, and merits further study.

This study has several potential limitations. It is a cooperative group study involving patients who qualified for clinical trials, who were on average younger than the median age at AML diagnosis in the United States, and were less likely to have antecedent MDS or to have therapy-related AML. Thus, these results may not be generalizable to the entire population of patients with AML. One key advantage of this design is the assurance that virtually every patient received identical initial therapy; thus, the study controls for therapy intensity as a potential confounder. In addition, we did not have data regarding socioeconomic status, and thus could not evaluate this known risk factor for poor outcome in patients with cancer.^{5,45} One future direction would involve performing a population-based study, enabling prospective comparisons of patients of different races, complete follow-up, and testing the generalizability of information about patients enrolled in cooperative group studies. This would allow enhanced examination of racial differences in older patients, and in patients with antecedent stem cell disorders or treatment-related AML.

In conclusion, African Americans and whites with AML differ with respect to some important prognostic factors. African American men have worse CR rates and overall survival than whites and African American women. These findings support more research concerning the specific biologic features as well as the development of novel therapeutic approaches in African American men, who should be considered a poor risk group. In addition, it should be recognized that African Americans are more likely than whites to present with good-risk AML (including APL or AML with a t(8;21)), and appropriate therapy should be initiated early. Future population-based studies should try to define these racial differences further by prospectively focusing on differential environmental exposure rates; differences in drug metabolism and bioavailability; and rates of therapy compliance, toxicities, and access to remission induction and postremission therapy, including SCT.

Acknowledgments

The authors thank the patients, investigators, nurses, and data managers who participated in these clinical trials. The authors also wish to thank Brian J. Bolwell, MD, for critical review of the manuscript, and A. David McCollum, MD, for assistance with study design. The contents of this manuscript are solely the responsibility of the authors and do not necessarily represent the views of the National Cancer Institute.

Appendix: CALGB members

The CALGB is chaired by Richard L. Schilsky.

Institution	Principal investigator
Main member institutions	
Dana-Farber Partners Cancer Care	George Canellos
Dartmouth Medical School - Norris Cotton Cancer Center	Marc Ernstoff
Duke University Medical Center	Jeffrey Crawford
Georgetown University Medical Center, Lombardi Cancer Center	Edward P. Gelmann
Memorial Sloan-Kettering Cancer Center	Clifford A. Hudis
Mount Sinai School of Medicine	Lewis Silverman
North Shore - Long Island Jewish Health System	Daniel Budman
The Ohio State University Medical Center	Clara D. Bloomfield
Rhode Island Hospital	William Sikov
Roswell Park Cancer Institute	Ellis Levine
SUNY Upstate Medical University	Stephen L. Graziano
University of California at San Diego	Stephen Seagren
University of California at San Francisco	Alan Venook
University of Chicago Medical Center	Gini Fleming
University of Illinois at Chicago	Thomas Lad
University of Iowa Hospitals	Gerald Clamon
University of Maryland Cancer Center	Martin Edelman
University of Massachusetts Medical Center	Pankaj Bhargava
University of Minnesota	Bruce Peterson
University of Missouri/Ellis Fischel Cancer Center	Michael Perry
University of Nebraska Medical Center	Anne Kessinger
University of North Carolina at Chapel Hill	Thomas Shea
Vermont Cancer Center	Hyman Muss
Wake Forest University School of Medicine	David D. Hurd
Walter Reed Army Medical Center	Joseph J. Drabek
Washington University School of Medicine	Nancy Bartlett
Weill Medical College of Cornell University	Scott Wadler
At large institutions/CCOPs	
Cedars-Sinai Medical Center	Alan T. Lefor
Christiana Care Health Services, Inc	Stephen Grubbs
McGill University	Gerald Batist
Missouri Baptist Medical Center	Alan P. Lyss
Mount Sinai Medical Center - Miami	Rogério Lilenbaum
Northern Indiana Cancer Research CCOP	Rafat Ansari
Southern Nevada Cancer Research Foundation	John Ellerton
Southeast Cancer Control Consortium, Inc	James N. Atkins
Syracuse Hematology-Oncology Associates	Jeffrey Kirshner
University of Texas Southwestern Medical Center	Debasish Tripathy
Van Andel Research Institute	George F. Vande Woude
Western Pennsylvania Cancer Institute	Richard K. Shaddock
Baptist Cancer Institute CCOP	Lee S. Schwartzberg
Evanston Northwestern Healthcare CCOP	Gershon Y. Locker
Grand Rapids Clinical Oncology Program CCOP	Kathleen J. Yost
Greenville CCOP	Jeffrey K. Giguere
Illinois Oncology Research Association CCOP	John W. Kugler
Kansas City Community Clinical Oncology Program CCOP	Jorge C. Paradelo
Massey Cancer Center MBCCOP	John D. Roberts

