

growth- and survival signals. Indeed, recent studies with CLL and MCL have provided support for this explanation (Buchner et al¹¹; M. F. M. de Rooij, J. J. Buggy, and M.S., manuscript in preparation). In this perspective, it is tempting to speculate that combining these agents with anti-CD47 may turn out to be a highly efficacious treatment for NHL patients: once the malignant B cells have been forced out of their protective growth- and survival-supporting microenvironment into the circulation, they are more accessible and vulnerable for the action of anti-CD47, preventing their dissemination and priming the fully exposed malignant B cells for being attacked and eaten by the macrophages. This would make a promising rational combination therapy for complete eradication of lymphoma, spread or not!

Conflict-of-interest disclosure: The author declares no competing financial interests. ■

REFERENCES

- Chao MP, Tang C, Pachynski RK, Chin R, Majeti R, Weissman IL. Extra-nodal dissemination of non-Hodgkin lymphoma requires CD47 and is inhibited by anti-CD47 antibody therapy. *Blood*. 2011;118(18):4890-4901.
- Matozaki T, Murata Y, Okazawa H, Ohnishi H. Functions and molecular mechanisms of the CD47-SIRPalpha signalling pathway. *Trends Cell Biol*. 2009;19(2):72-80.
- Jaiswal S, Jamieson CH, Pang WW, et al. CD47 is up-regulated on circulating hematopoietic stem cells and leukemia cells to avoid phagocytosis. *Cell*. 2009;138(2):271-285.
- Majeti R, Chao MP, Alizadeh AA, et al. CD47 is an adverse prognostic factor and therapeutic antibody target on human acute myeloid leukemia stem cells. *Cell*. 2009;138(2):286-299.
- Chao MP, Alizadeh AA, Tang C, et al. Therapeutic antibody targeting of CD47 eliminates human acute lymphoblastic leukemia. *Cancer Res*. 2011;71(4):1374-1384.
- Chao MP, Alizadeh AA, Tang C, et al. Anti-CD47 antibody synergizes with rituximab to promote phagocytosis and eradicate non-Hodgkin lymphoma. *Cell*. 2010;142(5):699-713.
- Pals ST, de Gorter DJ, Spaargaren M. Lymphoma dissemination: the other face of lymphocyte homing. *Blood*. 2007;110(9):3102-3111.
- Chao MP, Jaiswal S, Weissman-Tsukamoto R, et al. Calreticulin is the dominant pro-phagocytic signal on multiple human cancers and is counterbalanced by CD47. *Sci Transl Med*. 2010;2(63):63ra94.
- Spaargaren M, Beuling EA, Rurup ML, et al. The B cell antigen receptor controls integrin activity through Btk and PLCgamma2. *J Exp Med*. 2003;198(10):1539-1550.
- de Gorter DJ, Beuling EA, Kersseboom R, et al. Bruton's tyrosine kinase and phospholipase Cgamma2 mediate chemokine-controlled B cell migration and homing. *Immunity*. 2007;26(1):93-104.
- Buchner M, Baer C, Prinz G, et al. Spleen tyrosine kinase inhibition prevents chemokine- and integrin-mediated stromal protective effects in chronic lymphocytic leukemia. *Blood*. 2010;115(22):4497-4506.

● ● ● PHAGOCYTES & GRANULOCYTES

Comment on McDermott et al, page 4957, and on Dale et al, page 4963

Released on a WHIM

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In this issue of *Blood*, 2 groups (McDermott et al¹ and Dale et al²) independently report the results of phase I clinical trials using the CXCR4-specific chemokine receptor antagonist plerixafor (Mozobil) to target the hyperfunctional CXCR4 signaling axis in patients with the rare immunodeficiency disease WHIM syndrome.

Warts, hypogammaglobulinemia, immunodeficiency, and myelokathexis (WHIM) syndrome is an unusual disorder whose cardinal features are severe neutropenia despite an abundance of mature neutrophils in the bone marrow (myelokathexis), lymphopenia, and susceptibility to human papillomavirus infection.³ Initial speculation regarding the mechanism of neutropenia centered on inappropriate retention of mature neutrophils⁴ versus premature apoptosis.⁵ Genetic studies have revealed that the disease is caused by terminal truncations of the chemokine receptor CXCR4 cytoplasmic domain, a domain impor-

tant for receptor down-regulation.⁶ This discovery suggested that hyperactivation of the mutant receptor was the underlying pathogenic mechanism and that inhibition of signaling might be therapeutically effective (see figure). The highly specific CXCR4 antagonist plerixafor is currently approved as a stem cell mobilizing agent. The use of this agent as treatment for WHIM syndrome required establishing the safety of CXCR4 antagonism for chronic use and testing whether partial blockade would be effective in mobilizing hematopoietic populations that carried mutant CXCR4 receptors.

