The case against ergocalciferol (vitamin D$_2$) as a vitamin supplement$^{1,2}$

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ABSTRACT

Supplemental vitamin D is available in 2 distinct forms: ergocalciferol (vitamin D$_2$) and cholecalciferol (vitamin D$_3$). Pharmacopoeias have officially regarded these 2 forms as equivalent and interchangeable, yet this presumption of equivalence is based on studies of rickets prevention in infants conducted 70 y ago. The emergence of 25-hydroxyvitamin D as a measure of vitamin D status provides an objective, quantitative measure of the biological response to vitamin D administration. As a result, vitamin D$_3$ has proven to be the more potent form of vitamin D in all primate species, including humans. Despite an emerging body of evidence suggesting several plausible explanations for the greater bioefficacy of vitamin D$_3$, the form of vitamin D used in major preparations of prescriptions in North America is vitamin D$_2$. The case that vitamin D$_2$ should no longer be considered equivalent to vitamin D$_3$ is based on differences in their efficacy at raising serum 25-hydroxyvitamin D, diminished binding of vitamin D$_2$ metabolites to vitamin D binding protein in plasma, and a nonphysiologic metabolism and shorter shelf life of vitamin D$_2$. Vitamin D$_2$, or ergocalciferol, should not be regarded as a nutrient suitable for supplementation or fortification.

INTRODUCTION

Vitamin D is available in 2 distinct forms, ergocalciferol (vitamin D$_2$) and cholecalciferol (vitamin D$_3$). These are officially regarded as equivalent and interchangeable (1–3). Although sunshine exposure and fish consumption provide vitamin D in the form of D$_3$, a different bioactive, plant-derived form of vitamin D, named vitamin D$_2$, was produced in the early 1920s through ultraviolet exposure of foods. This process was patented and licensed to pharmaceutical companies, which led to the development of a medicinal preparation of vitamin D$_2$ called Viosterol (4). Because antirachitic bioassays were used to establish “rat units” for vitamin D (ie, the amount of vitamin D required for recalcification of the epiphyseal end of tibiae in rats), early workers found it extremely difficult to distinguish between the specific biological value of the 2 forms.

To this day, the major preparations of vitamin D for prescription use in North America are in the form of vitamin D$_2$, not vitamin D$_3$. Multivitamins may contain either vitamin D$_2$ or vitamin D$_3$, but most companies are now reformulating their products to contain vitamin D in the D$_3$ form. Here, we present the case that vitamin D$_2$ should no longer be considered equivalent to vitamin D$_3$ and that vitamin D$_2$, or ergocalciferol, should not be regarded as a nutrient suitable for supplementation or fortification.

INCORRECT PRESUMPTION OF VITAMIN D$_2$ AND D$_3$ EQUIVALENCE

Assumptions about the equivalency of the 2 forms of vitamin D were questioned shortly after the discovery of vitamin D$_2$. As early as 1930, Hess et al (5) suggested that the activity of cod liver oil (vitamin D$_3$) and Viosterol (vitamin D$_2$) used in the treatment of rickets may have different biologic values. They found that one unit of cod liver oil could be as effective in preventing rickets as 4 units of Viosterol. Over the next 10 y, more than 40 studies were conducted to determine whether the 2 distinct forms of vitamin D were equally effective. The results from these studies were confusing, and, in 1940, Park (6) noted that the work done was of poor quality, making a comparison of the 2 forms exceedingly difficult. Despite these misgivings, Park stated that any effect due to differences between the 2 forms would be minimal and concluded that, “For practical purposes, the vitamin D in Viosterol (vitamin D$_2$) may be regarded as being equal to the vitamin D of cod liver oil (vitamin D$_3$)” (6). As a result, the World Health Organization recommended in 1949 that 1 IU vitamin D be equivalent to 25 ng crystalline vitamin D$_3$, and no distinction was made between vitamin D$_2$ and vitamin D$_3$ (7). Shortly thereafter in Germany in the 1950s, formulations of vitamin D$_3$ were found to be $\approx$4 times as potent per unit mass as were formulations of vitamin D$_2$. Vigantol oil (Merck KGaA, Darmstadt, Germany),

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the most popular vitamin D supplement in Europe, was reformu-
lated to replace its vitamin D<sub>2</sub> content with vitamin D<sub>3</sub> (8).