References

1. Sekeres M, Dodge R, Bloomfield C, Larson R, Stone R. Racial differences in prognostic factors and outcome in acute myeloid leukemia (AML): a Cancer and Leukemia Group B (CALGB) study [abstract]. *Blood*. 2002;100:323a.
2. Lowenberg B, Downing JR, Burnett A. Acute myeloid leukemia. *N Engl J Med*. 1999;341:1051-1062.
3. Ries L, Eisner M, Kosary C, eds. SEER Cancer Statistics Review, 1973-1999. Bethesda, MD: National Cancer Institute; 2002. Available at: http://seer.cancer.gov/csr/1973_1999/. Accessed February 24, 2004.
4. Beart RW, Steele GD Jr, Menck HR, Chmiel JS, Ocwieja KE, Winchester DP. Management and survival of patients with adenocarcinoma of the colon and rectum: a national survey of the Commission on Cancer. *J Am Coll Surg*. 1995;181:225-236.
5. Bradley CJ, Given CW, Roberts C. Disparities in cancer diagnosis and survival. *Cancer*. 2001;91:178-188.
6. Marbella AM, Layde PM. Racial trends in age-specific breast cancer mortality rates in US women. *Am J Public Health*. 2001;91:118-121.
7. Hoffman RM, Gilliland FD, Eley JW, et al. Racial and ethnic differences in advanced-stage prostate cancer: the Prostate Cancer Outcomes Study. *J Natl Cancer Inst*. 2001;93:388-395.
8. Marcella S, Miller JE. Racial differences in colorectal cancer mortality: the importance of stage and socioeconomic status. *J Clin Epidemiol*. 2001;54:359-366.
9. Miller B, Kolonel L, Bernstein L, et al. Racial/ethnic patterns of cancer in the United States 1988-1992. Bethesda, MD: National Cancer Institute; 1996: 64-67. NIH Publication Number 96-4104.
10. Newman LA, Kuerer HM, Hunt KK, et al. Response to induction chemotherapy in black and white patients with locally advanced breast cancer. *Breast J*. 2000;6:242-246.
11. Roetzheim RG, Gonzalez EC, Ferrante JM, Pal N, Van Durme DJ, Krischer JP. Effects of health insurance and race on breast carcinoma treatments and outcomes. *Cancer*. 2000;89:2202-2213.
12. Pinkel D. Ethnicity and survival in children with acute lymphoid leukemia. *Leukemia*. 1993;7(suppl 2):S146-S147.
13. Pui CH, Boyett JM, Hancock ML, Pratt CB, Meyer WH, Crist WM. Outcome of treatment for childhood cancer in black as compared with white children: the St Jude Children's Research Hospital experience, 1962 through 1992. *JAMA*. 1995;273:633-637.
14. Bhatia S, Sather HN, Heerema NA, Trigg ME, Gaynon PS, Robison LL. Racial and ethnic differences in survival of children with acute lymphoblastic leukemia. *Blood*. 2002;100:1957-1964.
15. Douer D, Preston-Martin S, Chang E, Nichols PW, Watkins KJ, Levine AM. High frequency of acute promyelocytic leukemia among Latinos with acute myeloid leukemia. *Blood*. 1996;87:308-313.
16. Ruiz-Arguelles GJ. Promyelocytic leukemia in Mexican Mestizos. *Blood*. 1997;89:348-349.
17. Estey E, Thall P, Kantarjian H, Pierce S, Kornblau S, Keating M. Association between increased body mass index and a diagnosis of acute promyelocytic leukemia in patients with acute myeloid leukemia. *Leukemia*. 1997;11:1661-1664.
18. Howe H, Wingo P, Thun M, et al. Annual report to the nation on the status of cancer (1973 through 1998), featuring cancers with recent increasing trends. *J Natl Cancer Inst*. 2001;93:824-842.
19. Robbins AS, Whittemore AS, Van Den Eeden SK. Race, prostate cancer survival, and membership in a large health maintenance organization. *J Natl Cancer Inst*. 1998;90:986-990.
20. Roach M III, Cirrincione C, Budman D, et al. Race and survival from breast cancer: based on Cancer and Leukemia Group B trial 8541. *Cancer J Sci Am*. 1997;3:107-112.
21. Kalwinsky DK, Rivera G, Dahl GV, et al. Variation by race in presenting clinical and biologic features of childhood acute lymphoblastic leukaemia: implications for treatment outcome. *Leuk Res*. 1985;9:817-823.
22. Easaw SJ, Lake DE, Beer M, Seiter K, Feldman EJ, Ahmed T. Graft-versus-host disease: possible higher risk for African American patients. *Cancer*. 1996;78:1492-1497.
23. Chen CL, Liu Q, Pui CH, et al. Higher frequency of glutathione S-transferase deletions in black children with acute lymphoblastic leukemia. *Blood*. 1997;89:1701-1707.
24. Mayer RJ, Davis RB, Schiffer CA, et al. Intensive postremission chemotherapy in adults with acute myeloid leukemia. Cancer and Leukemia Group B. *N Engl J Med*. 1994;331:896-903.
25. Stone RM, Berg DT, George SL, et al. Granulocyte-macrophage colony-stimulating factor after initial chemotherapy for elderly patients with primary acute myelogenous leukemia. *Cancer and Leukemia Group B*. *N Engl J Med*. 1995;332:1671-1677.