In the present studies, patients with truncating mutation of CXCR4 who presented with the characteristic features of WHIM syndrome were treated with daily intramuscular injections of plerixafor over an escalating dose range. The baseline data in both study populations confirmed the profound neutropenia and lymphopenia that is characteristic of the disorder. A clinical response was observed in both cohorts, even at the lowest plerixafor dose, as assessed by serial complete blood counts, and dose responsiveness was observed up to the maximal dose used. Interestingly, lymphocyte populations showed a more robust response to treatment than neutrophils, with normalized or supranormal counts obtained even at submaximal doses. Neutrophil counts were responsive to treatment but never normalized, even at maximal plerixafor dosing. This pattern was unexpected as neutrophil mobilization in healthy subjects was greater than that of lymphocytes.⁷ Nonetheless, the neutrophil counts attained were in excess of 500 neutrophils/ μ L of blood in both studies, suggesting that therapeutic dosing was attainable within the range reported in these studies. In the study by McDermott and colleagues,¹ drug pharmacokinetic and pharmacodynamic properties were confirmed to be similar in the WHIM patients as in previously reported healthy controls. The safety profile after 1 week of use in both patient cohorts was acceptable, supporting further investigation of plerixafor as a therapeutic agent in WHIM syndrome.

In addition to the safety data presented, the results of the current clinical studies confirmed an interesting discrepancy between the relative responsiveness of lymphocyte, monocyte, and neutrophil populations in control versus WHIM syndrome subjects. In particular, B lymphocytes were highly mobilized from nearly undetectable levels to supranormal levels. It is not clear yet from which compartment lymphocytes were released by plerixafor treatment, but McDermott and colleagues speculate that the source is also the bone marrow.¹ The effect of chronic plerixafor lymphocyte mobilization on immune function in WHIM patients was not addressed in these studies, but the stage is now set for efficacy studies in which immune function can be characterized during a therapeutic trial. Clinical efficacy is likely with regard to prevention of neutropenia-related bacterial infections in light of the results obtained in both studies.

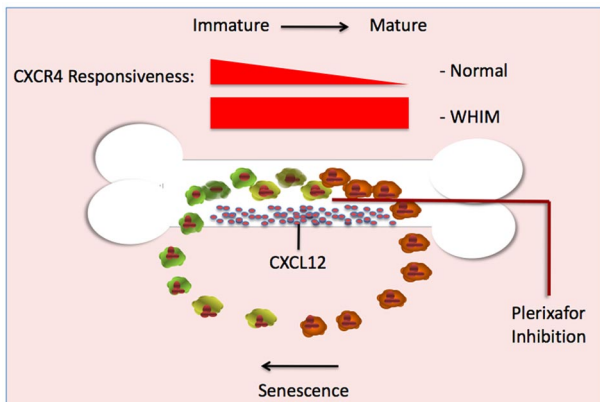


Illustration outlining the relationship between neutrophil maturation, bone marrow release, and cellular responsiveness of CXCR4 receptor to the ligand CXCL12. As neutrophils mature in healthy individuals, the cell-surface expression of CXCR4 and responsiveness to CXCL12 are reduced (darker cells). Once in the periphery, cell-surface expression and responsiveness to CXCL12 increase as these neutrophils age (lighter cells), allowing relocation to CXCL12 secreting niches. In cells expressing CXCR4 truncation mutants, down-regulation after receptor activation is impaired. As a result, the mutant receptor remains continuously responsive to CXCL12 produced by bone marrow stromal cells. Blockade of the CXCR4 receptor with plerixafor permits mutant neutrophils to escape the constant signaling loop that otherwise keeps mature neutrophils trapped in the bone marrow. This schema is presented as a probable mechanism for correction of the neutrophil trafficking defect. McDermott et al speculate that release from the bone marrow may also be the mechanism by which plerixafor corrects the lymphopenia of WHIM syndrome.¹