Sustained advancement in the characterization and metabol-
ism of vitamin D and its metabolites led to the proposed recom-
mendation in 1972 that 1 IU vitamin D be defined in moles or
molecules rather than in weight terms. Subsequently, both vita-
min D<sub>2</sub> and vitamin D<sub>3</sub> were defined to be 65 pmol, such that 1
IU vitamin D<sub>1</sub> (molecular weight: 384) and vitamin D<sub>2</sub> (molec-
ular weight: 396) would be equivalent to 25 ng and 25.78 ng,
respectively (9). Nevertheless, almost a half century later, British
and American pharmacopoeias continue to generalize the 2 nu-
tritional forms of vitamin D with the simple conversion of gram
quantity, where 1 IU of either vitamin D<sub>2</sub> or vitamin D<sub>3</sub> equals
25 ng (1, 3).

Despite early evidence of differences in potency between the
2 vitamin D forms on a per weight basis, it must be highlighted
that the widely practiced addition of vitamin D<sub>2</sub> to milk in the
United States and Europe in the 1930s served to successfully
eradicate rickets as a significant health problem. Additionally,
fortification of milk with either vitamin D<sub>2</sub> or vitamin D<sub>3</sub> to this
day has proven effective in the elimination of infantile rickets in
North America. To prevent infantile rickets, a minimal intake of
2.5 µg (100 IU) vitamin D/d in infants with little sun exposure
was shown to be efficacious (10). Thus, despite potential differ-
ences in the dose equivalence of vitamin D<sub>2</sub> and D<sub>3</sub>, it is likely
that vitamin D<sub>3</sub> is currently provided at a high enough dose per kg
infant body weight to maintain adequate bone mineral metabo-
lism. However, compared with the use of cruder markers (ie,
rickets or “units equivalence” of the bioassays shown by the older
rat data), the use of serum 25-hydroxyvitamin D [25(OH)D] as an
objective and quantitative marker of nutritional adequacy has
consistently shown specific differences in the biological re-
sponse of the 2 nutritional vitamin D forms.

The use of 25(OH)D as a biomarker in nonhuman species such
as birds showed vitamin D<sub>2</sub> to be only one-tenth as effective as
vitamin D<sub>3</sub> at increasing 25(OH)D (11). Likewise, in monkeys,
the concentrations of serum 25(OH)D maintained after intake of
vitamin D<sub>2</sub> were 2- to 3-fold those maintained with comparable
amounts of vitamin D<sub>3</sub> (12). In rats, however, vitamin D<sub>2</sub> was
found to be more effective (13). These differences have been
largely explained on the basis of the relative binding affinity of
vitamin D and its metabolites to the plasma vitamin D binding
protein (DBP) (14, 15). The weaker binding affinity of vitamin
D<sub>2</sub> metabolites to DBP would lead to a shorter circulating half-
life and an increased rate of clearance from circulation. Thus,
in the case of birds and monkeys, the 25(OH)D<sub>2</sub> metabolite is likely
less able to compete for binding sites on DBP. This difference in
the binding ability is potentially explained by the presence of a
methyl group at carbon 24 on the D<sub>2</sub> molecule (14).

In humans, vitamin D<sub>3</sub> is more effective than vitamin D<sub>2</sub> at
raising serum 25(OH)D concentrations. Although previous studi-
es that compared the 2 versions of vitamin D indicated a greater
effect of vitamin D<sub>3</sub> on raising 25(OH)D concentrations, evalua-
tion of potency was inconclusive due to the effects of confound-
ing variables (eg, seasonal solar exposure), insufficient sample
size, or both (16–18). In an effort to resolve the uncertainties of
earlier work, Trang et al (19) compared the ability of an equal
molar dose of vitamin D<sub>2</sub> or D<sub>3</sub> (~100 µg, or 4000 IU) to elevate
serum 25(OH)D over 2 wk between February and early May,
when vitamin D concentrations and solar exposure are minimal.

Both vitamin D<sub>2</sub> and vitamin D<sub>3</sub> increased serum 25(OH)D con-
centrations, yet the increase in 25(OH)D<sub>3</sub> was found to be 70%
greater (1.70 times) with vitamin D<sub>3</sub> than the increase obtained
with vitamin D<sub>2</sub>. When adjusted for concomitant changes in an
untreated group, the difference between the 2 groups was ~2-
fold. To further complement these findings, a 3-mo supplen-
tation study by Mastaglia et al (20) found that a dose of 250 µg
vitamin D<sub>2</sub>/d (2.5-fold) was needed to achieve similar serum
25(OH)D concentrations to those of the later study using a dose of
100 µg vitamin D<sub>3</sub>/d.

