

general population (SIR = 3.69, 95% CI: 1.39, 9.64; Table 1); however, because of the low number of cases (2 PV, 1 ET, and 1 myelofibrosis) such an association remains to be validated. The analyses stratified by sex (361 males/372 females), *JAK2V617F* mutational status (340 *JAK2V617F* mutated/137 wild-type), and cytotoxic therapy with hydroxyurea (59.6% of 659 evaluated) did not show any specific risk pattern (not shown in detail).

Our study is based on a smaller population compared with Frederiksen et al¹ and has the advantages of a slightly longer follow-up and of being conducted in a narrower area with an active local registry. Our carefully controlled patient series reduces the risk of possible misclassification at baseline, potentially present in a very large series obtained in a country-wide study, as mentioned by the authors.¹ In any case, we believe that the low SIR value (1.2 to 1.4) found in the Danish study,¹ together with our negative results, should call for much caution before accepting d'embledé the idea that the incidence of nonhematologic cancers is specifically increased in MPNs and, even more, before discussing such topic with the patients.

Maria Chiara Susini

Section of Hematology, University of Florence,
Florence, Italy

Giovanna Masala

Cancer Research and Prevention Institute-ISPO,
Florence, Italy

Elisabetta Antonioli

Section of Hematology, University of Florence,
Florence, Italy

Lisa Pieri

Section of Hematology, University of Florence,
Florence, Italy

Paola Guglielmelli

Section of Hematology, University of Florence,
Florence, Italy

Domenico Palli

Cancer Research and Prevention Institute-ISPO,
Florence, Italy

Alberto Bosi

Section of Hematology, University of Florence,
Florence, Italy

Alessandro M. Vannucchi

Section of Hematology, University of Florence,
Florence, Italy on behalf of the AGIMM Investigators

Acknowledgments: The study was funded by a grant from Associazione Italiana per la Ricerca sul Cancro (AIRC, Milano) "Special Program Molecular Clinical Oncology 5 × 1000" to AGIMM (AIRC-Gruppo Italiano Malattie Mieloproliferative), project number 1005. A detailed description of the AGIMM project is available at <http://www.progettoagimm.it>. The support of Fondazione A. Pofferi, Pistoia, is also acknowledged.

Contribution: M.C.S. performed research, analyzed data, and contributed to manuscript writing; G.M. analyzed data and contributed to manuscript writing; E.A., L.P., P.G., D.P., A.B. performed research; and A.M.V. designed research, analyzed data, and wrote the manuscript.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

Correspondence: Alessandro M. Vannucchi, MD, Section of Hematology, Department of Critical Care, University of Florence, Largo Brambilla 3, 50134 Florence, Italy; e-mail: amvannucchi@unifi.it.

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Response

Cancer risk in chronic myeloproliferative neoplasms

We thank Susini and colleagues for their comments¹ to our article on the risk of a new cancer in patients with chronic myeloproliferative neoplasms (CMPNs).² We have read the results from their CMPN cohort from Tuscany with interest. We agree that any novel finding must be confirmed before generally accepted and shared with patients. However, other studies have found results in line with ours. Fallah et al reported an increased risk of kidney cancer, melanoma and nonmelanoma skin cancer, as well as endocrine cancers among patients diagnosed with polycythemia vera (PV).³ Nielsen et al reported an increased risk of any cancer among *JAK2* V617F mutation-positive persons from the general population.⁴ In their study most new malignancies were CMPNs but also solid tumors were reported.⁴ Although we cannot rule out an effect of diagnostic misclassification of CMPNs in our study it is reassuring that we found similar risk of a new cancer among patients with chronic myeloid leukemia (CML) among whom diagnostic misclassification seems unlikely. We also found the expected increased risks of new hematologic malignancies. Furthermore, our results were robust across stratification according to a previous or current diagnosis of chronic obstructive pulmonary disease. The excess risk of nonhematologic cancers in our study is modest, ranging from 1.2 to 1.6, and might only be detectable in larger studies.

Henrik Frederiksen

Department of Clinical Epidemiology, Aarhus University Hospital,
and Department of Haematology, Odense University Hospital,
Odense, Denmark

Dóra Körmendiné Farkas

Department of Clinical Epidemiology, Aarhus University Hospital,
Aarhus, Denmark

Christian Fynbo Christiansen

Department of Clinical Epidemiology, Aarhus University Hospital,
Aarhus, Denmark

Hans Carl Hasselbalch

Department of Hematology, Roskilde Hospital,
Roskilde, Denmark

Henrik Toft Sørensen

Department of Clinical Epidemiology, Aarhus University Hospital,
Aarhus, Denmark

Conflict-of-interest disclosure: The authors declare no competing financial interests.

Correspondence: Henrik Frederiksen, Odense University Hospital Sdr, Boulevard 29, DK-5000 Odense C, Denmark; e-mail: hef@dadlnet.dk.