26. Moore JO, Dodge RK, Amrein PC, et al. Granulocyte-colony stimulating factor (filgrastim) accelerates granulocyte recovery after intensive postremission chemotherapy for acute myeloid leukemia with aziridiny benzoquinone and mitoxantrone. *Cancer and Leukemia Group B study 9022*. *Blood*. 1997;89:780-788.
27. Moore J, Powell B, Velez-Garcia E, et al. A comparison of sequential non-cross resistant therapy or ARA-C consolidation following complete remission in adult patients less than 60 years with acute myeloid leukemia: CALGB 9222 [abstract]. *Proc Am Soc Clin Oncol*. 1997;16:14a.
28. Lee E, George S, Caligiuri M, et al. Parallel phase I studies of daunorubicin given with cytarabine and etoposide with or without the multidrug resistance modulator PSC-833 in previously untreated patients 60 years of age or older with acute myeloid leukemia: results of Cancer and Leukemia Group B study 9420. *J Clin Oncol*. 1999;17:2831-2839.
29. Kollitz J, George S, Dodge R, et al. Consolidation therapy by cytogenetic risk in adults with acute myeloid leukemia (AML) < 60 years in first complete remission (CR): results of CALGB 9621 [abstract]. *Blood*. 2001;98:688a.
30. Farag SS, George SL, Lee EJ, et al. Postremission therapy with low-dose interleukin 2 with or without intermediate pulse dose interleukin 2 therapy is well tolerated in elderly patients with acute myeloid leukemia: Cancer and Leukemia Group B Study 9420. *Clin Cancer Res*. 2002;8:2812-2819.
31. Baer MR, George SL, Dodge RK, et al. Phase 3 study of the multidrug resistance modulator PSC-833 in previously untreated patients 60 years of age and older with acute myeloid leukemia: Cancer and Leukemia Group B Study 9720. *Blood*. 2002;100:1224-1232.
32. Cheson BD, Cassileth PA, Head DR, et al. Report of the National Cancer Institute-sponsored workshop on definitions of diagnosis and response in acute myeloid leukemia. *J Clin Oncol*. 1990;8:813-819.
33. Byrd JC, Mrozek K, Dodge RK, et al. Pretreatment cytogenetic abnormalities are predictive of induction success, cumulative incidence of relapse and overall survival in adult patients with de novo acute myeloid leukemia: results from CALGB 8461. *Blood*. 2002;100:4325-4336.
34. Baer MR, Stewart CC, Dodge RK, et al. High frequency of immunophenotype changes in acute myeloid leukemia at relapse: implications for residual disease detection (Cancer and Leukemia Group B Study 8361). *Blood*. 2001;97:3574-3580.
35. Kaplan E, Meier P. Nonparametric estimation from incomplete observations. *J Am Stat Assoc*. 1958;43:457-481.
36. Stone RM, Berg DT, George SL, et al. Postremission therapy in older patients with de novo acute myeloid leukemia: a randomized trial comparing mitoxantrone and intermediate-dose cytarabine with standard-dose cytarabine. *Blood*. 2001;98:548-553.
37. US Bureau of the Census. Census 2000 Redistricting Data (PL 94-171) Summary File for States. Washington, DC: US Bureau of the Census; 2001: Tables PL1, PL2, PL3, and PL4.
38. Perlin SA, Wong D, Sexton K. Residential proximity to industrial sources of air pollution: interrelationships among race, poverty, and age. *J Air Waste Manag Assoc*. 2001;51:406-421.
39. Wing S, Richardson D, Wolf S, Mihal J, Crawford-Brown D, Wood J. A case control study of multiple myeloma at four nuclear facilities. *Ann Epidemiol*. 2000;10:144-153.
40. Crist WM, Carroll AJ, Shuster JJ, et al. Poor prognosis of children with pre-B acute lymphoblastic leukemia is associated with the t(1;19)(q23;p13): a Pediatric Oncology Group study. *Blood*. 1990;76:117-122.
41. Laver JH, Hulsey TC, Jones JP, Gautreaux M, Barredo JC, Abboud MR. Assessment of barriers to bone marrow donation by unrelated African American potential donors. *Biol Blood Marrow Transplant*. 2001;7:45-48.
42. Perkins HA, Hansen JA. The U.S. National Marrow Donor Program. *Am J Pediatr Hematol Oncol*. 1994;16:30-34.
43. Dignam JJ, Colangelo L, Tian W, et al. Outcomes among African Americans and Caucasians in colon cancer adjuvant therapy trials: findings from the National Surgical Adjuvant Breast and Bowel Project. *J Natl Cancer Inst*. 1999;91:1933-1940.
44. Hodgson DC, Fuchs CS, Ayanian JZ. Impact of patient and provider characteristics on the treatment and outcomes of colorectal cancer. *J Natl Cancer Inst*. 2001;93:501-515.
45. Cella DF, Orav EJ, Kornblith AB, et al. Socioeconomic status and cancer survival. *J Clin Oncol*. 1991;9:1500-1509.