A comparison of the time course of serum 25(OH)D over a
period of 28 d after a single dose of either vitamin D<sub>2</sub> or vitamin
D<sub>3</sub> (2000 µg, or 50 000 IU, for both) was conducted by Armas et
al (21). Both forms of vitamin D produced similar rises in serum
25(OH)D concentration over the first 3 d, suggesting comparable
absorption of the 2 forms. In the vitamin D<sub>2</sub>-treated subjects,
serum 25(OH)D concentrations fell rapidly, reaching baseline
values by day 14. Interestingly, 25(OH)D concentrations then
continued to decline in this group and fell below baseline values
by day 28. In the D<sub>3</sub>-treated subjects, 25(OH)D continued to rise,
peaking by day 14 and remaining above baseline until at least
day 28. A comparison of the areas under the curve (concentration
versus time) showed a >3-fold potency with vitamin D<sub>3</sub>. Clearly,
vitamin D<sub>3</sub> would show efficacy in the treatment of severe vita-
min D deficiency; however, the authors note that 2000 µg
(50 000 IU) vitamin D<sub>2</sub> should be considered equivalent to ≤375
µg (15 000 IU) vitamin D<sub>3</sub>, and likely closer to 125 µg (5000 IU)
vitamin D<sub>3</sub> (21).

Several mechanisms could contribute to the greater capacity of
vitamin D<sub>3</sub> to maintain higher 25(OH)D concentrations over
time. Supplementation of vitamin D<sub>3</sub> produces appreciable
amounts of serum 25(OH)D<sub>3</sub> (22), which, as previously men-
tioned, has a lower affinity for DBP and results in a shorter
circulating half-life than that of 25(OH)D<sub>2</sub>. Others have sug-
gested a higher affinity of hepatic 25-hydroxylase for vitamin D<sub>3</sub>
than for vitamin D<sub>2</sub> (23). In the liver, hepatic enzyme 25-
hydroxylase places a hydroxyl group in the 25 position of the
molecule, resulting in the formation of 25(OH)D. This reaction
is the initial step in the activation of vitamin D before its metab-
olism in the kidney to its hormonally active form, 1,25(OH)<sub>2</sub>D.<npar
In rats, vitamin D 25-hydroxylase has been shown to exist in liver
mitochondrial and microsomal fractions. In humans, previous
work has shown that mitochondrial vitamin D 25-hydroxylase
presides and converts vitamin D<sub>2</sub> to 25(OH)D<sub>3</sub> 5 times as fast as it
does vitamin D<sub>3</sub> to form 25(OH)D<sub>2</sub> (23). The human micro-
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After 25-hydroxylation, 25(OH)D and 1,25(OH)2D undergo additional 24-hydroxylation in the kidney to form 24,25(OH)2D and 1,24,25(OH)3D, respectively. The formation of 1,24,25(OH)3D2 leads to deactivation of the vitamin D2 molecule, whereas the analogous vitamin D3 metabolite, 1,24,25(OH)3D3, must undergo additional side-chain oxidation to be biologically deactivated (27). In fact, 1,24,25(OH)3D3 has the ability to bind to the vitamin D receptor [VDR; \( \approx 40\% \) more than 25(OH)D3]. Thus, is able to potentially generate significant biological activity. It was suggested that this 24-hydroxylation of the side chain could occur only after 25-hydroxylation (15). Although this may be the case for vitamin D3, it does not appear to be a prerequisite for vitamin D2; evidence (28–30) suggests that 24-hydroxylation of the vitamin D2 side chain can also occur in the liver, resulting in a significant (20–50%) formation of 24(OH)D3 (29). Consequently, 1,24(OH)2D2, formed in the kidney from 24(OH)D2, has less affinity for VDR than do 1,25(OH)2D2 and 25(OH)D2 (31). Binding to VDR represents a molecular event important to the biological action of the vitamin D metabolites. Taken together, the most plausible explanations for the greater bioefficacy of vitamin D3 are conceivably due to the higher affinities of vitamin D3 and its metabolites than vitamin D2 for hepatic 25-hydroxylase, DBP, and VDR and because vitamin D3 is not directly metabolized to 24(OH)D as is vitamin D2.