The data presented by these groups culminate a series of investigations into the molecular pathogenesis of WHIM syndrome. In vitro studies confirmed that not only did mutant receptors signal more robustly than wild-type receptors, they also failed to internalize normally after stimulation with the CXCR4 ligand, CXCL12 (SDF-1).⁸ Two animal models of WHIM syndrome have been described in which a recurrent 19-amino acid truncation mutant (R334X) was used to generate a murine xenotransplant model⁹ and a transgenic zebrafish model.¹⁰ These preclinical models both suggested that neutrophil retention was the basis of myelokathexis. Furthermore, transgenic expression of the mutant receptor did not accelerate apoptosis directly in the mouse model, suggesting that the apoptotic cells observed in WHIM syndrome were a secondary phenomenon of neutrophil sequestration in the bone marrow. In the zebrafish model, inhibition of the endogenous CXCL12 expression corrected the neutrophil retention, providing in vivo evidence that suppression of CXCR4-CXCL12 signaling could correct the myelokathexis phenotype. Preclinical studies with plerixafor in cell culture models expressing the R334X mutant further supported the validity of this approach.¹¹ The development of plerixafor stemmed from earlier work to develop chemokine receptor antagonists that could be useful as agents that inhibited HIV cellular entry. The approval of the agent for hematopoietic precursor cell mobilization has now fortuitously permitted the rapid development of

a molecularly targeted therapy for use in an extremely rare immunodeficiency disease.

Not all cases of myelokathexis or WHIM syndrome result from mutation of CXCR4, but increased signaling by the receptor has been implicated even in cases with distinct genetic etiologies.¹² Genetic evaluation of patients with WHIM syndrome or myelokathexis may eventually be of direct clinical use to identify those patients whose inappropriately sequestered leukocytes can be released by plerixafor therapy, potentially releasing these patients from the grasp of their disease.

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● ● ● TRANSPLANTATION

Comment on Yu et al, page 5011

Th1 and Th17 join forces for acute GVHD

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In this issue of *Blood*, Yu and colleagues demonstrate that combined targeted disruption of transcription factors T-bet and ROR γ t have defective differentiation of donor CD4⁺ T cells toward T helper 1 (Th1) and Th17, and ameliorated graft-versus-host disease (GVHD) after allogeneic hematopoietic stem cell transplantation (SCT).¹

GVHD, a major complication after allogeneic hematopoietic SCT, is mediated by donor-derived T cells. On activation with alloantigens expressed on host tissues, donor-

REFERENCES

1. McDermott DH, Liu Q, Ulrick J, et al. The CXCR4 antagonist mozobil (AMD3100) corrects pancytopenia in patients with WHIM syndrome. *Blood*. 2011;118(18):4957-4962.
2. Dale DC, Bolyard AA, Kelley ML, et al. The CXCR4 antagonist plerixafor is a potential therapy for myelokathexis, WHIM syndrome. *Blood*. 2011;118(18):4963-4966.
3. Wetzler M, Talpaz M, Kleinerman ES, et al. A new familial immunodeficiency disorder characterized by severe neutropenia, a defective marrow release mechanism, and hypogammaglobulinemia. *Am J Med*. 1990;89(5):663-672.
4. Zeulzer WW. Myelokathexis, a new form of chronic granulocytopenia. Report of a case. *N Engl J Med*. 1964;270:699-70.
5. Aprikan AA, Liles WC, Park JR, Jonas M, Chi EY, Dale DC. Myelokathexis, a congenital disorder of severe neutropenia characterized by accelerated apoptosis and defective expression of bcl-x in neutrophil precursors. *Blood*. 2000;95(1):320-327.
6. Hernandez PA, Gorlin RJ, Lukens JN, et al. Mutations in the chemokine receptor gene CXCR4 are associated with WHIM syndrome, a combined immunodeficiency disease. *Nat Genet*. 2003;34(1):70-74.
7. Liles WC, Broxmeyer HE, Rodger E, et al. Mobilization of hematopoietic progenitor cells in healthy volunteers by AMD3100, a CXCR4 antagonist. *Blood*. 2003;102(8):2728-2730.
8. Kawai T, Choi U, Whiting-Theobald NL, et al. Enhanced function with decreased internalization of carboxy-terminus truncated CXCR4 responsible for WHIM syndrome. *Exp Hematol*. 2005;33(4):460-468.
9. Kawai T, Choi U, Cardwell L, et al. WHIM syndrome myelokathexis reproduced in the NOD/SCID mouse xenotransplant model engrafted with healthy human stem cells transduced with C-terminus-truncated CXCR4. *Blood*. 2007;109(1):78-84.
10. Walters KB, Green JM, Surfus JC, Yoo SK, Huttenlocher A. Live imaging of neutrophil motility in a zebrafish model of WHIM syndrome. *Blood*. 2010;116(15):2803-2811.
11. McDermott DH, Lopez J, Deng F, et al. AMD3100 is a potent antagonist at CXCR4(R334X), a hyperfunctional mutant chemokine receptor and cause of WHIM syndrome [published online ahead of print November 10, 2010]. *J Cell Mol Med*. doi:10.1111/j.1582-4934.2010.01210.x.
12. Kawai T, Malech HL. WHIM syndrome: congenital immune deficiency disease. *Curr Opin Hematol*. 2009;16(1):20-26.