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To the editor:

Regulation of human dendritic cells by B cells depends on the signals they receive

B cells are classically known for producing antibodies. However, several reports also indicate that B cells are potent regulators of immune responses including those mediated by dendritic cells (DCs), although direct effect of B cells on DC was relatively unexplored.¹⁻⁴ Recently, Morva et al demonstrated that CD40 + TLR9 (CpG)-stimulated human B cells restrain the differentiation of monocyte-derived DCs and their maturation.⁵ They further found that stimulation of B cells is necessary to trigger potent regulatory activities on DCs.

Under physiological conditions, there is a constant cross-talk between DCs and B cells. Although suppression of DCs may play a role in preventing the autoimmunity, normal functioning of DCs without suppression is also critical for immune homeostasis and immune response to tumor cells and foreign antigens. These views indicate that B cells are not continually inhibitory on DCs. As B cells can receive activation signals via diverse receptors including B-cell receptor (BCR), CD40 and TLR, we surmised that the effect of stimulated B cells on DCs depends on the type of signals they receive. In view of the importance of BCR signaling in B-cell activation and in generation of regulatory B cells,^{1,2} we explored the role of BCR-stimulated B cells either alone or in combination with TLR9 stimulation on the differentiation and maturation of DC.

We found that when monocytes were differentiated into DCs in the presence of cytokines GM-CSF and IL-4 and B cells that were activated by combination of BCR + TLR9 (CpG) stimuli, there was a significant reduction in the expression of DC markers such as DC-SIGN, CD83, HLA-DR, CD40, CD80, CD86, and CD58 (Figure 1A). Thus, in accordance with Morva et al,⁵ our results revealed a regulatory role of BCR + CpG-stimulated B cells on differentiation of DCs. However, B cells that received signals only via BCR were not inhibitory (Figure 1A), indicating that in the absence of TLR stimuli, activated B cells do not block differentiation of DCs. In addition, we report a novel mechanism of modulation of DCs by regulatory B cells. We found that BCR + CpG-stimulated B cells induce a high percentage of apoptosis in differentiating DCs (Figure 1B). Thus, regulatory B cells can modulate DC-mediated immune responses by controlling the number of DCs. Of note, Fas-FasL-mediated apoptosis of target cells is proposed to be one of the mechanisms of immune regulation by regulatory B cells.¹

Furthermore, compared with profound inhibition of LPS-mediated maturation of DCs by CD40 + CpG-activated B cells,⁵ we found that B cells that received BCR signaling alone were only partially inhibitory on DCs (Figure 1C). Together, our results indicate that regulation of DC differentiation and maturation by B cells depends on the type of stimuli they receive. B cells receiving BCR stimulation alone are not inhibitory on

DCs while under inflammatory conditions as in TLR9 stimulation; these TLR-9-stimulated B cells can act as inflammation-limiting factors in part via inhibition of DC activation. These functional differences of B-cell stimuli were also reflected in their ability to induce the expression of key molecules CD62L and CD80/CD86 on B cells (Figure 1D) that are proposed to be important in the regulation of DC and T-cell functions, respectively, by regulatory B cells.^{5,6}

Mohan S. Maddur

Inserm Unité 872,
Centre de Recherche des Cordeliers,
Equipe 16-Immunopathology and therapeutic immunointervention,
Université Pierre et Marie Curie,
Université Paris Descartes, Unité Mixte de Recherche 872, Inserm,
Paris, France

Srini V. Kaveri

Inserm Unité 872,
Centre de Recherche des Cordeliers,
Equipe 16-Immunopathology and therapeutic immunointervention,
Université Pierre et Marie Curie,
Université Paris Descartes, Unité Mixte de Recherche 872, Inserm,
Paris, France;
International Associated Laboratory IMPACT
(Inserm France-Indian Council of Medical Research, India),
National Institute of Immunohaematology,
Mumbai, India

Jagadeesh Bayry

Inserm Unité 872,
Centre de Recherche des Cordeliers,
Equipe 16-Immunopathology and therapeutic immunointervention,
Université Pierre et Marie Curie,
Université Paris Descartes, Unité Mixte de Recherche 872, Inserm,
Paris, France;
International Associated Laboratory IMPACT
(Inserm, France-Indian Council of Medical Research, India),
National Institute of Immunohaematology,
Mumbai, India

Acknowledgments: This work was supported by Inserm, Center National de la Recherche Scientifique (CNRS), Université Pierre et Marie Curie, Université Paris Descartes and European Community's Seventh Framework Programme (FP7/2007-2013) under Grant Agreement No. 260338 ALLFUN.

Contribution: M.S.M. designed and performed the experiments, analyzed the data, and wrote the paper; S.V.K. analyzed the results and wrote the paper; and J.B. designed the experiments, analyzed the data, and wrote the paper.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

Correspondence: Jagadeesh Bayry, Inserm Unité 872, Equipe 16-Centre de Recherche des Cordeliers, 15 rue de l'Ecole de Médecine, Paris, F-75006, France; e-mail: jagadeesh.bayry@crc.jussieu.fr.