**FORM OF VITAMIN D USED IN CLINICAL STUDIES**

As with all drugs that differ in molecular structure, care should be taken to distinguish the form of vitamin D used in clinical studies. For example, after vitamin D2 supplementation, Harris et al (32) reported almost double the increase in 25(OH)D concentrations in young men compared with those in older men. This result suggested a general age-related impairment in vitamin D metabolism. However, a follow-up study by the same researchers confirmed that although there are age-related changes in vitamin D metabolism, impairment with age pertains solely to vitamin D2 metabolism.

An older clinical trial in which vitamin D2 was used showed prevention loss of bone density on the basis of radiographs of the hand (34), whereas another trial showed no effect on the basis of dual-energy X-ray absorptiometry (35). One clinical trial showed that, similar to supplementation with vitamin D3 (36), supplementation with vitamin D2 caused a reduction in the rate of falls (37); however, another trial using vitamin D2 showed no effect (38). Studies using vitamin D3 have consistently shown preservation of bone density in older adults who adhered to a protocol of daily doses of \( \geq 10 \mu g \) (400 IU) (39, 40). In fact, all successful fracture trials have used vitamin D3 at doses of \( \approx 20 \mu g \) (800 IU)/d (41).

**Stability of vitamin D2 preparations**

Synthetic production of vitamin D2 is manufactured in a similar manner to that which occurs naturally in human and animal skin, via the production of 7-dehydrocholesterol from cholesterol and subsequent irradiation to its active D2 form. Conversely, vitamin D2 is synthetically produced from irradiation of ergosterol derived from the mold ergot (42). In addition to its lower bioactivity, the poor stability of vitamin D2 is worrisome, particularly upon exposure of crystalline D2 powder to varying temperatures, humidity, or even storage containers (43, 44). In contrast, vitamin D3 powder is not as labile. As a result, the vitamin D content by various manufacturers has been found to differ substantially from that of the labeled claim (17). It must be noted that comparative published data on the stability of vitamin D2 and D3 in oil is lacking. The poorer stability of and greater impurities in vitamin D2 powders may also lead to a higher risk of toxicity than that associated with the vitamin D3 metabolites. However, it is more likely that the weaker affinity of vitamin D2 metabolites to DBP produces higher and more biologically available proportions of free 25(OH)D2 and 1,25-(OH)2D2 and may thus be responsible for the greater risk of D2 toxicity (45).

**Assessment of vitamin D status after D2 supplementation: challenges to assay methodology**

The production of 25(OH)D2 as a result of vitamin D2 supplementation may additionally hinder the assessment of total circulating 25(OH)D, because common assay systems used for clinical purposes have either a diminished capacity or do not detect 25(OH)D2 with the same efficiency as 25(OH)D3 (46). Thus, clinical assays used to monitor vitamin D2 treatment may lead to an erroneous underestimation of vitamin D status. This occurrence may result in additional supplementation with potential adverse consequences, such as hypervitaminosis D.

**CONCLUSION**

Vitamin D2, if given in high enough doses, prevents infantile rickets and is capable of healing adult osteomalacia. However, the inefficiency of vitamin D2 compared with vitamin D3, on a per mole basis, at increasing 25(OH)D is now well documented, and no successful clinical trials to date have shown that vitamin D2 prevents fractures (19–21, 47). Given the assumption that the intake of any nutrient will deliver defined effects [ie, supplementation with vitamin D will lead to an increase in 25(OH)D or fracture prevention], it is clear that vitamin D2 does not fit this current nutritional notion. This is not to suggest that vitamin D2 is not efficacious, but, because the units of the 2 forms is clearly not equivalent, likely due to its distinct metabolic features and diminished binding of vitamin D2 metabolites to DBP in plasma, continual application of vitamin D2 in clinical use, including in research trials, only serves to confound our understanding of optimal vitamin D dosing recommendations. Furthermore, the public expects to derive the equivalent effect per unit dose of vitamin D, whether it is vitamin D2 or vitamin D3. The scientific community is aware that these molecules are not equivalent. Therefore, vitamin D2 should no longer be regarded as a nutrient appropriate for supplementation or fortification of foods.

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